Stem cell biology is population biology: differentiation of hematopoietic multipotent progenitors to common lymphoid and myeloid progenitors

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Additional File 1 – Supplementary Material

In this supplement, we show how the full set of cellular dynamics shown in Figure 1A in the main text can be simplified to Eqns 1-6 in the main text. Following [77-80], we describe cell dynamics using reaction kinetics, from which we derive ordinary differential equations that characterize the population dynamics of cell numbers. In [15], we gave a simplified stochastic version of the reaction kinetics that follow; we use a deterministic framework here because of the additional complexities we investigate. We first give the equations without feedback control and then explain the nature of the feedback control.

Stem Cells and Multipotent Progenitor Cells

We let \(S\) and \(MPP_j\) denote stem cell and the \(j^{th}\) \((j = 0, 1, 2, \ldots N)\) state of multipotent progenitor cells.
In the absence of feedback control, the reactions characterizing the dynamics of these cells are

\[
\begin{align*}
S & \xrightarrow{r_S} 2S \\
S & \xrightarrow{r_p} S + MPP_0 \\
S & \xrightarrow{r_p'} 2 \cdot MPP_0 \\
MPP_j & \xrightarrow{r_j} MPP_{j+1} \\
MPP_j & \xrightarrow{\lambda_j} 2 \cdot MPP_j
\end{align*}
\] (1) (2) (3) (4) (5)

We assume that MPP proliferation declines as the terminal differentiation is approached, which means that \(\lambda_j\) declines as \(j\) increases.

**Common Lymphoid Progenitors and Common Myeloid Progenitors**

After the \(N^{th}\) intermediate multipotent progenitor state, a MPP cell differentiates into a Common Lymphoid Progenitor (CLP) or Common Myeloid Progenitor (CMP) cell. Suppressing the dependence upon the concentrations of fully differentiated lymphoid and myeloid cells, we let \(0 \leq \rho \leq 1\) denote the fraction of MPP differentiations that follow the CLP route then the reaction kinetics are

\[
\begin{align*}
MPP_N & \xrightarrow{\rho_{d,MPP}} CLP \\
MPP_N & \xrightarrow{(1-\rho)\rho_{d,MPP}} CMP
\end{align*}
\] (6) (7)

**Fully Differentiated Cells**

We consider the following simplified system of fully differentiated cells: 1) fully differentiated lymphoid cells are B cells (B), T cells (T), and natural killer cells (NK); 2) fully differentiated megakaryocytes are:erythrocytes (E) and platelets (P); and 3) fully differentiated granulocytes (G) combine neutrophils, eosinophils, basophils, mast cells, and macrophages. It is clear how to remove the simplifications by expanding the granulocyte class if one wishes complexity before simplicity.
With obvious interpretation of the new parameters, we write

$$
CLP \xrightarrow{\rho_B r_{CLP}} B \quad (8)
$$

$$
CLP \xrightarrow{\rho_{NK} r_{CLP}} NK \quad (9)
$$

$$
CLP \xrightarrow{(1-\rho_B-\rho_{NK})r_{CLP}} T \quad (10)
$$

$$
CMP \xrightarrow{\rho_{ET} r_{CMP}} E \quad (11)
$$

$$
CMP \xrightarrow{\rho_{GT} r_{CMP}} G \quad (12)
$$

$$
CMP \xrightarrow{(1-\rho_E-\rho_G)r_{CMP}} P \quad (13)
$$

The total number of fully differentiated lymphoid cells, denoted by $L$, is the sum of the number of $B$, $T$, and $NK$ cells and the total number of myeloid cells, denoted by $M$ is the sum of the number of $E$, $P$, and $G$ cells. In evolutionary ecology, such sums would be called trophic species, since although there are differences in the cell (species) populations, they play the same general role in the ecosystem (organism).

**Feedback Control**

We do not explicitly model the cytokine-based feedback between the fully differentiated cells and the stem cells, nor the feedback within the niche. Instead, we follow [27] and modify the reaction rates, so that instead of being constant, they depend on the levels of fully differentiated products.

We assume that the niche can support at most $K$ stem cells and that in absence of all other feedback, the dynamics in the niche follow Gompertzian kinetics (justified in [15]). In addition to the within-niche feedback control, we let $\Phi_s(L, M)$ denote the feedback control from the fully differentiated cells [using the short-hand $L, M$ to avoid writing 6 arguments], so that $r_s$ in Eqn 1 is replaced by $r_s \cdot S \cdot \log(K/s) \cdot \Phi_s(L, M)$. Similarly $r_p$ in Eqn 2 is replaced by $r_p \cdot \Phi_s(L, M)$. We assume that there is additional feedback control on asymmetric differentiation of stem cells (Eqn 3) so that $r_p'$ is replaced by $r_p' \cdot \Phi_s(L, M) \cdot \Phi_p'(L, M)$. We assume that feedback control acts on each stage of MPP development so that each of the $\lambda_j$, $r_j$ are replaced by $\lambda_j \Phi_p(L, M)$ and $r_j \Phi_p(L, M)$.

