



Seasonal and diel variation in greenhouse gas emissions from an urban pond and its major drivers

Tamara J. H. M. van Bergen ^{1,2*} Nathan Barros,^{1,3} Raquel Mendonça,^{1,3} Ralf C. H. Aben,¹ Inge H. J. Althuisen,^{1,4} Vera Huszar,⁵ Leon P. M. Lamers,¹ Miquel Lürling,⁶ Fábio Roland,³ Sarian Kosten ¹

¹Department of Aquatic Ecology and Environmental Biology, Institute for Water and Wetland Research, Radboud University, Nijmegen, The Netherlands

²Department of Environmental Science, Institute for Water and Wetland Research, Radboud University, Nijmegen, The Netherlands

³Laboratory of Aquatic Ecology, Institute of Biology, Federal University of Juiz de Fora, Juiz de Fora, Brazil

⁴Department of Biological Sciences and Bjerknes Centre for Climate Research, University of Bergen, Bergen, Norway

⁵Department of Botany, National Museum, Federal University of Rio de Janeiro, Rio de Janeiro, Brazil

⁶Aquatic Ecology and Water Quality Management Group, Wageningen University, Wageningen, The Netherlands

Abstract

Small water systems are important hotspots of greenhouse gas (GHG) emission, but estimates are poorly constrained as data are scarce. Small ponds are often constructed in urban areas, where they receive large amounts of nutrients and therefore tend to be highly productive. Here, we investigated GHG emissions, seasonal and diel variation, and net ecosystem production (NEP) from an urban pond. In monthly 24-h field campaigns during 11 months, diffusive water–atmosphere methane (CH₄) and carbon dioxide (CO₂) fluxes and CH₄ ebullition and oxidation were quantified. With oxygen (O₂) measurements, NEP was assessed. The pond was a net GHG source the entire year, with an emission of 3.4 kg CO₂ eq m⁻² yr⁻¹. The dominant GHG emission pathway was CH₄ ebullition (bubble flux, 50%), followed by diffusive emissions of CO₂ (38%) and CH₄ (12%). Sediment CH₄ release was primarily driven by temperature and especially ebullition increased exponentially above a temperature threshold of 15°C. The pond's atmospheric CO₂ exchange was not related to NEP or temperature but likely to a high allochthonous carbon (C) input via runoff and anaerobic mineralization of C. We expect urban ponds to show a large increase in GHG emission with increasing temperature, which should be considered carefully when constructing ponds in urban areas. Emissions may partly be counteracted by pond management focusing on a reduction of nutrient and organic matter input.

Although lakes and ponds only cover a small area of the Earth's nonglaciated land area (~ 4%; Verpoorter et al. 2014), they play a disproportionately large role in the global carbon cycle due to the large amount of C they process, store, and transport (Tranvik et al. 2009). The majority of lakes and ponds is a significant source of CO₂ and CH₄ to the atmosphere (Cole et al. 1994; Bastviken et al. 2011), while at the same time, they bury large amounts of C in their sediments (Dean and Gorham 1998; Mendonça et al. 2017). Carbon

emission in lakes and ponds is largely driven by hydrological CO₂ inputs (Weyhenmeyer et al. 2015; Wilkinson et al. 2016) and by mineralization of organic matter (OM) within the system (Algesten et al. 2003). Conversely, primary producers can assimilate CO₂ at such high rates that the system turns into a CO₂ sink (Roland et al. 2010; Pacheco et al. 2013). However, global and regional studies suggest that only a small portion of lakes (6–14%) is undersaturated with respect to the atmospheric CO₂ (Cole et al. 1994; Kosten et al. 2010).

Annually, lentic inland waters emit 0.3 Pg C as CO₂ (Raymond et al. 2013) and 0.08 Pg C as CH₄ (Bastviken et al. 2011). This last value may be an underestimate, as ebullition is likely underestimated (Wik et al. 2016). Although small waterbodies (< 50 ha) comprise 98% of lentic inland waters by number and one third by volume (Downing et al. 2006; Verpoorter et al. 2014), small lakes and ponds are strongly underrepresented in literature (Downing 2010), and greenhouse gas

*Correspondence: t.vanbergen@science.ru.nl

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

Additional Supporting Information may be found in the online version of this article.

(GHG) inventories such as the United Nations Framework Convention on Climate Change. Yet, these small systems are estimated to be responsible for 15% and 41% of the global CO₂ and CH₄ diffusive emission from lentic freshwaters, respectively, and thus are important hotspots of C emission (Holgerson and Raymond 2016).

Here, we investigated GHG emissions from a small nutrient-rich urban pond. Ponds in urban areas are constructed for recreational reasons and also for microclimate regulation, for rainwater storage, or as ornamental features to improve public open spaces (Bolund and Hunhammar 1999). They provide several ecosystem services that enhance human wellbeing (e.g., recreation and microclimate regulation), but ponds also enhance biodiversity by increasing ecological connectivity in urban areas (Gledhill et al. 2008). Since the end of the 19th century, urbanization has increased exponentially and is expected to keep increasing, especially in developing countries (Champion 2001). Urban ponds are mostly shallow and stagnant, which makes them highly susceptible to anthropogenic disturbances and eutrophication (Waajen et al. 2014). They generally receive a large amount of nutrients, for example, from bird droppings, sewage overflow, and stormwater runoff (Scherer et al. 1995; Novotny 1999; Waschbusch et al. 1999; Wong et al. 2000), which frequently leads to phytoplankton blooms (Waajen et al. 2014) and could affect GHG emission.

