

Neuronal plasticity of trigeminal ganglia in mice following nerve injury

Randi Lynds^{1,2,*}

Chuang Lyu^{3,*}

Gong-Wei Lyu⁴

Xie-Qi Shi^{1,2}

Annika Rosén^{5,6}

Kamal Mustafa⁶

Tie-Jun Sten Shi⁷

¹Department of Dental Medicine, Karolinska Institutet, Huddinge, Sweden; ²Division of Oral and Maxillofacial Radiology, Department of Clinical Dentistry, Faculty of Medicine and Dentistry, University of Bergen, Bergen, Norway; ³State Key Laboratory of Veterinary Biotechnology, Harbin Veterinary Research Institute of Chinese Academy of Agricultural Sciences, ⁴Department of Neurology, The First Hospital of Harbin Medical University, Harbin, People's Republic of China; ⁵Division for Oral and Maxillofacial Surgery, ⁶Tissue Engineering Group, Department of Clinical Dentistry, Faculty of Medicine and Dentistry, ⁷Department of Biomedicine, University of Bergen, Bergen, Norway

*These authors contributed equally to this work

Background: Nerve injury may induce neuropathic pain. In studying the mechanisms of orofacial neuropathic pain, attention has been paid to the plastic changes that occur in the trigeminal ganglia (TGs) and nucleus in response to an injury of the trigeminal nerve branches. Previous studies have explored the impact of sciatic nerve injury on dorsal root ganglia (DRGs) and it has shown dramatic changes in the expression of multiple biomarkers. In large, the changes in biomarker expression in TGs after trigeminal nerve injury are similar to that in DRGs after sciatic nerve injury. However, important differences exist. Therefore, there is a need to study the plasticity of biomarkers in TGs after nerve injury in the context of the development of neuropathic pain-like behaviors.

Aim: The aim of this study was to investigate the plasticity of biomarkers associated with chronic persistent pain in TGs after trigeminal nerve injury.

Materials and methods: To mimic the chronic nature of the disorder, we used an intraoral procedure to access the infraorbital nerve (ION) and induced a nerve injury in mice. Immunohistochemistry and quantification were used for revealing the expression level of each biomarker in TGs after nerve injury.

Results: Two weeks after partial ION injury, immunohistochemistry results showed strongly upregulated expressions of activating transcription factor 3 and neuropeptide Y (NPY) in the ipsilateral TGs. Microglial cells were also activated after nerve injury. In regard to positive neuronal profile counting, however, no significant difference in expression was observed in galanin, substance P, calcitonin gene-related peptide, neuronal nitric oxide synthase, phosphorylated AKT, or P2X3 in ipsilateral TGs when compared to contralateral TGs.

Conclusion: In this study, the expression and regulation of biomarkers in TGs have been observed in response to trigeminal nerve injury. Our results suggest that NPY and Iba1 might play crucial roles in the pathogenesis of orofacial neuropathic pain following this type of injury. Further investigations on the relevance of these changes may help to target suitable treatment possibilities for trigeminal neuralgia.

Keywords: infraorbital nerve, orofacial pain, sensory neurons, neuropeptides, animal model

Introduction

Trigeminal neuralgia is regarded as being one of the most painful diseases known.¹ Pain resulting from this disease impairs patients' mood, quality of life, daily life activities, and work performance.² Known factors that can cause trigeminal neuralgia are injury, inflammation, and vascular compression of the trigeminal nerve.^{3,4} However, the mechanisms of trigeminal neuralgia are still poorly understood.

Correspondence: Tie-Jun Sten Shi
Department of Clinical Dentistry,
University of Bergen, Årstadveien 19, PO
Box 7804, 5020 Bergen, Norway
Tel +47 4539 2855
Email tjstenshi@gmail.com

