www.nature.com/ejcn

# ORIGINAL ARTICLE Dietary guidance normalizes large intestinal endocrine cell densities in patients with irritable bowel syndrome

T Mazzawi<sup>1,2,3</sup>, T Hausken<sup>2,3</sup>, D Gundersen<sup>4</sup> and M El-Salhy<sup>1,2,3</sup>

**BACKGROUND/OBJECTIVES:** To determine the large intestinal endocrine cell types affected following dietary guidance in patients with irritable bowel syndrome (IBS).

**SUBJECTS/METHODS:** The study included 13 IBS patients and 13 control subjects. The patients received three sessions of individualized dietary guidance. Both the control subjects and the patients were scheduled for colonoscopies at baseline and again for the patients at 3–9 months after dietary guidance. Biopsy samples were taken from the colon and rectum and were immunostained for all types of large intestinal endocrine cells. The endocrine cells were quantified using computerized image analysis.

**RESULTS:** The daily total consumption (mean ± s.e.m. values) of fruits and vegetables rich in FODMAPs (fermentable oligosaccharides, disaccharides, monosaccharides and polyols) decreased significantly from  $16.2 \pm 5.3$  g before receiving dietary guidance to  $9.2 \pm 3.2$  g after receiving dietary guidance (P = 0.02). In the total colon, the densities of serotonin cells were  $46.8 \pm 8.9$ ,  $10.5 \pm 2.1$  and  $22.6 \pm 3.2$  cells/mm<sup>2</sup> in control subjects and in IBS patients before and after receiving dietary guidance, respectively (P = 0.007); the corresponding densities of peptide YY cells were  $11.6 \pm 1.8$ ,  $10.8 \pm 1.7$  and  $16.8 \pm 2.1$  cells/mm<sup>2</sup>, respectively (P = 0.06). The cell densities for both serotonin and peptide YY did not change significantly in the rectum. The densities of somatostatin cells in the rectum were  $13.5 \pm 3.0$ ,  $13.2 \pm 3.0$ , and  $22.3 \pm 3.2$  cells/mm<sup>2</sup> for control subjects and for IBS patients before and after receiving dietary guidance, respectively (P = 0.01).

**CONCLUSIONS:** The densities of the large intestinal endocrine cells tend to normalize following dietary guidance that may have contributed to the improvement of the patients with IBS symptoms.

European Journal of Clinical Nutrition (2016) 70, 175-181; doi:10.1038/ejcn.2015.191; published online 25 November 2015

# INTRODUCTION

Irritable bowel syndrome (IBS) is a common chronic functional gastrointestinal (GI) disorder.<sup>1</sup> Besides its associated morbidity, IBS patients exhibit a high consumption of health-care resources that represents an economic burden to society as a whole.<sup>1-4</sup> There is increasing evidence that IBS can result from dysfunction

of the GI neuroendocrine system and GI endocrine cells in particular.<sup>5–16</sup>

The GI endocrine cells are scattered among the epithelial cells lining the GI lumen.<sup>17–19</sup> These endocrine cells interact with the GI luminal contents (especially nutrients) and respond by releasing specific hormones that affect different functions of the GI tract.<sup>5,20–31</sup>

Approximately two-thirds of IBS patients relate their symptom development to diet.<sup>32,33</sup> Dietary guidance has been proven to reduce the symptoms and improve the quality of life of IBS patients.<sup>34,35</sup>

A previous study found that changing the dietary intake by the provision of individualized dietary guidance altered the large intestinal total endocrine cells as detected by a general marker for endocrine cells (chromogranin A).<sup>36</sup> The observed change in chromogranin A cell density after receiving dietary guidance was toward the normal values of controls. However, which endocrine cell type was affected is not clear.<sup>36</sup>

The present study was undertaken to determine the types of large intestinal endocrine cells that are affected after receiving dietary guidance. The study was performed on the same cohort of IBS patients in whom the total population of endocrine cells was observed after receiving dietary guidance.<sup>36</sup>

## MATERIALS AND METHODS

#### Patients and controls

Patients of both sexes aged between 18 and 70 years and referred to the Division of Gastroenterology, Stord Hospital, Norway, who fulfilled Rome-III criteria for IBS diagnosis were included in the study. The exclusion criteria were being pregnant or lactating women patients with serious psychiatric or any organic/systemic diseases, drug abuse or previous abdominal surgery, with the exception of appendectomy, caesarean section or hysterectomy. Seven patients did not use any kind of medication. The remaining patients, however, used one or a combination of the following: proton pump inhibitors (n = 4), thyroxin substitution tablets (n = 2), astma inhalators (n = 1), angiotensin II receptor antagonist antihypertensive tablets (n = 1), antiallergic tablets (n = 3), contraceptive pills (n = 2) and antidepressants/anxiolytics (n = 2). These patients were instructed not to take any kind of proton pump inhibitors 1 week before starting or during the study.

