A novel MRI index for paraspinal muscle fatty infiltration: reliability and relation to pain and disability in lumbar spinal stenosis: results from a multicentre study

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Abstract

Background: Fatty infiltration of the paraspinal muscles may play a role in pain and disability in lumbar spinal stenosis. We assessed the reliability and association with clinical symptoms of a method for assessing fatty infiltration, a simplified muscle fat index (MFI).

Methods: Preoperative axial T2-weighted magnetic resonance imaging (MRI) scans of 243 patients aged 66.6 ± 8.5 years (mean \pm standard deviation), 119 females (49%), with symptomatic lumbar spinal stenosis were assessed. Fatty infiltration was assessed using both the MFI and the Goutallier classification system (GCS). The MFI was calculated as the signal intensity of the psoas muscle divided by that of the multifidus and erector spinae. Observer reliability was assessed in 102 consecutive patients for three independent investigators by intraclass correlation coefficient (ICC) and 95% limits of agreement (LoA) for continuous variables and Gwet's agreement coefficient (AC1) for categorical variables. Associations with patient-reported pain and disability were assessed using univariate and multivariate regression analyses.

Results: Interobserver reliability was good for the MFI (ICC 0.79) and fair for the GCS (AC1 0.33). Intraobserver reliability was good or excellent for the MFI (ICC range 0.86-0.91) and moderate to almost perfect for the GCS (AC1 range 0.55–0.92). Mean interobserver differences of MFI measurements ranged from -0.09 to -0.04 (LoA -0.32 to 0.18). Adjusted for potential confounders, none of the disability or pain parameters was significantly associated with MFL or GCS.

Conclusion: The proposed MFI demonstrated high observer reliability but was not associated with preoperative pain or disability.

Keywords: Magnetic resonance imaging, Paraspinal muscles, Patient-reported outcome measures, Psoas muscles, Spinal stenosis

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Key points

- Fatty infiltration of the paraspinal muscles can be quantified by magnetic resonance imaging (MRI).
- The proposed muscle fat index (MFI) uses routine lumbar MRI examinations.
- This novel MFI shows high observer reliability for the quantification of muscle fat.
- Increased muscle fat was seen in patients with lumbar spinal stenosis (LSS).
- Significant association between the MFI and symptoms of LSS was not found.

Background

Degenerative lumbar spinal stenosis (LSS) is a clinical condition caused by degenerative changes in the supporting structures of the lumbar spine [1]. Patients with LSS experience varying degrees of disability, low back pain, and radiating pain in lower extremities [2]. Fatty infiltration of the paraspinal muscles is a frequent finding in patients with LSS [3, 4]. Mainly formed by the multifidus (MF) and the erector spinae (ES), these muscles are innervated by the dorsal rami of the L1-L4 nerves. The main function of the paraspinal muscles is extension and rotation of the lumbar spine and to resist gravity [5]. Studies have demonstrated associations between the severity of fatty infiltration of the paraspinal muscles evaluated by magnetic resonance imaging (MRI), and pain and disability reported by patients with LSS [3, 6, 7]. It has been suggested that fatty infiltration of the paraspinal muscles can be used as a predictor of postoperative clinical outcomes and recovery of patients with symptomatic LSS, influencing the treatment decision process [8–10].

Imaging modalities can be used for the assessment and grading of the severity of fatty infiltration in the skeletal muscles. The Goutallier classification system (GCS) is a frequently used semiquantitative grading method for the assessment of muscle fatty infiltration [11]. This method was originally proposed by Goutallier et al. [12] for grading the severity of fatty infiltration in the shoulder rotator cuff muscles on computed tomography (CT) as a prognostic tool for tendon repairs, suggesting a poorer outcome when the cuff muscles had higher fatty infiltration. Fuchs et al. [13] demonstrated good or excellent interobserver reliability for the GCS on shoulder CT and MRI individually, but only fair to moderate correlation between the GCS grading performed on CT and MRI. Despite this inferior correlation, the GCS has been adopted for the evaluation of muscular fatty infiltration on MRI in various anatomical locations, including the paraspinal muscles [14–17]. Both quantitative and semiquantitative MRI methods have been used to assess the severity of fatty infiltration in the paraspinal muscles. It has been suggested that quantitative MRI methods have higher reliability than the semiquantitative methods [18–20]. The main drawbacks of the currently available quantitative methods are time consumption and the need for exporting the images into a third-party software for analysis, making these methods less practical in everyday clinical practice [7, 19, 21, 22].

The muscle fat index (MFI) is a quantitative measure used by researchers to assess the fat content of the paraspinal muscles on MRI, by calculating the ratio of the mean signal intensity of the muscle of interest to a homogenous area of the same or another muscle [23]. In the current study, we introduced a new method for calculating the MFI based on the signal intensity of the paraspinal and the psoas major (PM) muscles measured on axial T2-weighted images from routine lumbar spine MRI examinations, without a need for using a thirdparty software. To our knowledge, this simplified method for calculation of the MFI has not been used earlier. We hypothesised that this easily accessible method might yield higher reliability than the GCS and, furthermore, would associate with the clinical symptoms. The purpose of this study was to evaluate the reliability of this novel MFI and assess its association with pain and disability in patients with LSS.