We will describe the specific form of the feedback functions after giving the fully general reaction rate equations.
The Reaction Rate Equations

We let $\mu_i$ denote the rate of mortality of cell type $i$ (if fully developed cells also proliferate, we can capture that by setting $\mu_i < 0$) and $[\ ]$ concentration of cells. The dynamics of the full system is then

\[
\frac{d[S]}{dt} = [S] \cdot \log(K/[S])(r_s - r_p \cdot \Phi_p([L], [M])) \Phi_s([L], [M]) - \mu_s[S] \tag{14}
\]

\[
\frac{d[MPP_0]}{dt} = [S] \cdot \log(K/[S])(r_p + 2r_p \cdot \Phi_p([L], [M])) \Phi_s([L], [M]) + (\lambda_0 - r_0) \Phi_p([L], [M]) [MPP_0] - \mu_0[MPP_0] \tag{15}
\]

\[
\frac{d[MPP_j]}{dt} = r_{j-1} \Phi_p(L, M) [MPP_{j-1}]
+ (\lambda_j - r_j) \Phi_p(L, M) [MPP_j] - \mu_j[MPP_j], j = 1, 2, ..N - 1 \tag{16}
\]

\[
\frac{d[MPP_N]}{dt} = r_{N-1} \Phi_p(L, M) [MPP_{N-1}]
+ (\lambda_N - r_{d,MPP}) \Phi_p(L, M) [MPP_N] - \mu_N[MPP_N] \tag{17}
\]

\[
\frac{d[CLP]}{dt} = r_{d,MPP} \Phi_p(L, M) \rho([L], [M]) [MPP_N] - r_{CLP}[CLP] - \mu_{CLP}[CLP] \tag{18}
\]

\[
\frac{d[CMP]}{dt} = r_{d,MPP} \Phi_p(L, M)(1 - \rho([L], [M])) [MPP_N] \tag{19}
\]

\[ -r_{CMP}[CMP] - \mu_{CMP}[CMP] \]

\[
\frac{d[B]}{dt} = r_{CLP \rho_B}([B], [NK], [T])[CLP] - \mu_B[B] \tag{20}
\]

\[
\frac{d[NK]}{dt} = r_{CLP \rho_{NK}}([B], [NK], [T])[CLP] - \mu_{NK}[NK] \tag{21}
\]

\[
\frac{d[T]}{dt} = r_{CLP}(1 - \rho_B([B], [NK], [T]) - \rho_{NK}([B], [NK], [T]))[CLP] - \mu_T[T] \tag{22}
\]

\[
\frac{d[E]}{dt} = r_{CMP \rho_E}([E], [G], [P])[CMP] - \mu_E[E] \tag{23}
\]

\[
\frac{d[G]}{dt} = r_{CMP \rho_G}([E], [G], [P])[CMP] - \mu_G[G] \tag{24}
\]

\[
\frac{d[P]}{dt} = r_{CMP}(1 - \rho_E([E], [G], [P]) - \rho_G([E], [G], [P])[CMP] - \mu_P[P] \tag{25}
\]

Rapid Development and Differentiation of Progenitor Cells and Lumped Myeloid and Lymphoid Cells

If the development rate of MPP cells is fast (either because the reactions are fast or $N$ is small), we can use a quasi-steady state analysis as is commonly done for Michaelis-Menten kinetics [81]. We then write
Eqn 16 as

\begin{equation}
\frac{1}{r_j} \frac{d[MPP_j]}{dt} = \Phi_p(L, M)[MPP_{j-1}]
\end{equation}

\begin{equation}
+ \left[ \frac{\lambda_j}{r_j} - 1 \right] \Phi_p(L, M) - \frac{\mu_j}{r_j} [MPP_j]
\end{equation}

If we assume that the left-hand side of Eqn 26 and \( \frac{\mu_j}{r_j} \) are much less than 1, then Eqn 26 simplifies to

\begin{equation}
[MPP_j] = \frac{r_j-1}{r_j} (1 - \frac{\lambda_j}{r_j})^{-1} [MPP_{j-1}]
\end{equation}