Sensory neuron plasticity after peripheral injury is considered important for understanding the development of chronic persistent pain.⁵ Studies have revealed that the impact that sciatic nerve injury has on dorsal root ganglia (DRGs) causes dramatic changes in the expression of certain peptides, more specifically excitatory peptides such as substance P (SP) and calcitonin gene-related peptide (CGRP), as well as inhibitory and excitatory neuropeptides such as galanin (GAL) and neuropeptide Y (NPY).^{6,7} Neuronal nitric oxide synthase (nNOS) and P2X3 are also known to play a role in nociception.^{8,9} Phosphorylated AKT (pAKT) is believed to have neuroprotective effects and can be regulated in sensory ganglia after injury.^{5,10,11} All of these changes may have implications in the generation and modulation of neuropathic pain.¹² To our knowledge, only a limited number of studies have been conducted to examine neuronal plasticity of the trigeminal ganglion (TG) following trigeminal nerve injury. These studies show that the changes in neuropeptide expression of TGs after trigeminal nerve injury are similar to DRGs after sciatic nerve injury.^{13,14} However, important differences exist. For example, GAL is markedly upregulated in DRGs after sciatic nerve injury,¹⁵ whereas GAL expression in TGs has been inconclusive, with results showing an increase, no change, or even decrease in expression after facial nerve injury.^{13,16} Interestingly, systematic studies have not been conducted on the expression of neuropeptides in TGs after nerve injury in the context of the development of neuropathic pain-like behaviors, as in the case of sciatic nerve injury.¹⁷

Recently, several animal models have been developed to examine trigeminal nerve injury, inflammation, and trigeminal nerve root compression.^{18–21} Among these models, the infraorbital nerve (ION) constriction model of trigeminal neuralgia has been performed in both rats and mice.^{22–24} Interestingly, animal models of trigeminal neuralgia obtained by a chronic constriction injury of ION are shown to share many characteristics with clinical disorders in humans.^{25,26}

Because pain-related biomarkers give accurate signals, observing and understanding their changes may help us find potential targets for treating painful diseases. In this study, we investigated the expression of several biomarkers in TGs following trigeminal nerve injury. Expression and regulation of activating transcription factor 3 (ATF3), NPY, Iba1, GAL, CGRP, SP, nNOS, pAKT, and P2X3 were examined in a modified mouse model of trigeminal neuralgia.

Materials and methods

Animals

Adult male C57BL/6J Bommince mice (A/S Bomholtgaard, Ry, Denmark) weighing 28–30 g were used in this study.

All animals were kept under standard conditions on a 12-h day/night cycle with free access to food and water. The experiments were carried out according to the International Association for the Study of Pain Guidelines for the Use of Animals in Research and were approved by Karolinska Institutet ethics committee (registration number: N48/13).

Operation

Mice were anesthetized with isoflurane, received a transection to the left ION, and the contralateral side (right side) was used as a control. The surgery was performed intraorally, which allowed the hair on the snout and the vibrissae to be kept intact. A transection of ~0.5 cm was made unilaterally along the left gingivobuccal margin. The transection began at ~0.1 cm proximal to the first molar. The modification of surgery is based on two previously published studies.^{27,28} The mice were sacrificed 2 weeks after surgery.

Immunohistochemistry

All operated animals were deeply anesthetized with sodium pentobarbital (50 mg/kg, i.p.) and transcardially perfused with 20 mL of warm saline (0.9%; 37°C), followed by 20 mL of a warm mixture of paraformaldehyde (4%; 37°C) with 0.4% picric acid in 0.16 M phosphate buffer (pH 7.2) and then by 50 mL of the same, but ice-cold fixative. Both the ipsilateral and contralateral TGs were dissected out and postfixed in the same fixative for 90 min at 4°C and subsequently stored in 10% sucrose in phosphate-buffered saline (PBS; pH 7.4) containing 0.01% sodium azide (Sigma-Aldrich Co., St Louis, MO, USA) and 0.02% bacitracin (Sigma-Aldrich Co.) at 4°C for 2 days. Tissues were embedded with optimum cutting temperature compound (Tissue-Tek; Miles Laboratories, Elkhart, IN, USA), frozen and cut in a cryostat (Microm, Heidelberg, Germany) at 14 µm thickness, and then mounted onto chrome-alum-gelatin-coated slides. Thaw-mounted sections were dried at room temperature (RT) for 30 min and rinsed with PBS for 15 min. Sections were incubated for 24 h at 4°C in a humid chamber with rabbit anti-ATF3 (1:4,000; D1610, Santa Cruz Biotechnology, Dallas, TX, USA), NPY (1:8,000),²⁹ rabbit anti-GAL (1:8,000),³⁰ rabbit anti-SP (1:4,000),³¹ rabbit anti-CGRP (1:10,000),³² sheep anti-nNOS (1:4,000),³² rabbit anti-pAKT (1:4,000; G7441, Promega Corporation, Madison, WI, USA), and rabbit anti-P2X3 (1:4,000)³³ antisera diluted in PBS containing 0.2% (w/v) bovine serum albumin and 0.03% Triton X-100. Immunoreactivity was visualized using the tyramide signal amplification system (TSA Plus; NEN Life Science Products, Boston, MA, USA). Briefly, the slides were rinsed with Tris-HCL/NaCl/Tween-20 (TNT) buffer (0.1 M Tris-HCL, pH 7.5;