Methods

Study participants

The regional committees for medical research ethics approved the current cross-sectional study (reference number: 2011/2034 central region). The study adhered to the Declaration of Helsinki and all patients provided written informed consent. The participants in this study were consecutively enrolled from the spinal stenosis trial of the Norwegian Spinal Stenosis and Degenerative Spondylolisthesis (NORDSTEN) study. This multicentre trial includes symptomatic patients with LSS without degenerative spondylolisthesis who are scheduled for surgery. The study protocol and the settings for inclusion and exclusion of the patients have been published earlier [24]. The inclusion and exclusion criteria for the current study are provided in Table 1. After the initial consecutive enrolment of 300 patients (convenient sampling based on the availability of patient data), we excluded 57 patients due to inadequate or missing images, leaving 243 patients who were finally included (Fig. 1).

MRI protocol and assessments

The preoperative MRI examinations used in this study were performed at the local study sites of the NORD-STEN study between February 2013 and August 2016 using 1.5-T or 3.0-T units from several manufacturers, with patients in supine position. All images were anonymised and stored in a dedicated server. To maintain

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Inclusion criteria	Exclusion criteria
 Age between 18 and 80 years Clinical symptoms of LSS Not responding to at least 3 months of non-surgical treatment Radiological findings (foraminal, central canal, or lateral recess stenosis) corresponding to the clinical symptoms such as back pain, leg pain, or neurologic claudication Understanding the Norwegian language (spoken and written) 	 Previous surgery at the level of stenosis Previous fracture or fusion of the thoraco-lumbar spine Cauda equina syndrome (bowel or bladder dysfunction) or fixed complete motor deficit ASA grade 4 or 5 More than 20° lumbosacral scoliosis Distinct symptoms in lower limbs due to other diseases Stenosis in more than three lumbar levels Being unable to comply fully with the protocol Isthmic defect in pars interarticularis at the level of stenosis Participation in another clinical study that could interfere with the present trial Alcohol or substance abuse 2 3 mm spondylolisthesis verified on upright lateral view x-ray Axial T2-weighted MR images not covering the paraspinal and the psoas muscles at both sides of the spine or angulated more than 5° to the upper endplate of the vertebra at the level of measurement

ASA American Society of Anesthesiologists, LSS Lumbar spinal stenosis

homogeneity of the examinations, the performing institutions were provided with a standardised MRI protocol including axial and sagittal T2-weighted and sagittal T1weighted images. A board-certified radiologist (H.B.) verified whether the qualities of the images were adequate for the present study (*e.g.*, the axial images covering both the paraspinal and the PM muscles on both sides of the spine). All measurements for the present study were performed on the axial T2-weighted images (repetition time 1,500–6,548 ms; echo time 82–126 ms; slice thickness 3-4 mm; field of view from 160×160 to 220×220 mm²).

The paraspinal (ES and MF) and the PM muscles were evaluated bilaterally at the level with the upper endplates of L3, L4, and L5 (for both quantitative assessments of the MFI and semiquantitative assessments of the GCS). Inspired by previous studies [21, 23], the investigators segmented the paraspinal and the psoas muscles by drawing manual regions of interest around each muscle group. All segmentations were done using the integrated measurement tools in



a Picture Archiving and Communication System (PACS) (Sectra, Linkoping, Sweden) on personal laptops with nondiagnostic monitors. The mean signal intensity of the MF and the ES muscles was measured by drawing a region of interest around both muscles, excluding the epimuscular fat. The signal intensity of the muscles for each region of interest was calculated automatically by the PACS. To assess the relationship between the fatty infiltration of the paraspinal and the PM muscles, we used the PM muscle as a natural control. It has been suggested that the PM muscle is less prone to fatty infiltration [3, 7, 25]. The MFI was calculated as a continuous variable by dividing the mean signal intensity of the PM with the mean signal intensity of the MF and ES on the same image slice and side. In this way, values close to 1.0 indicated near equal proportions of fat and muscle fibres in the paraspinal muscles compared to the PM, suggesting a very low degree of fatty infiltration; values close to zero suggested a very high degree of fatty infiltration in the paraspinal muscles. An example of this measurement method is shown in Fig. 2.

In the next stage (during the same session and on the same image slice used for calculation of the MFI), the severity of fatty infiltration was graded using the GCS as grade 0 (no fatty streaks), grade 1 (some fatty streaks), grade 2 (fatty infiltration but still more muscle fibres than fat), grade 3 (equal amounts of fat and muscle fibres), or grade 4 (larger amounts of fat than muscle fibres) [12].

Assessment of observer reliability

Interobserver and intraobserver reliability for both methods were assessed for measurements performed at



Fig. 2 Axial T2-weighted magnetic resonance image obtained at the level of the upper endplate of L3. The muscle fat index (MFI) was calculated by dividing the mean signal intensity of the psoas major (PM) with the mean signal intensity of the erector spinae (ES) and the multifidus (MF) muscles

the levels from L2 to L5 for the first 102 consecutive patients. The investigators were three independent observers who were blinded to each other's measurements and to the severity of pain and disability of the patients. They were two orthopaedic spine surgeons (E.H. and J.A. with 10 and 6 years of experience, respectively) and a musculoskeletal radiologist (H.B. with 13 years of experience in spine imaging). To assess the intraobserver reliability and to maintain the independency of the testretest readings, all observers repeated the evaluations after a minimum of 6 weeks, blinded to the results of their first readings. Images with missing measurements or non-optimal axial T2-weighted images (e.g., incomplete imaging of the muscles) were excluded and only levels with measurements from all the three observers were included in the reliability analyses. Prior to the study start, the investigators discussed the measurement criteria for both methods, and the segmentation method was presented to the orthopaedic spine surgeons by the radiologist. They performed test measurements of both the MFI and the GCS on 10 randomly chosen MRI examinations from the study population. The results of the test readings were not included in the statistical calculations.