Especially the high perimeter to surface area ratio of small urban ponds could result in a relatively high input of terrestrial OM, which may play an important role in fueling heterotrophic respiration (Hanson et al. 2007). Even though eutrophic systems tend to have a high gross primary production (GPP; Smith 1979; Balmer and Downing 2011), ecosystem respiration (ER) tends to be high in these systems as well (Sand-Jensen and Staehr 2009). Hence, while in some eutrophic systems, the high primary production rates may lead to a net uptake of CO₂ (Pacheco et al. 2013), this is not necessarily the case as ER and the inflow of CO₂ via surface or groundwater may outweigh the primary producers' CO₂ uptake (Casper et al. 2000; Trolle et al. 2012). In addition, the high OM input, together with the phytoplankton-derived C that tends to be easily degradable, potentially leads to high CH₄ production rates in urban ponds (West et al. 2012; West et al. 2016; Martinez-Cruz et al. 2017).

The high relative contribution of small ponds to freshwater GHG emissions in combination with the scarcity of data regarding pond GHG emissions was our rationale for quantifying whole-year water-atmosphere CO₂ and CH₄ fluxes and their seasonal and diel variation in an urban pond. With an intensive measurement campaign, we aimed to not only quantify CO₂ and CH₄ emission from a temperate and hypereutrophic urban pond but also to obtain insight in temporal and spatial variation of fluxes to optimize future efforts to quantify GHG emissions from small systems. Additionally, we aimed to identify the main ecosystem components—sediment, phytoplankton, heterotrophic picoplankton, and fish—responsible for the within-system CO₂ production and consumption. Our approach combined daytime

and nighttime field measurements on a monthly basis with laboratory assays and modeling.

We hypothesized the pond to be an important GHG emitter as well as an important C sink, with a higher CH₄ emission during warm months. We further hypothesized that the pond would shift between a net C source in fall and winter to a net C sink in spring and summer, mainly due to fluctuations in CO₂ emission caused by phytoplankton growth—i.e., high CO₂ uptake by photosynthesis in summer and much less so in winter. Because of the high primary production, we also expected a high variability in diel CO₂ emission, especially in summer. Due to the shallowness of the pond (average depth of 1.3 m), we expected the sediment to be a major contributor to CO₂ emission but simultaneously an important site for C burial because of a high OM loading.

Materials and methods

In the field, we measured diffusive water-atmosphere fluxes of CH₄ and CO₂, ebullitive CH₄ fluxes, water layer CH₄ oxidation rates, and diel oxygen (O₂) concentrations. The pond's net ecosystem production (NEP), GPP, and ER were calculated from O₂ concentrations. A metabolic model was constructed that included aerobic respiration estimates for each organism group (organisms in the sediment, phytoplankton, heterotrophic picoplankton, and fish), based on temperature and O₂ consumption. The outcome of both the field and laboratory measurements, combined with data from an existing hydrological model for the pond area, and literature data, were joined in a mass balance to unravel the major contributors to the CO₂ flux. Finally, C burial rates in the pond were determined in order to compare C burial and emission rates.

Study site and sampling strategy

This study was performed in manmade urban pond situated in Malden, Gelderland, the Netherlands ("Korte Loef," 51°46'30"N, 5°51'11.22"E). The pond was constructed in 1994 to function as a rainwater overflow for the surrounding urban area. It covers an area of 4635 m², stores approximately 6000 m³ of water, and has distinct central (± 2 m) and littoral (≤ 1 m) zones (respectively, 44% and 56% of the total surface area). Monthly field campaigns were performed from July 2013 till May 2014. During the field campaigns, measurements were performed over 24-h periods at two sampling stations, one in the central and one in the littoral zone, that were placed at the same location every field campaign. Groundwater samples were taken from a well situated 20 m from the shore. Every 15 min, the O₂ concentration, temperature, and pH were measured. Diffusive gas fluxes were measured every 3 h during the campaign, whereas methane ebullition and oxidation were measured separately in the light and dark period. We did not sample in June due to logistic reasons, therefore values for this month were interpolated using the average values of May and July in order to obtain year-round estimates. Areal estimates of gas fluxes for the entire pond were calculated based on the

weighted average of the central and littoral area. In our analyses, we considered the months June, July, and August as the summer season; September, October, and November as fall; December, January, and February as winter; and March, April, and May as spring.

Water chemistry

A depth integrated water sample was taken from each of the sampling stations once per field campaign for chemical analyses. Unfiltered water samples were analyzed for total phosphorus (TP), by inductively coupled plasma spectrometry (ICP-OES iCAP 6000, Thermo Fisher Scientific), and for total inorganic nitrogen (TIN) by measuring and summing the ammonium (NH_4^+) and nitrate (NO_3^-) concentrations. Ammonium and nitrate were measured colorimetrically with an Auto Analyzer 3 system (Bran & Luebbe), using salicylate (Grasshoff and Johannsen 1972) and hydrazine sulfate (Kamphake et al. 1967), respectively. Phytoplankton chlorophyll- α (Chl α) concentration and division over major groups (brown, green, and blue Chl α) were determined using a phytoplankton analyzer system with a portable emitter-detector unit (Phyto-PAM and Phyto-ED, Walz).