0.15 M NaCl; 0.05% Tween-20) for 15 min at RT, blocked with Tris-HCL/NaCl/DuPont blocking reagent (TNB) buffer (0.1 M Tris-HCL; pH 7.5; 0.15 M NaCl; 0.5% DuPont blocking reagent) for 30 min at RT followed by a 30-min incubation with horseradish peroxidase-labeled swine anti-rabbit and/or anti-sheep antibody (1:200; Dako Denmark A/S, Glostrup, Denmark) diluted in TNB buffer. After a quick wash (15 min) in TNT buffer, all sections were exposed to biotiny tyramide-fluorescein (1:100) diluted in amplification diluent for 10 min, and finally washed in TNT buffer for 30 min (all steps performed at RT). Sections to be used for profile counts (with nucleus) were counterstained for 8–10 min in 0.001% (w/v) propidium iodide (PI; Sigma-Aldrich Co.) in PBS, rinsed twice for 5 min in PBS, and mounted with glycerol/PBS (9:1) containing 2.5% DABCO (Sigma-Aldrich Co.).

Image analysis and quantification

Specimens were analyzed on a Bio-Rad Radiance Plus confocal scanning microscope (Bio-Rad, Hemel Hempstead, UK) installed on a 710 LSM laser-scanning microscope equipped with $\times 10$ (0.5 numerical aperture [NA]), $\times 20$ (0.75 NA), and $\times 60$ (1.40 NA) oil objectives. Fluorescein labeling was excited using the 488 nm line of the argon ion laser and detected after passing a HQ 530/60 (Bio-Rad Laboratories Inc.) emission filter. All the slides were scanned in a series of 1 μm thick optical sections.

To determine the percentage of immunoreactive (IR) neuronal profiles (NPs), the counting was performed on 14- μm -thick sections, and every fifth section was selected. The total number of immunopositive NPs was divided by the total number of PI-stained NPs in the TG sections, and

the percentage of positive NPs was calculated. Four to eight sections of each TG from five animals in each group (i.e., ipsilateral and contralateral groups) were included in the analysis, and 1,200–1,800 NPs were counted in each ganglion.

Statistical analysis

Unpaired Student's *t*-test was used for the comparison of difference between ipsilateral and contralateral expression of each biomarker. $p < 0.05$ was taken as the criterion for statistical significance ($*p < 0.05$, $**p < 0.01$, and $***p < 0.001$).

Results

ATF3 expression in TGs after ION injury

ATF3-IR NPs were almost undetectable in the contralateral TGs 2 weeks after partial ION injury. However, a strong upregulation of ATF3-IR NPs was seen in the ipsilateral TGs, as shown in Figure 1A, B versus C and Figure 2 ($0.9 \pm 0.2\%$ vs. $30.1 \pm 5.8\%$; $p < 0.001$), mainly in the maxillary infraorbital neurons (Figure 1A; red box).

Expression of biomarkers in TGs after ION injury

In the contralateral TGs, very few NPs were NPY positive ($1.8 \pm 0.9\%$), while a dramatic increased expression of NPY-positive NPs was seen in the ipsilateral TGs ($12.9 \pm 5.6\%$; $p < 0.01$; Figures 2–4A and B). This increase is ~ 10 -fold when compared to the control. Regarding GAL expression, more positive NPs were found in the ipsilateral TGs, but this increase in expression was not statistically significant ($11.5 \pm 5.8\%$ vs. $13 \pm 4.8\%$; $p > 0.05$; Figures 2 and 3). No significant difference was detected between the ipsilateral and contralateral TGs for