Assessment of clinical symptoms

Patient-reported outcome measures were used for clinical assessment of pain and disability, including:

- The Oswestry disability index (ODI) [26], a pain and disability index for use in low back pain ranging from 0 to 100, where 0 denotes no disability and 100 indicates complete disability
- The Zurich claudication questionnaire (ZCQ) for pain and disability [27], a disease-specific questionnaire for LSS with several sub-scores including the severity of the symptoms and level of physical activity, ranging from 1 to 5, where 1 indicates the best clinical outcome
- A numeric rating scale (NRS) for back and leg pain ranging from 0 to 10, where 0 indicates no pain and 10 indicates the worst pain imaginable [28]

Statistical analyses

Continuous variables were described as means \pm standard deviations and categorical variables as frequencies and percentages. Intraclass correlation coefficient (ICC) was calculated using two-way random effects models for absolute agreement and was used to assess the interobserver and intraobserver reliability for the MFI. Bland-Altman plots were used to assess the mean differences and 95% limits of agreements (LoA) for repeated measurements. The categorical ratings of the GCS were unevenly distributed, and thus, we used Gwet's agreement coefficient (AC1) instead of κ statistics to assess the interobserver and intraobserver agreements (to avoid the so-called high agreement low kappa paradox) [29]. 95% confidence intervals (CIs) were calculated for both ICC and AC1. ICC values were interpreted to indicate poor (< 0.50), moderate (0.51–0.75), good (0.76–0.90), and excellent (> 0.91) agreement [30] and AC1 values to indicate poor (0.0), slight (0.01–0.20), fair (0.21–0.40), moderate (0.41–0.60), substantial (0.61–0.80), or almost perfect agreement (0.81–1.00) [31].

Observer 3 (radiologist H.B.) performed MFI measurements (continuous) and grading of the GCS (categorical) in the total study sample (243 patients). In few cases, the MFI values were higher than 1.0 and in the absence of apparent fatty infiltration in the PM, these values were redefined as 1.0. The measurements performed by observer 3 were used in the regression analyses and did not differ significantly between lumbar levels or sides (left/ right). Thus, the values representing the highest fatty infiltration (lowest MFI or highest GCS values) from the L2/L3 level were entered into univariate and multivariate regression models, treating all the patient-reported outcome measures as continuous variables. Regression coefficients with corresponding 95% CIs were reported. In the multivariate regression models, we adjusted for age, sex, body mass index, and smoking status (yes or no). Because of the low prevalence of higher GCS grades and for better clinical relevance, we trichotomised the GCS values into category 0 (GCS grade 0, no fatty infiltration), category I (GCS grade 1, mild fatty infiltration), and category II (GCS grades 2 to 4, moderate or severe fatty infiltration) (Table 2). Model assumptions were assessed by normality plots of the standardised residuals and the fitted values. To compare the goodness of fit between the regression models, we calculated the Akaike information criterion (AIC). The AIC is a goodness of fit measure for comparing two models, where the regression model with the lowest AIC value fits better to the data. It has been suggested that an AIC difference of 2 to 7 should be considered as a meaningful difference between two models [32]; others have suggested a minimum difference of 6 AIC units [33]. Values of p lower than 0.05 were considered statistically significant. STATA software (StataCorp. LLC 2017. Stata Statistical Software: Release 16.1 College Station, TX, USA) was used for the statistical analyses.

Results

Patient characteristics

Patient characteristics and distribution of the MRI findings are presented in Table 2. The mean age was 66.6 years and 119 of the 243 included patients (49%) were women. The mean MFI value was 0.53, suggesting overall more than twice fat inside the paraspinal muscles

Table	2 Patient	characteristics	and	distribution	of MR	I finding	1

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Patient characteristics (n = 243)	Mean ± standard deviation/count (%)
Age (years)	66.6 ± 8.5
Female	119 (49)
BMI	27.8 ± 4.0
Smokers	55 (23)
$DSCA < 75 \text{ mm}^2$	
L2/L3 L3/L4 L4/L5	31 (19) 106 (44) 157 (65)
ODI score (100-point scale)	40.66 ± 14.57
NRS back pain (10-point scale)	6.28 ± 2.19
NRS leg pain (10-point scale)	6.38 ± 2.10
ZCQ pain (5-point scale)	3.38 ± 0.55
ZCQ disability (5-point scale)	2.58 ± 0.52
MRI findings (L2/L3 level)	
MFI*	0.53 ± 0.18
GCS	
Category 0 (grade 0) Category 1 (grade 1) Category 2 (grades 2, 3, and 4)	119 (49) 100 (41) 24 (10)

BMI Body mass index, *DSCA* Dural sac cross-sectional area, *GCS* Goutallier classification system, *MFI* Muscle fat index, *NRS* Numeric rating scale, *ODI* Oswestry disability index, *ZCQ* Zurich claudication score *In one case, the value of the MFI was > 1.0, redefined as 1.0

compared to the PM. Most of the patients (n = 219, 90%) had GCS categories 0 or 1 (suggesting no or mild fatty infiltration in the paraspinal muscles) and 24/243 patients (10%) had moderate or severe fatty infiltration (categories 2 to 4). For the reliability part of the study (measurements at the L2–L5 levels), there were 424 GCS assessments and 418 MFI assessments per observer (not included in Table 2). There was an inverse relationship between the different grades of the GCS and the MFI values, indicating higher GCS grades in patients with lower MFI values (Table 3).