O₂, temperature, and pH

During the monthly 24-h field campaigns, we deployed a multiparameter portable meter (HQ40d, Hach) equipped with a luminescent/optical dissolved oxygen probe (LDO101) fitted with a temperature and barometric pressure sensor (model LDO101) and a pH-temperature sensor (model 9156BNWP) at the littoral and at the central station, at 0.5 m depth. Dissolved O₂ concentration, temperature, pH, and barometric pressure were recorded every 15 min. A third multimeter was used to sample vertical dissolved O₂ and temperature profiles at both stations every 3 h.

CO₂ and CH₄ emission

Diffusive CO₂ and CH₄ fluxes across the air-water interface were measured using a transparent Plexiglas floating chamber (30 cm diameter, 20 cm height) connected to a GHG analyzer (GGA) using Off-axis ICOS technology (GGA-24r-EP, Los Gatos Research). A closed loop between the chamber and the detector was established using an inlet and outlet on the top of the chamber. A third opening, fitted with an air-tight screw cap, was used to equilibrate the chamber with atmospheric pressure prior to each measurement. Measurements were conducted every 3 h, by placing the chamber three times (treated as triplicates) at the central and littoral zone. Each measurement took about 200 s, and the GGA was configured to log data every second. The flux of CO₂ and CH₄ was calculated based on the slope of the relationship between the concentration in the chamber headspace and time:

$$F = (V/A) \times \text{slope} \times [(P \times F1 \times F2)/(R \times T)] \quad (1)$$

where F is the gas flux ($\text{g m}^{-2} \text{d}^{-1}$), V is the chamber volume (m^3), A is the chamber surface area (m^2), slope is the slope of the relationship between the CH₄ or CO₂ concentration and the time (ppm s^{-1}), P is atmospheric pressure (Pa), $F1$ is molar mass of CO₂ (44) or CH₄ (16) (g mole^{-1}), $F2$ is the conversion factor of seconds to days, R is gas constant $8.3145 \text{ (m}^3 \text{ Pa K}^{-1} \text{ mol}^{-1}\text{)}$, and T is temperature (K). The values from the triplicate measurements were averaged to give the flux of each station (i.e., central or littoral zone). The fluxes of CO₂ and CH₄ were multiplied by their global warming potential (GWP, 1 and 34 over a 100 yr time horizon, respectively; Myhre et al. 2013) and then summed to obtain the total GHG flux in CO₂ equivalents (CO₂ eq).

Ebullition

In order to quantify the spontaneous release of bubbles from sediments to the atmosphere (ebullition), three inverted funnels with a diameter of 35 cm were connected to water-filled infusion bottles (volume of 0.5 liter) and employed at the pelagic zone of the pond (as in Almeida et al. [2016]). Infusion bottles were replaced at sunset or sundown, and after collection, the gas volume was measured by subtracting the weight of each bottle from the full filled weight (i.e., completely filled with water) of the bottle. The CH₄ concentration of the gas was analyzed on a HP 5890 gas chromatograph equipped with a Porapak Q column (80/100 mesh) and a flame ionization detector (GC-FID, Hewlett Packard).

CH₄ oxidation and sediment CH₄ release

CH₄ oxidation was measured as the decline in CH₄ concentration over time (as in Bastviken et al. [2008]), in three flexible medical blood bags (volume 500 mL) located in the shallow (0.5 m) or deep (1 or 2 m, depending on whether it was the littoral or central zone, respectively) part of the water column, in each sampling station. Separate blood bags were incubated with pond water from the corresponding depth, during day and night to evaluate potential temporal differences in CH₄ oxidation. The exact same procedure was followed as in Almeida et al. (2016). As a proxy for sediment CH₄ release, we used the sum of quantified CH₄ oxidation, CH₄ water-atmosphere diffusion, and CH₄ ebullition. In months where no CH₄ oxidation was quantified due to technical difficulties (January and May), oxidation was estimated as a function of the diffusive CH₄ flux. The rationale behind this is that the methane oxidation rate strongly depends on the methane concentration, which, in turn, affects CH₄ diffusion ($\text{CH}_4 \text{ oxidation} = 0.0476 \times \exp [28.97 \times \text{CH}_4 \text{ diffusion}]$; $R^2_{\text{adj}} = 0.87$).

NEP and respiration by different organism groups

We used 15-min interval O₂ measurements to calculate the O₂ change over time (ΔO_2 ; $\text{g O}_2 \text{ L}^{-1} \text{ h}^{-1}$) and subsequently calculated NEP by integrating ΔO_2 over 24 h (as in Cole et al. [2000] and Staehr et al. [2010]). From the NEP, GPP, and ER were calculated (*see* Supporting Information SI 1 for details). Subsequently, we distinguished different organism groups

contributing to the aerobic ER in the pond: phytoplankton, heterotrophic picoplankton, and methanotrophs in the water layer; organisms in the sediment; and fish. To assess the contribution of all separate organism groups to the ER, we constructed a metabolic model consisting of the relationship between water temperature and O₂ consumption for each organism group. These relationships were based on incubations and—for fish respiration—on literature-derived relationships (see Supporting Information SI 2 for details).