A control group of 13 subjects was included in the study. They comprised 9 females and 4 males with a mean age of 54 years (range 26–70 years). Four of the control subjects underwent colonoscopies

<sup>1</sup>Division of Gastroenterology, Department of Medicine, Stord Hospital, Stord, Norway; <sup>2</sup>Division of Gastroenterology, Department of Clinical Medicine, University of Bergen, Bergen, Norway; <sup>3</sup>National Centre for Functional Gastrointestinal Disorders, Department of Medicine, Haukeland University Hospital, Bergen, Norway and <sup>4</sup>Department of Research, Helse-Fonna, Haugesund, Norway. Correspondence: Dr T Mazzawi, Division of Gastroenterology, Department of Clinical Medicine, University of Bergen, Bergen 5021 Norway.

E-mail: tarek.mazzawi@med.uib.no

Received 2 December 2014; revised 2 September 2015; accepted 24 September 2015; published online 25 November 2015

npg

176

because of GI bleeding that identified the source as hemorrhoids (n = 3) or angiodysplasia (n = 1), whereas the other 9 control subjects underwent colonoscopies because of health worries caused by a family member being diagnosed with GI cancer.

The study was performed in accordance with the Declaration of Helsinki and was approved by the local Committee for Medical Research Ethics West, Bergen, Norway. All patients provided both oral and written consents to participate.

#### Study design

Forty-six patients (35 females and 11 males) with mean age of 35 years (range 18–69 years) were included in the study. All patients underwent physical examinations, and blood tests were performed to exclude inflammation, infection and other organic diseases. The patients were scheduled for three 45-min sessions of individualized dietary guidance, with a nurse experienced in diet and IBS, at intervals of at least 2 weeks. The patients were examined with colonoscopies before the first session and at 3–9 months (median, 4 months) following the third session of dietary guidance.

#### Individualized dietary guidance

Dietary guidance was delivered orally with the help of charts, and in written illustrations. During the first session, the patients received general information about IBS, with emphasis on the importance of a regular and healthy eating pattern and on the food items that worsen IBS symptoms such as insoluble dietary fiber and poorly absorbable FODMAPs (fermentable oligosaccharides, disaccharides, monosaccharides and polyols). The patients were informed that lactose-free milk and other lactosefree dairy products do not provoke IBS symptoms and hence they were allowed to consume such products daily during the study. The patients were supposed to test diets that were either rich or poor in protein, fat or carbohydrate, and to register for 2 weeks in a diary their daily consumption of food and fluids along with any associated symptoms, including the frequency and degree of abdominal pain and abdominal distension and the stool frequency and consistency. The consumption of food supplements containing probiotics, antibiotics and other medications was prohibited during the study unless otherwise specified.

In the second session, the nurse focused mainly on briefly repeating the information given during the first session and on using the patient's diary to identify the food items that triggered IBS symptoms. Based on the obtained information, the patients were asked to alter their diet proportions of protein, fat and carbohydrate, avoid FODMAPs-rich items as well as insoluble fiber and to consume vegetables and fruits that contained lower amounts of FODMAPs and insoluble fiber.

During the third session, each patient gave feedback about the dietary guidance to the nurse, and together with the nurse designed a suitable diet for the patient to follow until the end of the study.

#### Dietary assessment

Dietary intake was assessed using the MoBa food frequency questionnaire. This questionnaire reports the frequency and the portion sizes of food stuffs and beverages consumed over a defined period of time. Data analysis was conducted using software to calculate the nutrient content of the diet. The MoBa food frequency questionnaire was developed and validated by the Norwegian Institute of Public Health in Oslo, Norway.<sup>37,38</sup> This questionnaire inquires about the consumption of 225 foodstuffs and identifies the dietary habits of the participant, including the intake of any oral supplements, according to typical Norwegian meal patterns. The participants completed the MoBa food frequency questionnaire form before the first session and again at least 3 months after the third session of individualized dietary guidance.<sup>34</sup>

# Colonoscopies, biopsy sampling, histopathology and immunohistochemistry

Both the patients and controls were scheduled to receive colonoscopies, during which four biopsy samples were taken from each segment of the large intestine. The biopsy samples were then divided according to the segment they were taken from into right colon, left colon and rectum, where the right colon referred to the cecum, the ascending colon and the right half of the transverse colon. The left colon referred to the left half of the transverse colon, the descending colon and the sigmoid colon. In the rectum, biopsy samples were taken from the dorsal wall  $\sim\!15\,\text{cm}$  from the anus.