Interobserver and intraobserver reliability

The results of the reliability analyses are presented in Table 4. The agreement coefficients suggested good

Table 3 Relationship between the MFL and the	GCS
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GCS grade	Mean MFI	95% CI
0	0.61	0.54, 0.68
1	0.42	0.39, 0.44
2	0.35	0.32, 0.37
3	0.30	0.26, 0.34
4	0.29	0.23, 0.35

CI Confidence interval, GCS Goutallier classification system, MFI Muscle fat index

Table 4 Interobserver and intraobserver reliability

MFI	GCS
ICC (95% CI)	AC1 (95% CI)
0.79 (0.70, 0.85)	0.33 (0.27, 0.39)
0.91 (0.89, 0.92)	0.92 (0.89, 0.95)
0.86 (0.74, 0.91)	0.64 (0.59, 0.70)
0.91 (0.89, 0.93)	0.55 (0.49, 0.60)
	MFI ICC (95% CI) 0.79 (0.70, 0.85) 0.91 (0.89, 0.92) 0.86 (0.74, 0.91) 0.91 (0.89, 0.93)

AC1 Gwet's agreement coefficient, CI Confidence interval, GCS Goutallier classification system, ICC Intraclass correlation coefficient, MFI Muscle fat index

overall interobserver agreement for the MFI and only fair agreement for the GCS. Intraobserver agreement for the three observers was good or excellent for the MFI, while for the GCS, the agreement values ranged from moderate to almost perfect.

Measurement differences for the MFI between all observer pairs, as well as within the observers, are demonstrated by Bland-Altman plots in Figs. 3 and 4, respectively. Mean interobserver differences (*i.e.*, mean bias) ranged from -0.09 to -0.04 with 95% LoA ranging from -0.32 to 0.18. The narrowest LoA for measurements (*i.e.*, the smallest measurement differences) were observed between observers 2 and 3 (one of the two surgeons and the radiologist, Fig. 3c).

Mean intraobserver differences ranged from 0.01 to 0.05 with 95% LoA ranging from -0.15 to 0.22. The narrowest LoA was achieved for observer 1 (one of the two surgeons, Fig. 4a).

Association with clinical symptoms

The results of the univariate regression analyses are presented in Table 5. The estimated regression coefficients were generally small. We found a significant association only between NRS leg pain and the MFI (p = 0.042). A tendency towards lower AIC values was observed for the MFI (suggesting a better fitting to the univariate regression models of the MFI compared to the GCS). The results of the multivariate regression analyses are presented in Table 6. After adjusting for the potentially confounding factors, there were no significant associations between the patient-reported outcome measures and the MFI or the GCS. AIC values were consistently lower for the MFI and were 6 or 7 units lower in the analyses of the ODI and the ZCQ pain, suggesting better fitting of the MFI to the multivariate regression models.

Discussion

In this study, we found a high observer reliability for a novel quantitative MRI method (simplified MFI) in the assessment of fatty infiltration in the paraspinal muscles of patients with symptomatic LSS. For a more established semiquantitative method (the GCS), interobserver reliability was only fair and intraobserver reliability ranged from moderate to almost perfect. We found a significant association between leg pain and the MFI in the univariate regression analyses, but no significant associations in the multivariate analyses. However, the reliability coefficients and the AIC values suggested that the MFI presented here is a better fit to the regression models than the GCS.

Other quantitative methods have been used to assess the fatty infiltration of the paraspinal muscles on MRI. Researchers have used different software applications for texture analysis of the paraspinal muscles [22] or to assess the lean mass of the muscles by thresholding the signal intensity on MR images [7]. Both texture analysis and thresholding of the paraspinal muscles have shown high reliability [19, 21]. DIXON methods have gained increasing interest in spine imaging [34] and have been used for the quantification of fatty infiltration of the paraspinal muscles [35]. However, the need for exporting imaging data into a third-party software and performing additional MRI sequences makes these methods less practical in everyday practice. Whether the proposed MFI in the current study can be used on axial DIXON images of the lumbar spine needs further investigation.



Several studies have examined the relationship be-

tween fatty infiltration of the paraspinal muscles and

symptoms of degenerative diseases of the lumbar spine

[6, 15, 23, 44, 45], but not all studies have considered

the role of the PM muscle in this relationship [6, 15, 45].

To estimate the degree of fatty imbalance between the

PM and the paraspinal muscles, we calculated the MFI

by dividing the signal intensity of the PM with that of

the MF and ES. The calculated mean MFI of 0.53 in the

current study suggested overall less fatty infiltration in

the PM compared to the MF and ES muscles, which is

in accordance with previous research [3, 7, 25]. The im-

portant role of denervation in atrophy and fatty infiltra-

tion of the skeletal muscles have been demonstrated

[46–48]. Higher fatty infiltration in the paraspinal mus-

cles compared to the PM may support the role of damage of the dorsal rami of the lumbar nerves as a cause of

fatty infiltration [49-51]. It is unclear whether nerve

damage can be a common cause for fatty infiltration of

the paraspinal muscles and leg pain in patients with LSS.