Hydrological inputs and within-pond CO₂ production

For each season, we assessed the relative importance of hydrological CO₂ input via groundwater, within-pond CO₂ production from ecosystem productivity (NEP) and from photochemical mineralization of chromophoric dissolved organic carbon (cDOC) (see Supporting Information SI 3 for details). The CO₂ input from groundwater was calculated based on the estimated groundwater inflow and the CO₂ concentration in the groundwater. Photochemical mineralization cDOC was calculated as a direct response to solar radiation exposure and solely based on the average daily light intensity (as in Vähätalo et al. [2000] and Weyhenmeyer et al. [2015]). Light intensity data were obtained from weather station Volkel, Noord-Brabant, The Netherlands.

C burial

Organic C burial rates were estimated by taking six sediment cores (6 cm diameter) with a piston sampler at locations distributed over the central zone of the pond, determining total C content and dividing the estimated areal C stock of the pond by its age (according to Mendonça et al. [2014]; see Supporting Information SI 4 for details).

Statistics

All statistical analyses were performed using R (version 3.5.0). Statistical significance was determined at $p < 0.05$. The effect of location (littoral or center) and time of day (day or night) on GHG fluxes and methane oxidation were tested using linear mixed-effects and generalized linear mixed-effects models (LMM and GLMM, respectively) using the “lme4” package (Bates et al. 2015; see Supporting Information SI 5 for details). For methane oxidation, the effect of water depth (shallow or deep) was also evaluated. Assumptions of linearity, homoscedasticity, and normality were checked by visual inspection of residual plots and a Shapiro Wilk’s test (function *shapiro.test*), respectively (see Supporting Information SI 5 for details). Simple linear regression and nonlinear regression were performed using the *lm* and *nls* function, respectively. Coefficients and p values for Pearson correlations were obtained using the *rcorr* function (“Hmisc” package). All error values in the manuscript give the standard deviation.

Results

Water chemistry

The yearly average concentrations of TP and TIN were 4.5 ± 0.9 and $54.0 \pm 32.3 \mu\text{mol L}^{-1}$, respectively. The average

Chl α concentration was $81 \pm 29 \mu\text{g L}^{-1}$, with the lowest concentration observed in April ($37 \mu\text{g L}^{-1}$) and highest concentration in July ($119 \mu\text{g L}^{-1}$). The water layer was dominated by cyanobacteria during summer and fall and by green algae during winter and spring. The observed water temperature varied from 4.3°C during winter to 27.8°C during summer and was $13.0^\circ\text{C} \pm 0.7^\circ\text{C}$ on average. The O₂ concentration, measured at 0.5 m depth, varied between 1.6 and 13.5 mg L^{-1} during the year and was on average $9.1 \pm 2.6 \text{ mg L}^{-1}$. The O₂ concentration near the bottom of the pond ranged between approximately 0 and 11 mg L^{-1} . During July, August, and September, diurnal stratification occurred, with mixing occurring at night. During the other months, the pond was homogeneously mixed all day. The pH had a neutral value on average (7.2 ± 0.2), with slightly acidic values in summer (pH around 6.5) and slightly basic values in spring and fall (pH around 8).

Carbon flux and burial

The urban pond was a net C source all year with an average C (CO₂-C and CH₄-C) emission of $1.07 \pm 0.43 \text{ g C m}^{-2} \text{ d}^{-1}$. On average, the diffusive CO₂ emission was $0.95 \pm 0.40 \text{ g CO}_2\text{-C m}^{-2} \text{ d}^{-1}$, with the highest values measured in summer and winter season (Figs. 1, 3). CO₂ diffusion was not related to temperature ($F_{1,9} = 0.204$; $p = 0.66$, $R^2 = 0.02$) or Chl α concentration ($F_{1,9} = 0.008$; $p = 0.93$, $R^2 = 0.001$). Both diffusive and ebullitive CH₄ emission were positively related to temperature ($R^2_{\text{adj}} = 0.86$ and $R^2_{\text{adj}} = 0.82$, respectively), and thus similarly the total CH₄ emission as a sum of these fluxes was positively related to temperature (Fig. 2). The diffusive and ebullitive CH₄ emission were, respectively, 0.02 ± 0.30 and $0.10 \pm 0.49 \text{ g CH}_4\text{-C m}^{-2} \text{ d}^{-1}$. Most CH₄ was emitted via the ebullitive emission pathway (75%), and only approximately 25% was emitted via diffusion. When taking the GWP into account, we found a yearly GHG emission (CO₂ and CH₄ emission combined) of $9.19 \pm 7.99 \text{ g CO}_2 \text{ eq m}^{-2} \text{ d}^{-1}$. The yearly average ebullitive CH₄ emission was $4.63 \pm 6.70 \text{ g CO}_2 \text{ eq m}^{-2} \text{ d}^{-1}$, which was considerably higher than the other fluxes. The yearly average diffusive CO₂ and CH₄ emissions were respectively 3.46 ± 1.48 and $1.09 \pm 4.09 \text{ g CO}_2 \text{ eq m}^{-2} \text{ d}^{-1}$. An annual average C burial rate of $29 \text{ g C m}^{-2} \text{ yr}^{-1}$ was found, which is substantially lower than the annual C emission (CO₂ and CH₄ emission combined) of $391 \text{ g C m}^{-2} \text{ yr}^{-1}$.