The biopsy samples were fixed in 4% buffered paraformaldehyde overnight, embedded in paraffin and cut into sections with a thickness of 5 µm. The biopsy samples were examined histopathologically. They were stained with hematoxylin-eosin and immunostained using the Avidin-Biotin complex (ABC) method with the Vectastain ABC kit (Vector Laboratories, Burlingame, CA, USA) and the chromogen 3,3'-diaminobenzidine peroxidase substrate (DAB) kit (Vector Laboratories). The immunostaining method has been described in detail previously.<sup>13</sup> In brief, the sections were incubated for 2 h with the primary antibodies and then for another 30 min with biotinylated swine anti-mouse IgG (in the case of monoclonal antibodies) or goat anti-rabbit IgG (in the case of polyclonal antibodies), both diluted to 1:200, and for 30 min with ABC diluted to 1:200. The slides were submerged in DAB and counterstained with hematoxylin. Incubation was carried out at room temperature. The primary antibodies used were monoclonal mouse anti-serotonin (code no. 5HT-209, Dako, Glostrup, Denmark), polyclonal anti-porcine peptide YY (PYY; code PYY 11A, Alpha Diagnostic, San Antonio, TX, USA), polyclonal rabbit antisynthetic human pancreatic polypeptide (code no. 114, Diagnostic Biosystems, Pleasanton, CA, USA), polyclonal rabbit anti-porcine oxyntomodulin (code BP508, Acris Antibodies, Herford, Germany) and polyclonal rabbit antisynthetic human somatostatin (code no. A566, Dako); these antibodies were used in dilutions of 1:1500, 1:1000, 1:800, 1:400 and 1:200, respectively.

#### Computerized image analysis

The densities of various types of endocrine cell were quantified using computer software Cell^D (Olympus, Tokyo, Japan). The number of immunoreactive endocrine cells and the area of the epithelial cells were measured in 10 randomly chosen fields using a  $\times 40$  objective. Each field represented a tissue area of 0.14 mm<sup>2</sup>. The density of endocrine cells is expressed herein as the number of cells per square mm of epithelium.

Table 1.         Demographic characteristics of participants						
IBS subtype/controls	Number	Age (mean)ª	Females/males			
IBS-D	6	30-44 (37.2)	2/4			
IBS-C	5	27-42 (32.4)	4/1			
IBS-M	2	21-23 (22)	2/0			
Controls	13	26–70 (54)	9/4			

Abbreviations: IBS, irritable bowel syndrome; IBS-C, IBS patients with constipation as a predominant symptom; IBS-D, IBS patients with diarrhea as the predominant symptom; IBS-M, IBS patients with mixed diarrhea and constipation. <sup>a</sup>Years.

**Table 2.** Densities of serotonin- and PYY-immunoreactive cells in thetotal colon, right colon and left colon of control subjects and of IBSpatients before and after receiving dietary guidance

Hormone/location	Cell density (cells/mm <sup>2</sup> )				
	Control	Before guidance	After guidance	P-value	
Serotonin Total colon	468+89	105+21	226+32	0.007 <sup>b</sup>	
Right colon	$25.9 \pm 5.4$	$1.2 \pm 0.8$	$10.7 \pm 1.6$	< 0.0001 <sup>c</sup>	
	20.9 ± 4.0	0.9 ± 1.7	10.0 ± 1.9	0.55	
Total colon	11.6± 1.8	10.8 ± 1.7	16.8 ± 2.1	0.06	
Right colon Left colon	3.9 ± 0.8 7.7 ± 1.4	2.9 ± 0.8 7.9 ± 1.0	5.5 ± 1.1 11.5 ± 1.1	0.1 0.04ª	
Left colon	$3.9 \pm 0.8$ 7.7 ± 1.4	$2.9 \pm 0.0$ $7.9 \pm 1.0$	$3.3 \pm 1.1$ $11.5 \pm 1.1$	0.1 0.04 <sup>a</sup>	

Abbreviations: IBS, irritable bowel syndrome; PYY, peptide YY. Data are presented as mean  $\pm$  s.e.m. <sup>a</sup>P < 0.05, <sup>b</sup>P < 0.01 and <sup>c</sup>P < 0.0001.

## Statistical analysis

The Kruskal–Wallis nonparametric test with Dunn's test as a post test was used to compare between the controls and patients before dietary guidance and also between controls and patients after dietary guidance. The paired *t*-test was used to compare patients before and after receiving dietary guidance. The data are presented as mean  $\pm$  s.e.m. values. *P*-values of < 0.05 were considered significant.

# RESULTS

#### Patients and controls

Forty-six patients were included in the study and received sessions of individualized dietary guidance. After the first session and colonoscopy, four patients were excluded after being diagnosed with celiac disease (n=2) and lupus (n=1) and because of



**Figure 1.** Serotonin cell densities in the total colon (**A**), right colon (**B**) and left colon (**C**) of IBS patients before and after receiving dietary guidance. The dashed lines labeled 'a' and 'c' indicate the upper and lower limits of the 95% confidence interval for control subjects, respectively, whereas line 'b' indicates the mean serotonin cell density. \*\*P < 0.01, \*\*\*\*P < 0.0001.



technically difficult colonoscopy (n = 1). The remaining patients (n = 42) went through the second session of individualized dietary guidance. Two patients were also excluded during the study because of pregnancy (n = 1) and moving abroad (n = 1). Forty patients received the third session of individualized dietary guidance. Following the third session, 24 patients withdrew their consent, either because of symptom improvement after receiving



Figure 2. Serotonin-immunoreactive cells in the total colon of a control subject (a) and of an IBS patient before (b) and after (c) receiving dietary guidance.