Applying a similar analysis to Eqn 17 leads to

\begin{equation}
[MPP_N] = \frac{r_N-1}{r_d} (1 - \frac{\lambda_N}{r_d})^{-1} \prod_{j=1}^{N-1} \frac{r_j-1}{r_j} (1 - \frac{\lambda_j}{r_j})^{-1} [MPP_0]
\end{equation}

which we write as

\begin{equation}
[MPP_N] = \Omega_N [MPP_0]
\end{equation}

with the obvious definition of \( \Omega_N \). Eqs 18 and 19 become

\begin{equation}
\frac{d[CLP]}{dt} = \frac{r_d}{r_d} \Phi_p(L, M) \rho([L], [M]) \Omega_L [MPP_0] - \frac{r_{CLP}}{r_{CLP}} [CLP] - \mu_{CLP} [CLP]
\end{equation}

\begin{equation}
\frac{d[CMP]}{dt} = \frac{r_d}{r_d} \Phi_p(L, M) (1 - \rho([L], [M])) \Omega_L [MPP_0] - \frac{r_{CMP}}{r_{CMP}} [CMP] - \mu_{CMP} [CMP]
\end{equation}

We now denote progenitors by MPP, lump B, NK, and T cells into a lymphoid class, with concentration denoted by \([L]\) and E, G, and P cells into a myeloid class, with concentration denoted by \([M]\), equations 14-25 simplify to
\[
\frac{d[S]}{dt} = [S \cdot \log(K/[S])] (r_s - r_{p'} \Phi_{p'}([L], [M])) \Phi_s([L], [M]) - \mu_s [S] \tag{33}
\]
\[
\frac{d[MPP]}{dt} = [S \cdot \log(K/[S])] (r_p + 2 r_{p'} \Phi_{p'}([L], [M])) \Phi_s([L], [M]) \\
+ (\lambda - r_{d,MPP}) \Phi_p([L], [M]) [MPP] - \mu_p [MPP] \tag{34}
\]
\[
\frac{d[CLP]}{dt} = r_{d,MPP} \Phi_p(L, M) \rho([L], [M]) \Omega_N [MPP] \\
- r_{CLP}[CLP] - \mu_{CLP}[CLP] \tag{35}
\]
\[
\frac{d[CMP]}{dt} = r_{d,MPP} \Phi_p(L, M) (1 - \rho([L], [M])) \Omega_N [MPP] \\
- r_{CMP}[CMP] - \mu_{CMP}[CMP] \tag{36}
\]
\[
\frac{d[L]}{dt} = r_{CLP}[CLP] + (r_l - \mu_l)[L] \tag{37}
\]
\[
\frac{d[M]}{dt} = r_{CMP}[CMP] + (r_m - \mu_m)[M]. \tag{38}
\]

It is these equations that form the bases of the analysis in the main text, and complement the variety of other models of the dynamics of the HSC system (Supplementary Table 1)
**Supplementary Table S1 An Overview of Some Key Models Applied to HSCs and Their Descendants**

<table>
<thead>
<tr>
<th>Source</th>
<th>Kind of Model</th>
<th>Focus</th>
</tr>
</thead>
<tbody>
<tr>
<td>(65) Till et al 1964</td>
<td>Branching Process</td>
<td>Origins of Variability</td>
</tr>
<tr>
<td>(82) Vogel et al 1969</td>
<td>Stochastic branching processes</td>
<td>Development of erythroblastic colonies</td>
</tr>
<tr>
<td>(83) Mackey 1978</td>
<td>Nonlinear ordinary differential equations for cycling cells</td>
<td>Aplastic anemia</td>
</tr>
<tr>
<td>(76) Novak and Necas 1994</td>
<td>Steady state algebraic analysis</td>
<td>Lineage proliferation in the steady state</td>
</tr>
<tr>
<td>(84) Abkowitz et al 1996</td>
<td>Markov Birth and Death Process</td>
<td>Evidence that hematopoiesis is stochastic</td>
</tr>
<tr>
<td>(85) Haurie et al 1998</td>
<td>Delay differential equations</td>
<td>Origins of periodic hematological disorders</td>
</tr>
<tr>
<td>(29) Abkowitz et al 2000</td>
<td>Stochastic comparent model</td>
<td>Patterns of individual variation following transplants</td>
</tr>
<tr>
<td>(86) Bernard et al 2004</td>
<td>Nonlinear ordinary differential equations</td>
<td>Origins of oscillatory WBC diseases</td>
</tr>
<tr>
<td>(87) Colijn and Mackey 2005</td>
<td>Delay-differential equations</td>
<td>Oscillating leukemia</td>
</tr>
<tr>
<td>(52) Dingli and Pacheco 2006</td>
<td>Allometric scaling methods</td>
<td>Estimating the size of the active HSC pool</td>
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