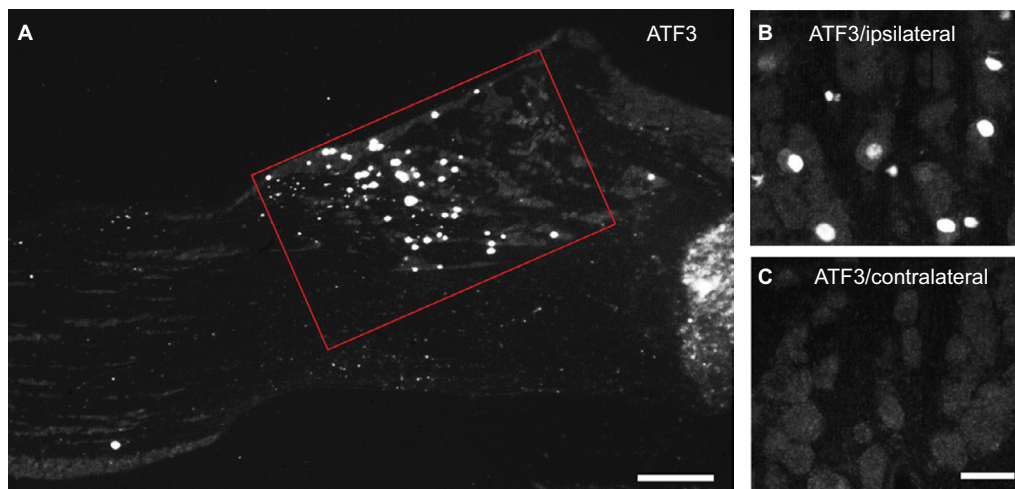


Figure 1 Expression of ATF3-LI in TGs 2 weeks after partial ION transection.

Notes: (A) ATF3-LI is mainly seen in the infraorbital neurons (red box) in ipsilateral TGs. (B and C) High magnification micrographs show the expression of ATF3-IR neurons in ipsilateral (B) and contralateral (C) TGs. Scale bars indicate 250 μm (A) and 50 μm (B and C).

Abbreviations: ATF3, activating transcription factor 3; ION, infraorbital nerve; IR, immunoreactive; LI, like immunoreactivity; TGs, trigeminal ganglia.

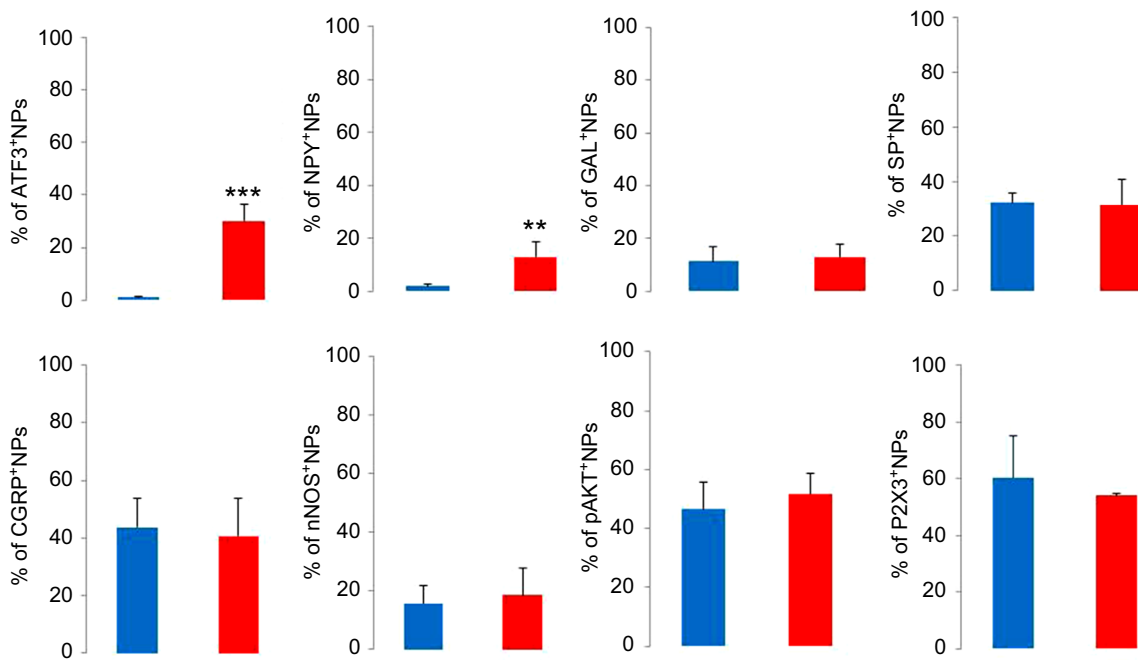


Figure 2 Percentage of positive NPs in contralateral and ipsilateral TGs 2 weeks after partial ION transection.