178

dietary guidance and/or because they were unwilling for a second colonoscopy to be performed. Additional two patients were excluded because of noncompliance (n = 2). One patient was excluded because of a bout of gastroenteritis under the study (n = 1). Thus, 13 of the original 46 patients completed the entire study, comprising 8 females and 5 males with a mean age of 34 years (range 20–45 years). The demographic characteristics are summarized in Table 1.

# Dietary assessment

The change in diet in the present study has been described in detail elsewhere.<sup>34</sup> In brief, the daily total consumption of fruits and vegetables rich in FODMAPs decreased significantly from 16.2±5.3 g before receiving dietary guidance to 9.2±3.2 g after receiving dietary guidance (P=0.02). However, the daily consumption of fiber did not differ significantly between before receiving dietary guidance (27.4±2.5 g) and after receiving dietary guidance (23.1±2.2 g, P=0.09).<sup>34</sup>

## Colonoscopies, histopathology and immunohistochemistry

The colonoscopies indicated that the colon and rectum were normal both macroscopically and microscopically in patients as well as controls, with no signs of microscopic colitis. Immunoreactive endocrine cells were found in the mucosa of both the colon and rectum of the patients and controls. These cells were either basket or flask shaped, and sometimes exhibited a long basal cytoplasmic process. The numbers of pancreatic polypeptide- and oxyntomodulin (enteroglucagon)-immunoreactive cells in the biopsy samples of the colon and rectum used in the study were too low to allow reliable quantification. There were also too few somatostatin cells in the colon to allow reliable guantification. Computerized image analyses

#### Colon

Serotonin cell density: The densities of serotonin cells in the controls and patients are listed in Table 2 and illustrated in Figures 1 and 2. The density of serotonin cells in the total colon and the right colon of IBS patients increased significantly (P = 0.007 and P < 0.0001, respectively) after receiving dietary guidance. The density of serotonin cells in the left colon also increased after receiving dietary guidance, but this increase was not statistically significant (P = 0.53). The densities of serotonin cells in controls and IBS subtypes before and after receiving guidance are listed in Table 3.

PYY cell density: The densities of PYY cells in the controls and patients are summarized in Table 2 and illustrated in Figure 3. The density of PYY cells in the left colon of IBS patients increased significantly (P=0.04) after receiving dietary guidance. The densities of PYY cells also increased in the total colon and the right colon following dietary guidance, but the change did not reach the cutoff for statistical significance (P=0.06 and P=0.1, respectively). The densities of PYY cells in controls and IBS subtypes before and after receiving guidance are listed in Table 3.

*Rectum.* The densities of serotonin, PYY and somatostatin cells in controls and patients are reported in Table 4.

Serotonin cell density: The serotonin cell density in the rectum of IBS patients did not differ significantly (P = 0.06) between before and after receiving dietary guidance (Figure 4A).

PYY cell density: The density of PYY cells in the rectum of IBS patients did not change significantly (P = 0.13) after receiving dietary guidance (Figure 4B).

Somatostatin cell density: The somatostatin cell density of IBS patients increased significantly (P = 0.01) after receiving dietary guidance (Figure 4C).

Hormone/location	Endocrine cell densities (cell/mm²)		Endocrine cell densities (cell/mm <sup>2</sup> )		P-value, before guidance vs controls	P-value, after guidance vs controls	P-value, before vs after quidance
	Control	IBS subtype	Before guidance	After guidance			2
Serotonin							
Total colon	46.8 ± 8.9	IBS-D	11.17 ± 3.4	19.83 <u>+</u> 3.7	0.009 <sup>b</sup>	0.2	0.12
		IBS-C	11.75 ± 3.8	22.75 ± 8.1	0.06	0.26	0.24
		IBS-M	$6.0 \pm 2.0$	$30.5 \pm 0.5$	0.16	0.66	0.06
Right colon	25.9 <u>+</u> 5.4	IBS-D	2.16 ± 1.6	10.3 ± 1.7	0.002 <sup>b</sup>	0.22	0.009 <sup>b</sup>
5		IBS-C	$0.75 \pm 0.48$	11.5 ± 4.7	0.03 <sup>a</sup>	0.43	0.1
		IBS-M	0	10.7 ± 2.6	0.02 <sup>a</sup>	0.42	0.05
Left colon	$20.9 \pm 4.8$	IBS-D	9.0 ± 2.9	9.5 ± 2.1	0.39	0.59	0.89
		IBS-C	$10.0 \pm 3.1$	9.0 ± 3.7	0.42	0.33	0.83
		IBS-M	$6.0\pm2.0$	$18.0\pm3.0$	0.39	0.9	0.25
ΡΥΥ							
Total colon	11.6 ± 1.8	IBS-D	$11.17 \pm 2.4$	19.17 ± 2.5	0.9	0.051	0.04 <sup>a</sup>
		IBS-C	$12.5 \pm 3.5$	11.5 ± 3.3	0.9	0.9	0.83
		IBS-M	$6.5 \pm 3.5$	$20.5 \pm 7.5$	0.43	0.37	0.42
Right colon	$3.9 \pm 0.8$	IBS-D	3.83 ± 1.3	$6.0 \pm 1.4$	0.9	0.31	0.39
-		IBS-C	3.5 ± 1.7	4.3 ± 2.5	0.9	0.9	0.78
		IBS-M	$0.33 \pm 0.33$	6.3 ± 2.4	0.09	0.8	0.15
Left colon	7.7 ± 1.4	IBS-D	7.3 ± 1.7	$13.2 \pm 1.4$	0.9	0.04 <sup>a</sup>	0.002 <sup>b</sup>
		IBS-C	9.4 ± 1.5	8.6 ± 1.6	0.95	0.9	0.72
		IBS-M	6.0 + 3.0	13.5 + 3.5	0.87	0.25	0.45