Sediment CH₄ release and CH₄ oxidation

The sediment CH₄ release was estimated by summing the average CH₄ diffusion, ebullition, and oxidation in the water column for each month. The estimated sediment CH₄ release was positively related to water temperature (T_w), with the highest release rate observed at the highest water temperature (Fig. 2). There seems to be a threshold around 15°C , above which the sediment CH₄ release increases substantially (from April up to and including September). The estimated sediment CH₄ release varied considerably throughout the year, from $0.01 \pm 0.005 \text{ g CH}_4\text{-C m}^{-2} \text{ d}^{-1}$ in March to $1.04 \pm 0.46 \text{ g CH}_4\text{-C m}^{-2} \text{ d}^{-1}$ in July (Fig. 2). The percentage of estimated sediment CH₄ emission that was oxidized

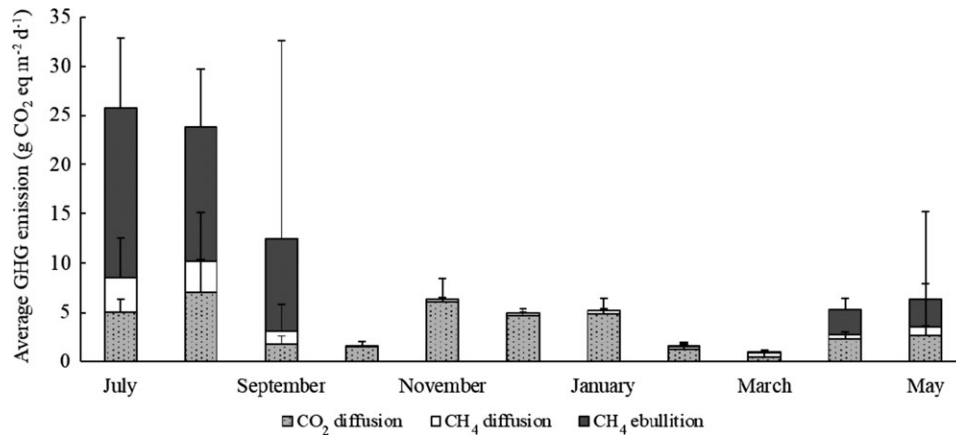


Fig. 1. Measured CO₂ diffusion, CH₄ diffusion, and CH₄ ebullition in CO₂ equivalents (g CO₂ eq m⁻² d⁻¹) ± SD. At some dates, the error bars (SD) are too small to show in the figure.

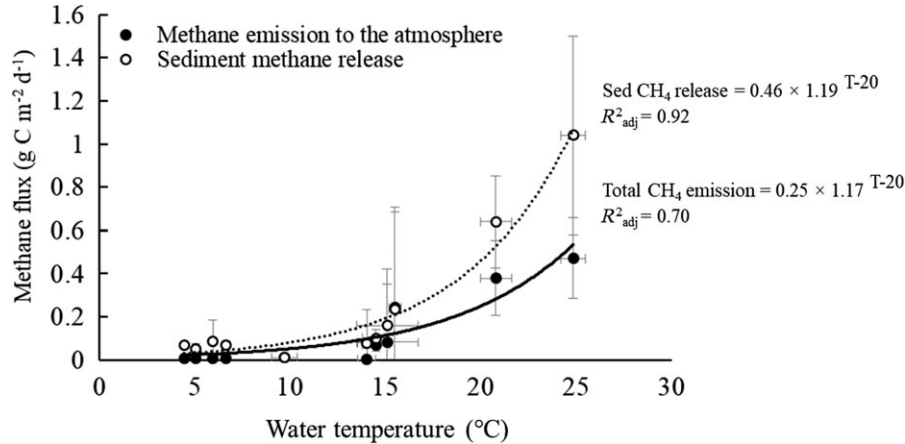


Fig. 2. Relationship between water temperature and CH₄ fluxes in the pond. All average values are shown with their standard deviation, but at some dates, the error bars are too small to show. The solid line shows the relationship of water temperature with the average total CH₄ emission to the atmosphere (black dots), which includes water–atmosphere diffusion and ebullition. The dashed line shows the relationship of water temperature with sediment CH₄ release (white dots), which was calculated as the sum of water–atmosphere CH₄ diffusion and ebullition and water column CH₄ oxidation. Note that CH₄ oxidation rates were not measured in January and May. In these months, we estimated CH₄ oxidation rates based on the relationship between CH₄ diffusion and oxidation (see Materials and Methods section).

before reaching the atmosphere, varied from 0% to 97% over the year (average 57%), with values ranging between 0 and 0.58 g CH₄-C m⁻² d⁻¹. On average, 0.16 g CH₄-C m⁻² d⁻¹ was oxidized. We found a significant location–depth interaction effect (GLMM: Wald $Z = 6.31$; $p < 0.001$): the effect of deep vs. shallow incubation on CH₄ oxidation rates is larger for the littoral than the central zone of the pond. There was also a significant time–location–depth interaction (GLMM: Wald $Z = -15.59$; $p < 0.001$).