Note: The columns represent contralateral (blue) and ipsilateral (red) TGs, respectively (n=5 in each group, **p<0.01, ***p<0.001).

Abbreviations: ATF3, activating transcription factor 3; CGRP, calcitonin gene-related peptide; GAL, galanin; ION, infraorbital nerve; nNOS, neuronal nitric oxide synthase; NPs, neuronal profiles; NPY, neuropeptide Y; pAKT, phosphorylated AKT; SP, substance P; TGs, trigeminal ganglia.

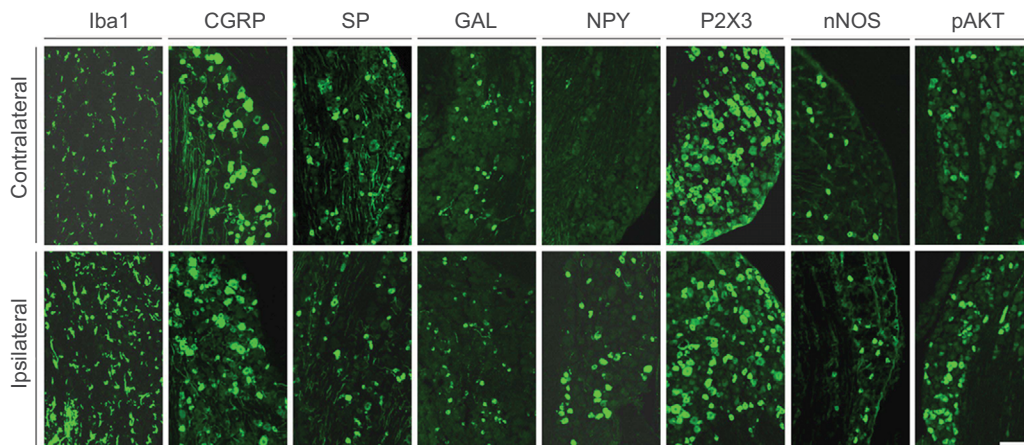


Figure 3 Immunofluorescent micrographs show the expression of biomarkers in contralateral (top panel) and ipsilateral (bottom panel) TGs 2 weeks after partial ION transection.

Notes: Sections were incubated with antisera of Iba1, CGRP, SP, GAL, NPY, P2X3, nNOS, and pAKT, respectively. Scale bar indicates 100 μ m.

Abbreviations: CGRP, calcitonin gene-related peptide; ION, infraorbital nerve; nNOS, neuronal nitric oxide synthase; NPY, neuropeptide Y; pAKT, phosphorylated AKT; SP, substance P; TGs, trigeminal ganglia.

CGRP (43.7 \pm 9.9% vs. 40.9 \pm 13.0%; Figures 2, 3, and 5A), SP (32.2 \pm 3.7% vs. 31.6 \pm 9.1%; Figures 2, 3, and 5B), P2X3 (60.4 \pm 15.8% vs. 54.0 \pm 1.3%; Figures 2 and 3), or nNOS (15.9 \pm 6.0% vs. 18.6 \pm 9.3%; Figures 2 and 3), respectively. pAKT-like immunoreactivity (pAKT-LI) was seen in many neurons in intact TGs (Figure 5C), especially small-sized ones. pAKT had a relatively high intensity in ipsilateral TGs but the percentage of positive-NPs remained the same 2 weeks after ION injury (46.6 \pm 9.2% vs. 51.5 \pm 7.1%; Figures 2 and 3).