Table 3. Densities of serotonin- and PYY-immunoreactive cells in the total colon, right colon and left colon of controls and patients with different IBS

Abbreviations: IBS, irritable bowel syndrome; IBS-C, IBS patients with constipation as a predominant symptom; IBS-D, IBS patients with diarrhea as the predominant symptom; IBS-M, IBS patients with mixed diarrhea and constipation; PYY, peptide YY. Data are presented as mean  $\pm$  s.e.m. <sup>a</sup>P < 0.05 and <sup>b</sup>P < 0.01.

## DISCUSSION

The dropout rate in the present study was 52%, somewhat higher than previously reported rates (33–48%).<sup>35,39–42</sup> This higher dropout rate could be explained by the demanding design of the study, involving two colonoscopies and requiring the patients to follow a strict diet for at least 3 months. Moreover, an additional 20% of the patients were excluded from the study for other reasons (diagnosis of celiac disease, pregnancy, moving abroad, noncompliance, gastroenteritis and technical difficulties experienced during the colonoscopy). However, despite the small number of the sample who completed the study, changing the diet in these patients was shown to have a significant impact on the large intestinal endocrine cells. It is worth mentioning that neither age nor gender affects the densities of the endocrine cells in the large intestine.<sup>36,43</sup>

A previous study involving the same cohort of IBS patients investigated in the present  $study^{36}$  found that the total endocrine cells, as detected by chromogranin A, changed in the colon, but not in the rectum. The present findings



**Figure 3.** PYY cell densities in the total colon (**A**), right colon (**B**) and left colon (**C**) of IBS patients before and after receiving dietary guidance. The symbols are the same as in Figure 1. \*P < 0.05.

show that these changes in the colon are brought about by changes in serotonin and PYY cells. The elevation in the density of somatostatin cells in the rectum seen in the

Table 4. Densities of serotonin-, PYY- and somatostatin-<br/>immunoreactive cells in the rectum of control subjects and of IBS<br/>patients before and after receiving dietary guidance

Hormone	Endocrin	P-value, before vs after guidance		
	Control	Before guidance	After guidance	
Serotonin PYY Somatostatin	$\begin{array}{c} 37.7 \pm 10.5 \\ 32.5 \pm 5.1 \\ 13.5 \pm 3.0 \end{array}$	$31.9 \pm 6.4$ $38.4 \pm 5.0$ $13.2 \pm 3.0$	$\begin{array}{c} 19.6 \pm 4.5 \\ 31.5 \pm 3.7 \\ 22.3 \pm 3.2 \end{array}$	0.06 0.13 0.01 <sup>a</sup>

Abbreviations: IBS, irritable bowel syndrome; PYY, peptide YY. Data are presented as the mean  $\pm$  s.e.m.  $^aP<0.05.$ 



**Figure 4.** Cell densities of (**A**) serotonin, (**B**) PYY and (**C**) somatostatin in the rectum of IBS patients before and after receiving dietary guidance. The symbols are the same as in Figure 1. \*P < 0.05.

present study does not seem to have affected the total densities of endocrine cells.

Serotonin is known to activate the submucosal sensory branch (Meissner's plexus) of the enteric nervous system that conveys sensation from the GI tract to the central nervous system and modulates the visceral sensitivity of the GI tract.<sup>7,44–47</sup> Serotonin also stimulates the motility of the large intestine, and accelerates the transit time through both the small and large intestines.<sup>44–52</sup> The serotonin cell density has been found to be low in patients with IBS.<sup>11</sup> After receiving dietary guidance in the present study, the serotonin cell density in the colon increased significantly toward the values for the control subjects. The serotonin cell density in the rectum of IBS patients has been reported to be normal.<sup>9</sup> The serotonin cell density in the present study after receiving dietary guidance.

PYY stimulates the absorption of water and electrolytes, and is considered to be a major regulator of the 'ileal brake'.<sup>7,53</sup> The cell density of PYY has been reported to be low in the colon<sup>11</sup> and rectum<sup>9</sup> of IBS patients. The cell density of PYY increased significantly only in the left colon toward the values of control subjects, after receiving dietary guidance. The increase in the PYY cell densities in the total and right colon was not significant, and this could be a type II error because of the small sample size being studied. The cell densities of PYY did not significantly change in the rectum.