However, it is important to be aware of this possible



Previous research has mainly focussed on the associations between back pain and fatty infiltration [6, 8, 22, 36-38]. Leg pain is a frequent symptom in patients with LSS and it is generally accepted that this symptom is a radiating pain from the lumbar spine [2]. There are, however, some controversies on whether this pain is solely generated by the lumbar nerves or can partly be caused by supporting structures of the spine [39, 40]. It has been suggested that patients with LSS and predominant leg pain are more likely to benefit from surgical decompression compared to those with predominant back pain [41]. We did not find a significant association between fatty infiltration of the paraspinal muscles and leg pain after adjusting for potential confounding factors. To our knowledge, this is the first study to examine this association in patients with LSS. Two studies not concerning LSS assessed leg pain in relation to fatty infiltration of the MF muscle with conflicting results. Fatty infiltration of the MF muscle was associated with leg pain in a retrospective study of 78 patients with low back pain [42] but not in a population-based cohort study of young individuals with a history of leg or back pain [43].

Clinical	MFI			GCS*			
parameter	Coefficient (95% CI)	р	AIC	Category	Coefficient (95% CI)	p	AIC
ODI	-6.90 (-17.38, 3.57)	0.195	1940	1	2.65 (-3.91, 9.21)	0.427	1945
				2	10.28 (-6.40, 26.96)	0.226	
NRS leg pain	-1.58 (-3.11, -0.06)	0.042	973	1	0.56 (-0.40, 1.52)	0.249	978
				2	0.66 (-1.72, 3.04)	0.584	
NRS back pain	-0.41 (-2.03, 1.21)	0.618	1011	1	0.86 (-0.15, 1.87)	0.093	1010
				2	1.48 (-1.02, 3.97)	0.245	
ZCQ pain	-0.30 (-0.71, 0.10)	0.136	388	1	0.06 (-0.20, 0.33)	0.629	394
				2	0.35 (-0.29, 0.99)	0.281	
ZCQ disability	-0.25 (-0.63, 0.13)	0.191	368	1	0.08 (-0.16, 0.32)	0.514	370
				2	0.51 (-0.09, 1.11)	0.098	

Table 5 Univariate regression analyses

AIC Akaike information criterion, CI Confidence interval, GCS Goutallier classification system, MFI Muscle fat index, NRS Numeric rating scale, ODI Oswestry disability index, ZCQ Zurich claudication questionnaire

*For the GCS, only categories with fatty infiltration are presented. The coefficient values for the GCS show the estimates for the regression equation of categories 1 and 2, respectively, on category 0 (GCS grade 0, no fatty infiltration)

Clinical parameter	MFI			GCS*			
	Coefficient (95% CI)	р	AIC	Category	Coefficient (95% CI)	р	AIC
ODI	-3.74 (-14.69, 7.21)	0.502	1863	1	0.10 (-5.69, 7.69)	0.769	1869
				2	7.06 (-0.26, 23.38)	0.395	
NRS leg pain	-0.59 (-2.13, 0.96)	0.455	913	1	0.21 (-0.70, 1.12)	0.649	916
				2	-0.23 (-244, 1.98)	0.837	
NRS back pain	0.80 (-0.84, 2.44)	0.339	950	1	0.63 (-0.34, 1.60)	0.199	952
				2	0.75 (-1.61, 3.10)	0.533	
ZCQ pain	-0.22 (-0.64, 0.20)	0.205	364	1	-0.01 (-0.26, 0.26)	0.996	371
				2	0.24 (-0.39, 0.86)	0.454	
ZCQ disability	-0.01 (-0.30, 0.03)	0.979	344	1	0.01 (-0.24, 0.25)	0.972	347
				2	0.40 (-0.19, 0.99)	0.186	

Table 6 Multivariate regression analyses

AIC Akaike information criterion, CI Confidence interval, GCS Goutallier classification system, MFI Muscle fat index, NRS Numeric rating scale, ODI Oswestry disability index, ZCQ Zurich claudication questionnaire

*For the GCS, only categories with fatty infiltration are presented. The coefficient values for the GCS show the estimates for the regression equation of categories 1 and 2, respectively, on category 0 (GCS grade 0, no fatty infiltration)

association in clinical practice. The MFI presented in this study provides a reliable and easy-to-perform quantitative method for assessment of fatty infiltration in the paraspinal muscles on a standard clinical MRI examination without a need of additional software resources and with a high potential to widespread use.

A limitation of this study was the highly symptomatic surgical sample, potentially leading to an underestimation of any association between fatty infiltration and symptoms (due to potential restriction of range) [52]. Furthermore, the results of this study are limited to patients with LSS. Another limitation that may have influenced the reliability was the heterogeneity of the MR images. Images obtained from different MRI units and manufacturers can differ in brightness, affecting the perception of the signal intensity. This may partly explain the lower reliability for the subjectively evaluated GCS in this study, but hardly affected the MFI measurements. We excluded the epimuscular fat of the paraspinal muscles in the MFI measurements; some studies have included this fat in quantitative measurements. There is, however, a lack of consensus on whether the epimuscular fat should be included or excluded from the measurements [20].

We did not measure time consumption in this study, but time is an important factor in clinical and radiological everyday practice. Quantitative MRI methods are generally more time-consuming compared to semiquantitative and qualitative methods [20]. The advent of artificial intelligence methods for automated segmentation of muscles and the integration of these methods with clinical PACS solutions are expected to resolve the timeconsumption issue [53]. We used the signal intensity of the muscles for the assessment of fatty infiltration. It can be argued that the proportion of fat and muscle fibres (used in the GCS) can be applied in artificial intelligence methods to improve the assessment of fatty infiltration in the paraspinal muscles as well (*e.g.*, by calculating the lean muscle to fat ratio). Whether such method would result in better reliability and association with the clinical symptoms of patients with LSS is yet to be examined.