Iba1, a marker for activated microglial cells, was activated in the ipsilateral TGs as compared to contralateral ones 2 weeks after partial ION injury (Figures 3 and 4C–F). The expression of Iba1-LI in the contralateral TGs presented a typical resting microglial morphology (Figures 3 and 4C and E). By contrast, the enlarged and amoeboid morphological features of activated microglia cell bodies were seen in the ipsilateral TGs (Figures 3 and 4D and F), which are consistent with previous findings.³⁴

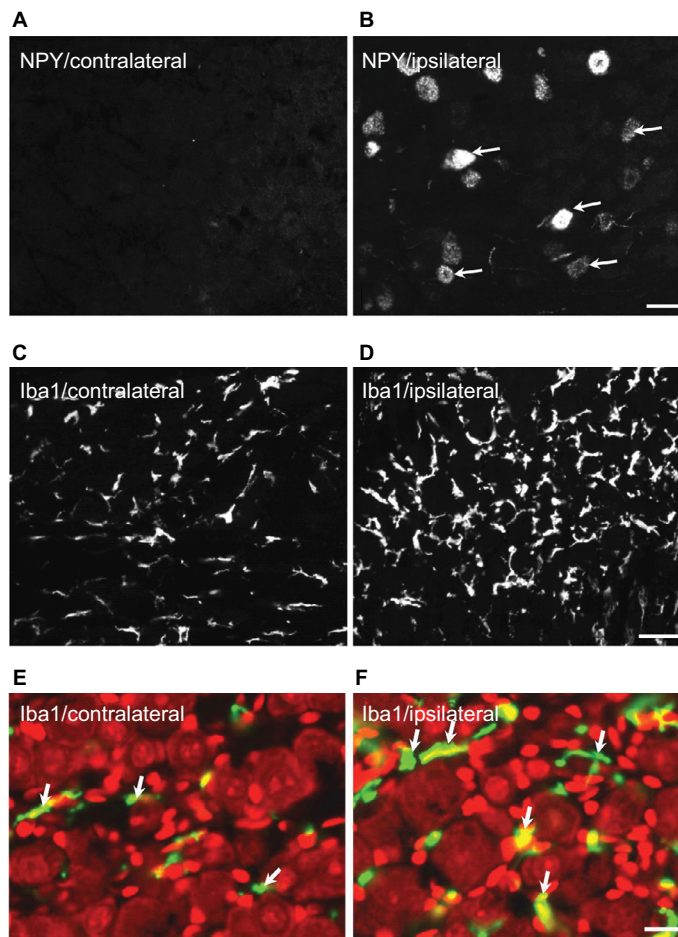


Figure 4 Expression of NPY and Iba1 in ipsilateral and contralateral TGs 2 weeks after partial ION transection.

Notes: NPY-LI is upregulated in ipsilateral TG (B) compared with the contralateral one (A). Immunofluorescence micrographs show a hypertrophic morphology of Iba1-LI in ipsilateral (D and F) vs. contralateral TGs (C and E), counterstaining with PI (red) (E and F). Arrows indicate the NPY-positive neurons (B) and Iba1-positive microglial cells (E and F), respectively. Scale bars indicate 50 μ m (A and B), 30 μ m (C and D), and 25 μ m (E and F).

Abbreviations: ION, infraorbital nerve; LI, like immunoreactivity; NPY, neuropeptide Y; PI, propidium iodide; TGs, trigeminal ganglia.

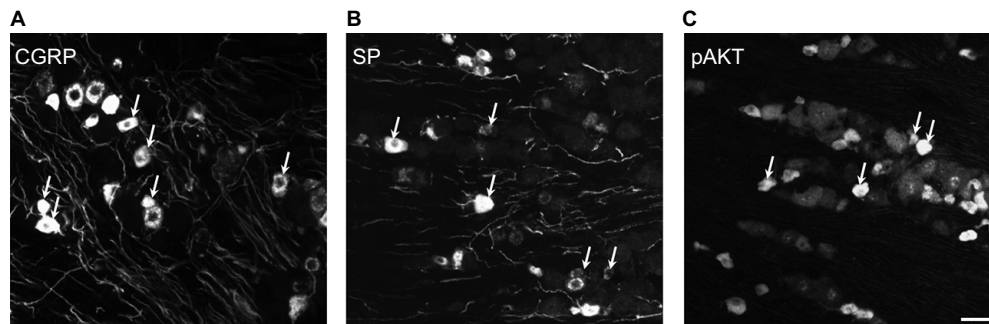


Figure 5 Immunofluorescent micrographs show the expression of biomarkers in contralateral TGs 2 weeks after partial ION transection.

Notes: CGRP-LI (A), SP-LI (B), and pAKT-LI (C) are present in the TG neurons. Arrows indicate positive neurons. Scale bar indicates 50 μ m.