A previous study on the same cohort of IBS patients showed a significant improvement in pain (P < 0.001) and diarrhea (P < 0.05) domains but not in constipation using Birmingham IBS symptom questionnaire after receiving dietary guidance compared with before guidance.<sup>34</sup> It is possible that the significant changes in the cell densities of serotonin and PYY in IBS-D patients could be related to the observed symptomatic changes after receiving dietary guidance in the mentioned article. The different changes in the cell densities in different parts of the colon could be a type II error because of the small sample size. In the same manner, the nonsignificant changes in the cell densities in the

Somatostatin inhibits intestinal motility and exocrine and neuroendocrine secretion.<sup>8,54</sup> The rectal somatostatin cell density has been found to be higher in patients with IBS.<sup>9</sup> The somatostatin cell density in the rectum increased significantly after receiving dietary guidance. This increase in somatostatin cell density is difficult to explain. It is possible that it may be caused by a compensatory reaction to the increased cell density of PYY after receiving dietary guidance.

The changes in the endocrine cell densities after receiving dietary guidance differed between the colon and rectum, probably because of their different functions: the colon absorbs water, sodium and some fat-soluble vitamins, whereas the rectum acts only as a fecal reservoir before defecation.<sup>13</sup>

Nutrients in the lumen of the GI tract are the main triggers for the endocrine cells releasing various GI hormones that control and regulate several functions of the GI tract.<sup>8,55</sup> The stem cells in the GI tract require between 2 and 6 days to differentiate into different endocrine cells.<sup>56,57</sup> It can therefore be speculated that changing the pattern of food intake can alter the differentiation of the endocrine cells.

In conclusion, this study is the first to show that the densities of the endocrine cells in the large intestines are affected by the type of food consumed. A change in diet of IBS patients following the provision of dietary guidance can normalize the densities of these endocrine cells and recover their malfunctioning, and may have resulted in the improvement of IBS symptoms.

#### **CONFLICT OF INTEREST**

The authors declare no conflict of interest.

## ACKNOWLEDGEMENTS

We thank Professor Hans Olav Fadnes, Head of the Department of Medicine, Stord Hospital, for his support and for reading the manuscript. This study was supported by a grant from Helse-Fonna.

### REFERENCES

- 1 El-Salhy M, Gundersen D, Hatlebakk JG, Hausken T. *Irritable Bowel Syndrome*. Nova Scientific Publisher: New York, 2012.
- 2 Whitehead WE, Burnett CK, Cook EW 3rd, Taub E. Impact of irritable bowel syndrome on quality of life. *Dig Dis Sci* 1996; **41**: 2248–2253.
- 3 Talley NJ, Gabriel SE, Harmsen WS, Zinsmeister AR, Evans RW. Medical costs in community subjects with irritable bowel syndrome. *Gastroenterology* 1995; 109: 1736–1741.
- 4 Spanier JA, Howden CW, Jones MP. A systematic review of alternative therapies in the irritable bowel syndrome. *Arch Intern Med* 2003; **163**: 265–274.
- 5 El-Salhy M. Ghrelin in gastrointestinal diseases and disorders: a possible role in the pathophysiology and clinical implications (review). Int J Mol Med 2009; 24: 727–732.
- 6 El-Salhy M. Irritable bowel syndrome: diagnosis and pathogenesis. World J Gastroenterol 2012; **18**: 5151–5163.
- 7 El-Salhy M, Gilja OH, Gundersen D, Hatlebakk JG, Hausken T. Endocrine cells in the ileum of patients with irritable bowel syndrome. *World J Gastroenterol* 2014; 20: 2383–2391.
- 8 El-Salhy M, Gundersen D, Gilja OH, Hatlebakk JG, Hausken T. Is irritable bowel syndrome an organic disorder? *World J Gastroenterol* 2014; 20: 384–400.
- 9 El-Salhy M, Gundersen D, Hatlebakk JG, Gilja OH, Hausken T. Abnormal rectal endocrine cells in patients with irritable bowel syndrome. *Regul Pept* 2013; **188C**: 60–65.
- 10 El-Salhy M, Gundersen D, Hatlebakk JG, Hausken T. Low-grade inflammation in the rectum of patients with sporadic irritable bowel syndrome. *Mol Med Rep* 2013; 7: 1081–1085.
- 11 El-Salhy M, Gundersen D, Ostgaard H, Lomholt-Beck B, Hatlebakk JG, Hausken T. Low densities of serotonin and peptide YY cells in the colon of patients with irritable bowel syndrome. *Dig Dis Sci* 2012; **57**: 873–878.
- 12 El-Salhy M, Lomholt-Beck B, Hausken T. Chromogranin A as a possible tool in the diagnosis of irritable bowel syndrome. *Scand J Gastroenterol* 2010; 45: 1435–1439.
- 13 El-Salhy M, Mazzawi T, Gundersen D, Hausken T. Chromogranin A cell density in the rectum of patients with irritable bowel syndrome. *Mol Med Rep* 2012; 6: 1223–1225.
- 14 El-Salhy M, Rauma J. Low density of ghrelin cells in the oxyntic mucosa correlated to slow gastric emptying in patients with type 1 diabetes. *Mol Med Rep* 2009; 2: 893–896.
- 15 El-Salhy M, Vaali K, Dizdar V, Hausken T. Abnormal small-intestinal endocrine cells in patients with irritable bowel syndrome. *Dig Dis Sci* 2010; 55: 3508–3513.
- 16 El-Salhy M, Wendelbo IH, Gundersen D. Reduced chromogranin A cell density in the ileum of patients with irritable bowel syndrome. *Mol Med Rep* 2013; 7: 1241–1244.
- 17 Moran GW, Leslie FC, Levison SE, Worthington J, McLaughlin JT. Enteroendocrine cells: neglected players in gastrointestinal disorders? *Therap Adv Gastroenterol* 2008; 1: 51–60.
- 18 Moran-Ramos S, Tovar AR, Torres N. Diet: friend or foe of enteroendocrine cells-how it interacts with enteroendocrine cells. *Adv Nutr* 2012; **3**: 8–20.
- 19 Buffa R, Capella C, Fontana P, Usellini L, Solcia E. Types of endocrine cells in the human colon and rectum. *Cell Tissue Res* 1978; **192**: 227–240.
- 20 Sandstrom O, El-Salhy M. Ageing and endocrine cells of human duodenum. Mech Ageing Dev 1999; 108: 39–48.
- 21 Tolhurst G, Reimann F, Gribble FM. Intestinal sensing of nutrients. *Handb Exp Pharmacol* 2012309–335.
- 22 Lee J, Cummings BP, Martin E, Sharp JW, Graham JL, Stanhope KL et al. Glucose sensing by gut endocrine cells and activation of the vagal afferent pathway is impaired in a rodent model of type 2 diabetes mellitus. Am J Physiol Regul Integr Comp Physiol 2012; 302: R657–R666.
- 23 Parker HE, Reimann F, Gribble FM. Molecular mechanisms underlying nutrientstimulated incretin secretion. Expert Rev Mol Med 2010; 12: e1.
- 24 Raybould HE. Nutrient sensing in the gastrointestinal tract: possible role for nutrient transporters. *J Physiol Biochem* 2008; **64**: 349–356.