This simplified MFI method using routine MR images should be investigated in a broader patient population with LSS, also including patients without the need of surgical treatment, as well as to see whether fatty infiltration of the paraspinal muscles can be used as a predictor for postoperative outcomes of LSS.

In conclusion, the novel MFI proposed in this study presents a highly reliable method for the assessment of fatty infiltration in the paraspinal muscles using routine spine MRI examinations and measurement tools available in the PACS solutions. This MFI was not significantly associated with pain and disability in LSS but may provide better explanation for symptoms related to fatty infiltration in the paraspinal muscles, compared to the GCS.

Abbreviations

AIC: Akaike information criterion; CI: Confidence interval; CT: Computed tomography; ES: Erector spinae; GCS: Goutallier classification system; ICC: Intraclass correlation coefficient; LoA: Limits of agreements; LSS: Lumbar spinal stenosis; MF: Multifidus; MFI: Muscle fat index; MRI: Magnetic resonance imaging; NRS: Numeric rating scale; ODI: Oswestry disability index; PACS: Picture Archiving and Communication System; PM: Psoas major; ZCQ: Zurich claudication questionnaire.

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Authors' contributions

HB, EH, JA, AN, MA, KS, MG, AE, KI, and CH have designed the current study and have been involved in the acquisition and interpretation of data. HB, TÅM, AE, MG, AN, JIB, CW, and HeB have evaluated the methods and statistics. HB is the primary investigator for this study and has drafted the manuscript under the supervision of EH and AN. All the listed authors have critically revised the manuscript and approved the final version.

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Availability of data and materials

The datasets produced during this study are available from the corresponding author upon a reasonable request.

Declarations

Ethics approval and consent to participate

This study was approved by the Norwegian regional committees for medical research ethics (Reference number: 2011/2034 Central region). All patients signed written informed consent.

Consent for publication

Not applicable

Competing interests

None

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References

- Genevay S, Atlas SJ (2010) Lumbar spinal stenosis. Best Pract Res Clin Rheumatol 24:253–265. https://doi.org/10.1016/j.berh.2009.11.001
- Katz JN, Harris MB (2008) Clinical practice. Lumbar spinal stenosis. N Engl J Med 358:818–825. https://doi.org/10.1056/NEJMcp0708097
- Chen YY, Pao JL, Liaw CK, Hsu WL, Yang RS (2014) Image changes of paraspinal muscles and clinical correlations in patients with unilateral lumbar spinal stenosis. Eur Spine J 23:999–1006. https://doi.org/10.1007/ s00586-013-3148-z

- Yarjanian JA, Fetzer A, Yamakawa KS, Tong HC, Smuck M, Haig A (2013) Correlation of paraspinal atrophy and denervation in back pain and spinal stenosis relative to asymptomatic controls. PM R 5:39–44. https://doi.org/1 0.1016/j.pmrj.2012.08.017
- Kalimo H, Rantanen J, Viljanen T, Einola S (1989) Lumbar muscles: structure and function. Ann Med 21:353–359. https://doi.org/10.3109/0785389890914 9220
- Cooley JR, Walker BF, Ardakani EM, Kjaer P, Jensen TS, Hebert JJ (2018) Relationships between paraspinal muscle morphology and neurocompressive conditions of the lumbar spine: a systematic review with meta-analysis. BMC Musculoskelet Disord 19:351. https://doi.org/10.1186/s12 89-018-2266-5
- Fortin M, Lazáry À, Varga PP, Battié MC (2017) Association between paraspinal muscle morphology, clinical symptoms and functional status in patients with lumbar spinal stenosis. Eur Spine J 26:2543–2551. https://doi. org/10.1007/s00586-017-5228-y
- He K, Head J, Mouchtouris N et al (2020) The implications of paraspinal muscle atrophy in low back pain, thoracolumbar pathology, and clinical outcomes after spine surgery: a review of the literature. Global Spine J 10: 657–666. https://doi.org/10.1177/2192568219879087
- Hori Y, Hoshino M, Inage K et al (2019) Clinical importance of trunk muscle mass for low back pain, spinal balance, and quality of life—a multicenter cross-sectional study. Eur Spine J 28:914–921. https://doi.org/10.1007/ s00586-019-05904-7
- Storheim K, Berg L, Hellum C et al (2017) Fat in the lumbar multifidus muscles - predictive value and change following disc prosthesis surgery and multidisciplinary rehabilitation in patients with chronic low back pain and degenerative disc: 2-year follow-up of a randomized trial. BMC Musculoskelet Disord 18:145. https://doi.org/1 0.1186/s12891-017-1505-5
- Somerson JS, Hsu JE, Gorbaty JD, Gee AO (2016) Classifications in Brief: Goutallier classification of fatty infiltration of the rotator cuff musculature. Clin Orthop Relat Res 474:1328–1332. https://doi.org/10.1 007/s11999-015-4630-1
- Goutallier D, Postel JM, Bernageau J, Lavau L, Voisin MC (1994) Fatty muscle degeneration in cuff ruptures. Pre- and postoperative evaluation by CT scan. Clin Orthop Relat Res 304:78–83
- Fuchs B, Weishaupt D, Zanetti M, Hodler J, Gerber C (1999) Fatty degeneration of the muscles of the rotator cuff: assessment by computed tomography versus magnetic resonance imaging. J Shoulder Elbow Surg 8: 599–605. https://doi.org/10.1016/s1058-2746(99)90097-6
- Battaglia PJ, Maeda Y, Welk A, Hough B, Kettner N (2014) Reliability of the Goutallier classification in quantifying muscle fatty degeneration in the lumbar multifidus using magnetic resonance imaging. J Manipulative Physiol Ther 37:190–197. https://doi.org/10.1016/j.jmpt.2013.12.010
- Mandelli F, Nüesch C, Zhang Y et al (2021) Assessing fatty infiltration of paraspinal muscles in patients with lumbar spinal stenosis: Goutallier classification and quantitative MRI measurements. Front Neurol 12:656487. https://doi.org/10.3389/fneur.2021.656487
- Klemt C, Simeone FJ, Melnic CM, Tirumala V, Xiong L, Kwon YM (2021) MARS MRI assessment of fatty degeneration of the gluteal muscles in patients with THA: reliability and accuracy of commonly used classification systems. Skeletal Radiol 50:665–672. https://doi.org/10.1007/ s00256-020-03611-9
- Thompson SM, Reilly P, Emery RJ, Bull AM (2012) A comparison of the degree of retraction of full-thickness supraspinatus tears with the Goutallier grading system. J Shoulder Elbow Surg 21:749–753. https://doi.org/10.1016/ j.jse.2011.09.019
- Han G, Jiang Y, Zhang B, Gong C, Li W (2021) Imaging evaluation of fat infiltration in paraspinal muscles on MRI: a systematic review with a focus on methodology. Orthop Surg 13:1141–1148. https://doi.org/1 0.1111/os.12962
- Mannil M, Burgstaller JM, Thanabalasingam A et al (2018) Texture analysis of paraspinal musculature in MRI of the lumbar spine: analysis of the lumbar stenosis outcome study (LSOS) data. Skeletal Radiol 47:947–954. https://doi. org/10.1007/s00256-018-2919-3
- Hodges PW, Bailey JF, Fortin M, Battié MC (2021) Paraspinal muscle imaging measurements for common spinal disorders: review and consensus-based recommendations from the ISSLS degenerative spinal phenotypes group. Eur Spine J 30:3428–3441. https://doi.org/10.1007/ s00586-021-06990-2