Abbreviations: CGRP, calcitonin gene-related peptide; ION, infraorbital nerve; LI, like immunoreactivity; SP, substance P; TGs, trigeminal ganglia; pAKT, phosphorylated AKT.

Discussion

In this study, we investigated the expression and regulation of several biomarkers in TGs 2 weeks after partial ION injury in mice. Following the ION transection and based on the NP counting method, dramatic changes of ATF3, Iba1,

and NPY-LIs were observed but minor to no changes of SP, CGRP, nNOS, pAKT, or P2X3 were found in injured TGs as compared with contralateral uninjured controls. These findings suggest that distinct mechanisms of adaptation to nerve injury may exist in TGs when compared to spinal

ganglia, and that NPY and activated glial cells may play a more important role in trigeminal neuralgia.

Several animal models have been developed for mimicking the injured condition of the human trigeminal nerve.^{35,36} Regarding the model of ION injury, the surgery has mostly been performed on rats by incising the skin above the unilateral eye.^{37,38} It is known that many nerve fibers of the facial nerve are distributed around eyes, both in superficial and deep layers. Therefore, exposing the ION through the extraoral approach can inevitably cause damage to the facial nerve and other tissues. This extra tissue damage may disturb the results of behavioral studies and make it more difficult to determine the origin of pain. The model used in this study was modified from a previous report, i.e., the surgery was carried out intraorally, and the damage of other cranial nerves and tissues was greatly limited.²⁷

ATF3 is often used as a marker of damaged neurons, as its upregulation indicates subtle damage to the neurons and/or their axons.^{39,40} Here, we report that ATF3 is significantly upregulated in the ipsilateral TGs vs. the contralateral TGs 2 weeks after partial injury of ION. ATF3-LI was seen in the axotomized neurons, especially in the nucleus and sometimes cytoplasm. In addition, most maxillary infra-orbital neurons were ATF3 positive in the ipsilateral TGs, indicating that the axons of these neurons projecting to ION were injured following partial ION transection. This result further demonstrates that the transection of ION can induce a specific nocuous stimulation on the corresponding territory innervated by ION afferents, triggering dramatic plasticity changes in neuronal receptive fields in the TGs, and thus further reflects that the partial ION injury model of trigeminal neuralgia in mouse used in this study has achieved an injured specificity. These results are in line with previous findings that an increase of ATF3-LI was seen in the ipsilateral TG neurons, in the ION-innervated region, after partial ION ligation.⁴¹

Recent studies suggest that microglial cells are vital players in the immune response to injury and are largely implicated in chronic pain conditions. In most cases, activation of microglial cells accompanies an increased expression of a biomarker, i.e., Iba1.^{42,43} Moreover, microglial activation in the spinal cord contributes to pain hypersensitivity, and it has been shown that pharmacological modulation of spinal microglial responses can effectively mitigate chronic pain.⁴⁴ In the present study, microglial activation accompanying phenotype changes was observed in the ipsilateral TGs stained with anti-Iba1 antisera following ION injury, indicating that activated microglial cells may contribute to the initiation and

maintenance of neuropathic orofacial pain after trigeminal nerve injury.

NPY is widely distributed in the central and peripheral nervous systems.⁴⁵ Both inhibitory and excitatory effects have been reported for NPY, depending on which receptor is involved.^{46,47} Our findings that NPY is low in the contralateral TGs, but significantly upregulated in the ipsilateral TGs after ION injury, are similar to NPY expression in DRG neurons after sciatic nerve injury.^{16,48–50} Our data are also in agreement with several previous reports, which indicate that NPY was upregulated in TGs following inferior alveolar nerve (IAN) injury.^{49,51,52} A very recent study using a mental nerve injury model in rats also showed an increased expression of NPY in ipsilateral TGs.⁵³ Thus, upregulation of NPY in injured TG neurons indicates a role of NPY in persistent abnormal sensations.

GAL, a 29-amino acid peptide, appears to play a role in pain modulation.⁵⁴ GAL and its mRNA are found in TGs. After peripheral injury, no change, upregulation, or downregulation of GAL in TGs has been reported.^{16,55,56} Our data here are in agreement with previous reports that GAL is also expressed in mouse TGs. However, in this study, no significant change of GAL was found in injured TGs after ION injury, indicating that peripheral nerve injury has less influence on GAL synthesis in the TGs than in the DRGs. The quantitative differences in GAL expression from these studies may be related to variations in time periods, models, or species.