- 25 San Gabriel A, Nakamura E, Uneyama H, Torii K. Taste, visceral information and exocrine reflexes with glutamate through umami receptors. *J Med Invest* 2009; **56**: 209–217.
- 26 Rudholm T, Wallin B, Theodorsson E, Naslund E, Hellstrom PM. Release of regulatory gut peptides somatostatin, neurotensin and vasoactive intestinal peptide by acid and hyperosmolal solutions in the intestine in conscious rats. *Regul Pept* 2009; **152**: 8–12.
- 27 Sternini C, Anselmi L, Rozengurt E. Enteroendocrine cells: a site of 'taste' in gastrointestinal chemosensing. *Curr Opin Endocrinol Diabetes Obes* 2008; 15: 73–78.
- 28 Sternini C. Taste receptors in the gastrointestinal tract. IV. Functional implications of bitter taste receptors in gastrointestinal chemosensing. *Am J Physiol Gastrointest Liver Physiol* 2007; **292**: G457–G461.
- 29 Buchan AM. Nutrient tasting and signaling mechanisms in the Gut III. Endocrine cell recognition of luminal nutrients. *Am J Physiol* 1999; **277**: G1103–G1107.
- 30 Montero-Hadjadje M, Elias S, Chevalier L, Benard M, Tanguy Y, Turquier V et al. Chromogranin A promotes peptide hormone sorting to mobile granules in constitutively and regulated secreting cells: role of conserved N- and C-terminal peptides. J Biol Chem 2009; 284: 12420–12431.
- 31 Shooshtarizadeh P, Zhang D, Chich JF, Gasnier C, Schneider F, Haikel Y *et al.* The antimicrobial peptides derived from chromogranin/secretogranin family, new actors of innate immunity. *Regul Pept* 2010; **165**: 102–110.
- 32 Simren M, Mansson A, Langkilde AM, Svedlund J, Abrahamsson H, Bengtsson U *et al.* Food-related gastrointestinal symptoms in the irritable bowel syndrome. *Digestion* 2001; **63**: 108–115.
- 33 Williams EA, Nai X, Corfe BM. Dietary intakes in people with irritable bowel syndrome. BMC Gastroenterol 2011; 11: 9.
- 34 Mazzawi T, Hausken T, Gundersen D, El-Salhy M. Effects of dietary guidance on the symptoms, quality of life and habitual dietary intake of patients with irritable bowel syndrome. *Mol Med Rep* 2013; **8**: 845–852.
- 35 Ostgaard H, Hausken T, Gundersen D, El-Salhy M. Diet and effects of diet management on quality of life and symptoms in patients with irritable bowel syndrome. *Mol Med Rep* 2012; **5**: 1382–1390.
- 36 Mazzawi T, Gundersen D, Hausken T, El-Salhy M. Increased chromogranin a cell density in the large intestine of patients with irritable bowel syndrome after receiving dietary guidance. *Gastroenterol Res Pract* 2015; 2015: 823897.
- 37 Masson LF, McNeill G, Tomany JO, Simpson JA, Peace HS, Wei L et al. Statistical approaches for assessing the relative validity of a food-frequency questionnaire: use of correlation coefficients and the kappa statistic. Public Health Nutr 2003; 6: 313–321.
- 38 Brantsaeter AL, Haugen M, Alexander J, Meltzer HM. Validity of a new food frequency questionnaire for pregnant women in the Norwegian Mother and Child Cohort Study (MoBa). *Matern Child Nutr* 2008; 4: 28–43.
- 39 Enck P, Klosterhalfen S, Kruis W. Determination of placebo effect in irritable bowel syndrome. Dtsch Med Wochenschr 2005; 130: 1934–1937.
- 40 Abdul-Baki H, El H II, Elzahabi L, Azar C, Aoun E, Skoury A *et al.* A randomized controlled trial of imipramine in patients with irritable bowel syndrome. *World J Gastroenterol* 2009; **15**: 3636–3642.
- 41 Zernicke KA, Campbell TS, Blustein PK, Fung TS, Johnson JA, Bacon SL et al. Mindfulness-based stress reduction for the treatment of irritable bowel syndrome symptoms: a randomized wait-list controlled trial. Int J Behav Med 2013; 20: 385–396.