Temporal and spatial variation in C fluxes

The CO₂ : CH₄ emission ratio (in CO₂ equivalents) varied strongly over the year (Supporting Information Table S2). In March, the ratio was 1 and there was an equal share of GHG emitted. From April until September, the ratio was well below 1, indicating the importance of CH₄ emission. The ebullitive CH₄ flux

dominated the GHG emission, with a share of 75% of total CH₄ emission and 50% of total GHG emission. From October until February, CO₂ formed the majority of GHG emission, with a share of 80% to 96%. The CH₄ emissions were low during these coldest months (< 0.5 g CO₂ eq m⁻² d⁻¹), and more than 85% of methane released by the sediment was oxidized (Fig. 3). No significant temporal (LMM: $F_{1,10} = 2.87$; $p = 0.12$) and spatial (LMM: $F_{1,10} = 4.09$; $p = 0.07$) difference in diffusive CO₂ flux was found. Similarly, there was no significant temporal (LMM: $F_{1,10} = 3.01$; $p = 0.11$) nor spatial (LMM: $F_{1,10} = 2.61$; $p = 0.14$) difference in diffusive CH₄ emission, although the emission was mostly higher in the central zone (Supporting Information Fig. S1). For ebullition, we found a significant main effect of location (GLMM: Wald $t = 2.18$; $p = 0.029$), with higher ebullition in the center of the pond. In addition, we found a significant

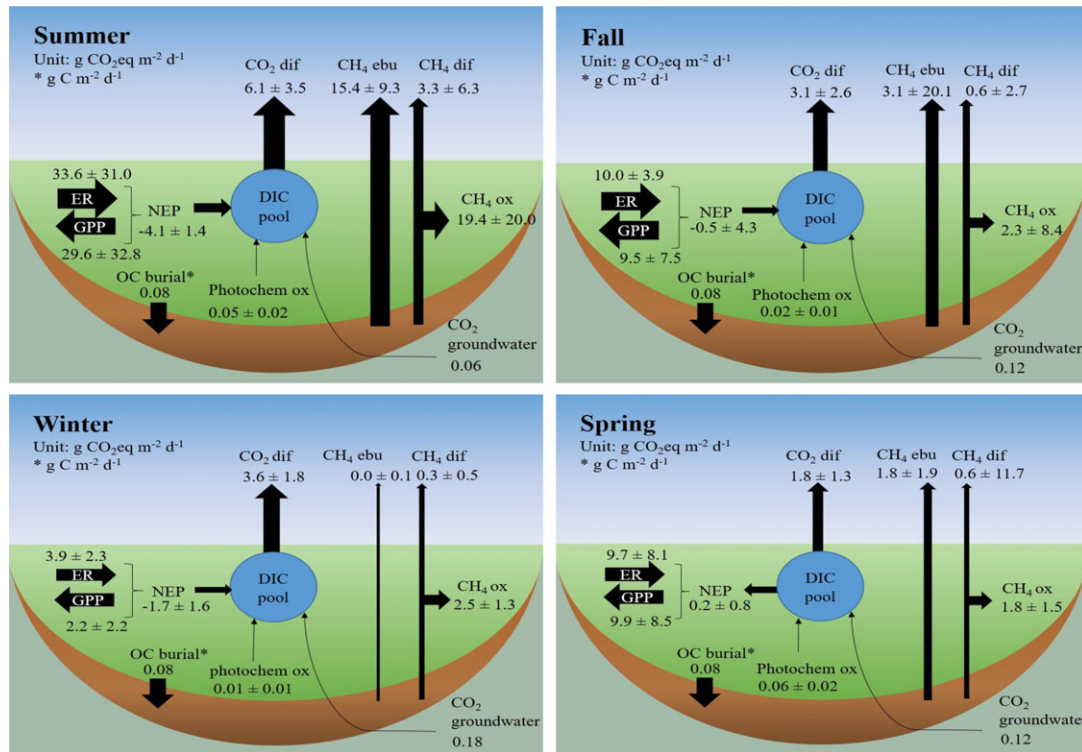


Fig. 3. Seasonal carbon fluxes measured in the pond \pm SD. All values are in $\text{g CO}_2\text{eq m}^{-2}\text{ d}^{-1}$, except for OC burial ($\text{g C m}^{-2}\text{ d}^{-1}$). A negative NEP indicates ER exceeded GPP, thereby adding to the DIC pool. A positive NEP ($\text{GPP} > \text{ER}$) indicates consumption of DIC. Arrows depict the direction of C transport.

interaction (GLMM: Wald $t = 2.66$; $p = 0.008$) between time and location: the effect of the center vs. the littoral zone of the pond was higher for nighttime than for daytime CH_4 ebullition rates.