It has been established that CGRP and SP have excitatory effects on DRG and spinal neurons.⁵⁷ Regarding regulation in the DRGs, peripheral nerve injury reduces the expression of CGRP and SP, while peripheral inflammation induced by complete Freund's adjuvant or carrageenan instead increases their expressions.^{58–60} Interestingly, in ferret TGs, the expression of CGRP and SP was either downregulated after IAN transection or showed no change after IAN ligation.^{13,16} Here, we report no significant changes of CGRP or SP in mouse TGs following partial ION injury. Our data suggest that these two neuropeptides may have less impact on ION injury-induced abnormal sensations in mice.

Recent studies indicate that purinergic signaling plays important roles in physiological and pathophysiological processes.^{61,62} Purinergic receptors are widely distributed in mammalian tissues, and these receptors are generally divided into three classes: P1, P2X, and P2Y receptors. P2X receptors are ionotropic and ATP sensitive and have been shown to be involved in nociceptive processing.⁶³ P2X3 is mainly expressed in small-sized, nonpeptidergic neurons but also

seen in some medium- and large-sized neurons in DRGs and can be regulated in different animal models.^{64–66} In rat TGs, transient expression of P2X3 has been detected in response to trigeminal nerve injury.^{64,67} Although the proportion of P2X3-positive NPs in mouse TGs remains unchanged at 2 weeks postoperation in the present study, a role of P2X3 in mediating the abnormal nociceptive process may still exist via enhanced sensitivity of P2X3 receptors as suggested by a previous study.⁶³

It is known that nNOS produces nitric oxide (NO) in the nervous system.⁶⁸ As a neuronal neurotransmitter and/or neuromodulator, NO is involved in multiple biological processes, including modulation of pain.^{69–71} nNOS is regulated following the peripheral nerve injury in both DRGs and TGs,^{72,73} and nerve injury-induced hypersensitivity or allodynia can be attenuated by a pharmacological inhibition of nNOS.^{74–76} In the present study, we quantified the number of nNOS-positive NPs in TGs 2 weeks after ION transection. We did not find a significant difference in nNOS expression between the ipsilateral and contralateral TGs. In contrast to our findings, a significant upregulation of nNOS-IR neurons was seen in ipsilateral TGs 7 days after IAN injury, accompanied by significantly increased levels of two NO indicators, nitrate and nitrite.⁷⁵ Interestingly, the same group also demonstrated that the levels of the two indicators in ipsilateral TGs went back to the control levels 14 days after injury, suggesting that a transient expression pattern of nNOS exists in injured TGs. Furthermore, the dramatic increase of nNOS in TGs 7 days, but not 14 days, after injury implies that NO may play a role at the early stage of neuropathic pain.

Phosphatidylinositol 3-kinase (PI3-K)/AKT signaling is believed to be important for promoting neuronal cell survival and axonal outgrowth.⁷⁷ We have previously investigated the expression of pAKT in mouse DRGs and spinal cord following sciatic nerve axotomy or carrageenan-induced inflammation.⁵ In normal control animals, pAKT was found in both peptidergic and nonpeptidergic DRG neurons. After nerve injury, pAKT-IR neurons were seen in ipsilateral DRG neurons coexpressed with GAL but not NPY. In addition, we also found that the intensity of pAKT-LI in DRGs was increased in axotomized animals. To our knowledge, pAKT expression in TGs has been much less characterized. In this study, we reported that peripheral ION injury did not change the proportion of pAKT-positive NPs. However, even without quantification, an increased intensity of pAKT-LI was detected in the ipsilateral TGs. Our findings suggest that there is a PI3-K/AKT signaling pathway in trigeminal system.

Conclusion

Trigeminal nerve injury causes marked plasticity of biomarkers in TGs. Significant changes observed in injured TGs suggest that NPY and Iba1 may play crucial roles in the pathogenesis of orofacial neuropathic pain, in response to peripheral nerve injury.

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Author contributions

RL, CL, GL, and T-JSS performed experiments, analyzed the data, and wrote the manuscript. XS, AR, and KM analyzed the data and wrote the manuscript. All authors contributed toward data analysis, drafting and critically revising the paper and agree to be accountable for all aspects of the work.

Disclosure

The authors report no conflicts of interest in this work.

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