- 42 Halmos EP, Power VA, Shepherd SJ, Gibson PR, Muir JG. A diet low in FODMAPs reduces symptoms of irritable bowel syndrome. *Gastroenterology* 2014; **146**: 67–75.e5.
- 43 Sandstrom O, el-Salhy M. Human rectal endocrine cells and aging. *Mech Ageing Dev* 1999; **108**: 219–226.
- 44 Gershon MD, Tack J. The serotonin signaling system: from basic understanding to drug development for functional GI disorders. *Gastroenterology* 2007; **132**: 397–414.
- 45 Gershon MD. Plasticity in serotonin control mechanisms in the gut. *Curr Opin Pharmacol* 2003; **3**: 600–607.
- 46 Gershon MD. 5-Hydroxytryptamine (serotonin) in the gastrointestinal tract. Curr Opin Endocrinol Diabetes Obes 2013; 20: 14–21.
- 47 Gershon MD. Serotonin is a sword and a shield of the bowel: serotonin plays offense and defense. *Trans Am Clin Climatol Assoc* 2012; **123**: 268–280. discussion 80.
- 48 Tack JF, Janssens J, Vantrappen G, Wood JD. Actions of 5-hydroxytryptamine on myenteric neurons in guinea pig gastric antrum. Am J Physiol 1992; 263: G838–G846.
- 49 Michel K, Sann H, Schaaf C, Schemann M. Subpopulations of gastric myenteric neurons are differentially activated via distinct serotonin receptors: projection, neurochemical coding, and functional implications. *J Neurosci* 1997; 17: 8009–8017.
- 50 Tack J, Coulie B, Wilmer A, Andrioli A, Janssens J. Influence of sumatriptan on gastric fundus tone and on the perception of gastric distension in man. *Gut* 2000; 46: 468–473.
- 51 Gershon MD. Review article: roles played by 5-hydroxytryptamine in the physiology of the bowel. *Aliment Pharmacol Ther* 1999; **13**: 15–30.
- 52 Gershon MD, Wade PR, Kirchgessner AL, Tamir H. 5-HT receptor subtypes outside the central nervous system. Roles in the physiology of the gut. *Neuropsychopharmacology* 1990; **3**: 385–395.
- 53 El-Salhy M, Mazzawi T, Gundersen D, Hatlebakk JG, Hausken T. The role of peptide YY in gastrointestinal diseases and disorders (review). *Int J Mol Med* 2013; **31**: 275–282.
- 54 El-Salhy M, Gilja OH, Gundersen D, Hatlebakk JG, Hausken T. Interaction between ingested nutrients and gut endocrine cells in patients with irritable bowel syndrome (review). Int J Mol Med 2014; **34**: 363–371.
- 55 El-Salhy M, Hatlebakk JG, Gilja OH, Hausken T. Irritable bowel syndrome: recent developments in diagnosis, pathophysiology, and treatment. *Expert Rev Gastroenterol Hepatol* 2014; **8**: 435–443.
- 56 Hocker M, Wiedenmann B. Molecular mechanisms of enteroendocrine differentiation. *Ann NY Acad Sci* 1998; **859**: 160–174.
- 57 Inokuchi H, Fujimoto S, Kawai K. Cellular kinetics of gastrointestinal mucosa, with special reference to gut endocrine cells. *Arch Histol Jpn* 1983; **46**: 137–157.

This work is licensed under a Creative Commons Attribution 4.0 International License. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in the credit line; if the material is not included under the Creative Commons license, users will need to obtain permission from the license holder to reproduce the material. To view a copy of this license, visit http://creativecommons.org/licenses/ by/4.0/