- Fortin M, Battié MC (2012) Quantitative paraspinal muscle measurements: inter-software reliability and agreement using OsiriX and ImageJ. Phys Ther 92:853–864. https://doi.org/10.2522/ptj.20110380
- 22. Mannil M, Burgstaller JM, Held U, Farshad M, Guggenberger R (2019) Correlation of texture analysis of paraspinal musculature on MRI with different clinical endpoints: Lumbar Stenosis Outcome Study (LSOS). Eur Radiol 29:22–30. https://doi.org/10.1007/s00330-018-5552-6
- D'Hooge R, Cagnie B, Crombez G, Vanderstraeten G, Dolphens M, Danneels L (2012) Increased intramuscular fatty infiltration without differences in lumbar muscle cross-sectional area during remission of unilateral recurrent low back pain. Manual Therapy 17:584–588. https://doi.org/10.1016/j.math.2 012.06.007
- Hermansen E, Austevoll IM, Romild UK et al (2017) Study-protocol for a randomized controlled trial comparing clinical and radiological results after three different posterior decompression techniques for lumbar spinal stenosis: the Spinal Stenosis Trial (SST) (part of the NORDSTEN Study). BMC Musculoskelet Disord 18:121. https://doi.org/10.1186/s12891-017-1491-7
- Arbanas J, Pavlovic I, Marijancic V et al (2013) MRI features of the psoas major muscle in patients with low back pain. Eur Spine J 22:1965–1971. https://doi.org/10.1007/s00586-013-2749-x
- Fairbank JC, Pynsent PB (2000) The Oswestry disability index. Spine (Phila Pa 1976) 25:2940–2952; discussion 2952. https://doi.org/10.1097/00007632-2 00011150-00017
- Stucki G, Daltroy L, Liang MH, Lipson SJ, Fossel AH, Katz JN (1996) Measurement properties of a self-administered outcome measure in lumbar spinal stenosis. Spine (Phila Pa 1976) 21:796–803. https://doi.org/10.1097/ 00007632-199604010-00004
- Ferreira-Valente MA, Pais-Ribeiro JL, Jensen MP (2011) Validity of four pain intensity rating scales. Pain 152:2399–2404. https://doi.org/10.1016/j.pain.2 011.07.005
- Gwet KL (2008) Computing inter-rater reliability and its variance in the presence of high agreement. Br J Math Stat Psychol 61:29–48. https://doi. org/10.1348/000711006x126600
- Koo TK, Li MY (2016) A guideline of selecting and reporting intraclass correlation coefficients for reliability research. J Chiropractic medicine 15: 155–163. https://doi.org/10.1016/j.jcm.2016.02.012
- 31. Landis JR, Koch GG (1977) The measurement of observer agreement for categorical data. Biometrics 33:159–174
- Burnham KP, Anderson DR, Huyvaert KP (2011) AIC model selection and multimodel inference in behavioral ecology: some background, observations, and comparisons. Behav Ecol Sociobiol 65:23–35. https://doi. org/10.1007/s00265-010-1029-6
- Stylianou C, Pickles A, Roberts SA (2013) Using Bonferroni, BIC and AIC to assess evidence for alternative biological pathways: covariate selection for the multilevel Embryo-Uterus model. BMC Med Res Methodol 13:73–73. https://doi.org/10.1186/1471-2288-13-73
- Zanchi F, Richard R, Hussami M, Monier A, Knebel JF, Omoumi P (2020) MRI of non-specific low back pain and/or lumbar radiculopathy: do we need T1 when using a sagittal T2-weighted Dixon sequence? Eur Radiol 30:2583– 2593. https://doi.org/10.1007/s00330-019-06626-6
- Zhao Y, Huang M, Serrano Sosa M et al (2019) Fatty infiltration of paraspinal muscles is associated with bone mineral density of the lumbar spine. Arch Osteoporos 14:99. https://doi.org/10.1007/s11657-019-0639-5
- Steffens D, Hancock MJ, Maher CG, Williams C, Jensen TS, Latimer J (2014) Does magnetic resonance imaging predict future low back pain? A systematic review. Eur J Pain 18:755–765. https://doi.org/10.1002/j.1532-214 9.2013.00427.x
- Endean A, Palmer KT, Coggon D (2011) Potential of magnetic resonance imaging findings to refine case definition for mechanical low back pain in epidemiological studies: a systematic review. Spine (Phila Pa 1976)36:160– 169. https://doi.org/10.1097/BRS.0b013e3181cd9adb
- Ranger TA, Cicuttini FM, Jensen TS et al (2017) Are the size and composition of the paraspinal muscles associated with low back pain? A systematic review. Spine J 17:1729–1748. https://doi.org/10.1016/j.spinee.201 7.07.002
- Kellgren JH (1938) Referred pains from muscle. Br Med J 1:325–327. https:// doi.org/10.1136/bmj.1.4023.325
- Sinclair DC, Feindel WH et al (1948) The intervertebral ligaments as a source of segmental pain. J Bone Joint Surg Br 30b:515–521
- 41. Pearson A, Blood E, Lurie J et al (2011) Predominant leg pain is associated with better surgical outcomes in degenerative spondylolisthesis and spinal