Relative importance of hydrological inputs and within-pond CO_2 production

High GPP rates were found throughout the year (average of $3.30 \pm 2.79\text{ g CO}_2\text{-C m}^{-2}\text{ d}^{-1}$); however, high rates of ER were observed as well ($3.66 \pm 2.66\text{ g CO}_2\text{-C m}^{-2}\text{ d}^{-1}$). On average, ER rates exceeded GPP rates, which led to net heterotrophy in the system (average NEP of $-0.36 \pm 0.69\text{ g CO}_2\text{-C m}^{-2}\text{ d}^{-1}$). We found net heterotrophy in the system in 8 out of 11 months (NEP of $-0.64 \pm 0.48\text{ g CO}_2\text{-C m}^{-2}\text{ d}^{-1}$) and (minor) net autotrophy in the months February, March, and September (with a respective NEP of 0.02, 0.30, and $1.08\text{ g CO}_2\text{-C m}^{-2}\text{ d}^{-1}$). In months where ER exceeded GPP, the contribution of the NEP to the measured CO_2 emission from the pond was approximately 50% on average. In months with net autotrophy ($\text{GPP} > \text{ER}$), the pond still emitted CO_2 . When CO_2 inputs from groundwater (yearly average of $0.03 \pm 0.01\text{ g C m}^{-2}\text{ d}^{-1}$) and CO_2 formed by photochemical mineralization of DOC (yearly average of $0.009 \pm 0.007\text{ g C m}^{-2}\text{ d}^{-1}$) were added to the NEP in months with net heterotrophy (indicating CO_2 production), the contribution to the measured CO_2 emission was 56%.

Partitioning of ER between organism groups

Empirical relations found to predict organism group-specific respiration (Supporting Information Table S1), based on the relationship between temperature and respiration rates, failed to accurately predict the measured ER of the system (based on the nighttime O_2 decrease at 50 cm depth; Supporting Information Fig. S2). The sum of the modeled respiration of the different organism groups only accounted for roughly half (56%) of the aerobic ER, estimated from the nighttime O_2 curve (Supporting Information Fig. S3). Phytoplankton respiration contributed most to the modeled aerobic ER, with 34.7% on average. After that, the heterotrophic picoplankton and sediment organisms in the lake contributed most to ER with 26.8% and 24.3%, respectively. Fish and methanotrophs contributed the least to ER, with 8.9% and 5.3%, respectively.

Discussion

Our findings demonstrate that the temperate urban pond was a year-round source of GHG, with a particularly large CH_4 emission. The areal GHG emitted by the pond was $3.4\text{ kg CO}_2\text{eq m}^{-2}\text{ yr}^{-1}$, which corresponds to 15.5 t of CO_2 equivalents emitted from the pond surface per year. CH_4 ebullition was by far the most important GHG emission pathway ($\sim 50\%$ of total emission), especially during warm months as we hypothesized ($> 15^\circ\text{C}$, mostly in spring and summer), while

diffusive CH₄ and CO₂ emission contributed with 12% and 38%, respectively. The diffusive CH₄ emission from this small urban pond was three times higher than the average emission reported for small waterbodies within the same size class (on average 0.01 g CH₄ m⁻² d⁻¹, size class 0.001–0.01 km²; Holgerson and Raymond 2016). Moreover, the total CH₄ emission (168 mg CH₄ m⁻² d⁻¹) is moderately high compared to other temperate lakes, irrespective of lake size (Supporting Information Table S3). The OC burial rate corresponded to approximately 6% of the total C emission from the pond (443 g C m⁻² yr⁻¹), which is rather low, as a recent global analyses indicate that on average lakes and reservoir C burial corresponds to roughly 20% of their emissions (Mendonça et al. 2017). Opposed to our hypothesis, the pond did not shift between a net C source in fall and winter to a net C sink in spring and summer. The pond emitted CO₂ to the atmosphere throughout the entire year, even in months where diel oxygen curves indicated the pond was net autotrophic, suggesting that external CO₂ sources other than within-system aerobic respiration play an important role. Overall, the urban pond acted as a “carbon hub,” receiving carbon from different sources, processing, and redirecting the carbon in different ways.

Sediment CH₄ release was strongly related to water temperature and increased exponentially above 15°C (Fig. 2), mainly due to increased ebullition. This result corroborates findings in other systems in various climate regions where temperature-induced exponential increases in ebullition were observed (Aben et al. 2017). Nutrients and organic C limitation may constrain sediment CH₄ release and thereby impede its increase with warming (Kelly and Chynoweth 1981; Schwarz et al. 2008; DelSontro et al. 2016). This is, however, unlikely to occur in hypereutrophic urban ponds, where labile OM availability tends to be high throughout the year both due to internal production and external inflow. The high OM availability and anoxic conditions in pond sediments, combined with the shallow water layer, readily explain the high rates of CH₄ ebullition in our pond (Sobek et al. 2012; Martinez-Cruz et al. 2017). As these conditions are common in urban ponds (Waajen et al. 2014), our results imply that CH₄ emission from these systems is high and strongly temperature dependent. The increase of CH₄ oxidation with rising temperature was much weaker than that of sediment CH₄ release, which resulted in a high summer CH₄ emission (Fig. 2) and is in accordance with a previous laboratory study (Sepulveda-Jauregui et al. 2018). In fall and winter, a high proportion (over 85%) of CH₄ was oxidized (Fig. 3) and emissions were low. CH₄ oxidation rates in our pond were comparable with rates found in other eutrophic systems, which were in a range between 0 and 0.32 g C m⁻² d⁻¹ (Bastviken 2009). Only during July—when water temperature was the highest recorded (25°C)—we found a rate exceeding the reported range (0.58 g C m⁻² d⁻¹). Notably, our estimates do not incorporate oxidation at the sediment–water interface, which can be considerable (Bastviken 2009; Martinez-Cruz et al. 2018).