stenosis: results from the Spine Patient Outcomes Research Trial (SPORT). Spine (Phila Pa 1976) 36:219–229. https://doi.org/10.1097/BRS.0b013e3181 d77c21

- Kader DF, Wardlaw D, Smith FW (2000) Correlation between the MRI changes in the lumbar multifidus muscles and leg pain. Clin Radiol 55:145– 149. https://doi.org/10.1053/crad.1999.0340
- Hebert JJ, Kjaer P, Fritz JM, Walker BF (2014) The relationship of lumbar multifidus muscle morphology to previous, current, and future low back pain: a 9-year population-based prospective cohort study. Spine (Phila Pa 1976) 39:1417–1425. https://doi.org/10.1097/brs.00000000000424
- Fortin M, Gibbons LE, Videman T, Battié MC (2015) Do variations in paraspinal muscle morphology and composition predict low back pain in men? Scand J Med Sci Sports 25:880–887. https://doi.org/10.1111/sms.12301
- Kjaer P, Bendix T, Sorensen JS, Korsholm L, Leboeuf-Yde C (2007) Are MRIdefined fat infiltrations in the multifidus muscles associated with low back pain? BMC Med 5:2–2. https://doi.org/10.1186/1741-7015-5-2
- Haig AJ (2002) Paraspinal denervation and the spinal degenerative cascade. Spine J 2:372–380. https://doi.org/10.1016/s1529-9430(02)00201-2
- Hodges P, Holm AK, Hansson T, Holm S (2006) Rapid atrophy of the lumbar multifidus follows experimental disc or nerve root injury. Spine (Phila Pa 1976) 31:2926–2933. https://doi.org/10.1097/01.brs.0000248453.51165.0b
- Liu X, Laron D, Natsuhara K, Manzano G, Kim HT, Feeley BT (2012) A mouse model of massive rotator cuff tears. J Bone Joint Surg Am 94:e41. https:// doi.org/10.2106/jbjs.K.00620
- Chon J, Kim H-S, Lee JH et al (2017) Asymmetric atrophy of paraspinal muscles in patients with chronic unilateral lumbar radiculopathy. Ann Rehabil Med 41:801–807. https://doi.org/10.5535/arm.2017.41.5.801
- Sun D, Liu P, Cheng J, Ma Z, Liu J, Qin T (2017) Correlation between intervertebral disc degeneration, paraspinal muscle atrophy, and lumbar facet joints degeneration in patients with lumbar disc herniation. BMC Musculoskelet Disord 18:167–167. https://doi.org/10.1186/s12891-017-1522-4
- Rantanen J, Hurme M, Falck B et al (1993) The lumbar multifidus muscle five years after surgery for a lumbar intervertebral disc herniation. Spine (Phila Pa 1976)18:568–574. https://doi.org/10.1097/00007632-199304000-00008
- 52. Warne RT (2017) Statistics for the social sciences: a general linear model approach. Cambridge University Press, Cambridge, pp 383–390
- Osorno-Castillo K, Fonnegra RD, Díaz GM (2020) Integration of machine learning models in pacs systems to support diagnostic in radiology services. In: Figueroa-García JC, Garay-Rairán FS, Hernández-Pérez GJ, Díaz-Gutierrez Y (eds) Applied computer sciences in engineering. Springer International Publishing, Cham, pp 233–244

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