Diffusive CO₂ emission was, after CH₄ ebullition, the most important contributor to the pond’s total GHG emission. The internally produced CO₂ (NEP), calculated based on diel O₂ curves, was always lower than the measured CO₂ emission rates (comprising on average ~ 50% of the emission rate). Even during phytoplankton blooms in summer, when GPP exceeded ER, CO₂ was emitted to the atmosphere (6.1 ± 3.5 g CO₂-C m⁻² d⁻¹). Daytime and nighttime CO₂ emissions were not statistically different. This indicates that the pond’s atmospheric CO₂ exchange was not strongly regulated by primary production within the lake, opposed to our hypothesis and to findings of Pacheco et al. (2013). CO₂ input from groundwater and photochemical mineralization of DOC was responsible for approximately 6% of the measured CO₂ emission. The unexplained fraction of the CO₂ emission likely originates from endogenous anaerobic CO₂ production or exogenously produced CO₂ (entering via runoff or local groundwater originating from e.g., leaves from trees, street dirt, or bird droppings). Endogenous anaerobic CO₂ may arise from denitrification, which seems likely as Dutch groundwater often contains a large amount of nitrate (Smolders et al. 2006). Allochthonous C is known to drive CO₂ emission in small and shallow ponds, owing to their high perimeter to volume ratio (Finlay et al. 2010; Holgerson 2015; Wilkinson et al. 2016). Other possible factors that may explain the discrepancy between the measured CO₂ emission and estimated internal CO₂ production include inaccurate estimates of oxygen saturation and gas transfer velocity, as well as a possible underestimation of daytime respiration (Del Giorgio and Williams 2005; Staehr et al. 2010; Brothers et al. 2017). Estimating respiration rates per organism group is a difficult practice as well and comes with several uncertainties and caveats (Del Giorgio and Williams 2005; Martínez-García et al. 2013), which was confirmed by the fact that our modeled ER only accounted for roughly half of the oxygen diel-curve-derived ER.

Seasonal trends in GHG emissions from temperate ponds have generally been disregarded based on the observation that GHG emissions in these systems are often not primarily driven by the seasonally varying within-system metabolism (Jonsson et al. 2003; Holgerson 2015). Although in our pond NEP played a minor role in driving C emission as well, we did find a clear seasonal pattern in GHG emissions (Figs. 1, 3). This seasonal trend is strongly determined by ebullition (Fig. 1; Supporting Information Table S2). We found a lower CO₂ : CH₄ emission ratio with increasing temperature (Supporting Information Table S2), largely due to an increase in CH₄ emission, in accordance with other studies (Yvon-Durocher et al. 2014; DelSontro et al. 2016). Unexpectedly, we did not find clear differences in spatial and temporal (daytime or nighttime) diffusive GHG emission. Both, CH₄ ebullition and diffusion were, however, clearly related to temperature and ebullition was significantly higher in the central zone of the pond. The latter contradicts other studies that suggest the littoral zone plays a more important role in GHG emissions (Natchimuthu et al. 2016). Because of the high spatiotemporal variation in total GHG emission and the high importance of CH₄

ebullition, we recommend future studies to explicitly incorporate ebullition measurements and to take spatial differences into account even in small systems, such as ponds. Furthermore, future monitoring efforts in comparable temperate and eutrophic systems should be concentrated in the months May through September, as in these months the highest fluxes and highest variation occurs. A low number of measurements in fall and winter can be used to estimate cold season emissions.

Extrapolating our findings to all urban ponds in the Netherlands (extrapolated from data by Waajen et al. [2014]), we find a GHG emission of 0.27 Mt CO₂ eq yr⁻¹. This equals the annual emission of approximately 193,000 cars (based on driving 13,200 km yr⁻¹). City planners consider ponds as natural cooling mechanisms to mitigate urban heat stress (Inard et al. 2004; Theeuwes et al. 2013); however, our findings indicate that this is not a GHG neutral option. In addition, hypereutrophic urban ponds are more likely to be affected by raising temperatures, as water temperatures of small and shallow water systems are closely correlated to air temperature, making them sensitive to rapid warming (Toffolon et al. 2014; Wik et al. 2014). Shallow, urban ponds with a high OM input may thus likely exhibit enhanced CH₄ emissions with global warming induced rising of water temperatures (O'Reilly et al. 2015). Considering their already high GHG emission rates, their inclusion in regional GHG emission assessments is important. Climate change will likely feedback on urban ponds GHG emissions and additionally worsen eutrophication and OM loading to the system (Moss et al. 2011). In line with the idea of using local management strategies to mitigate global stressors (Scheffer et al. 2015), management strategies for urban ponds should be aimed at reducing eutrophication and OM loading. In this way, local management may counteract the warming induced increase of GHG emissions as well.

References

- Aben, R. C., and others. 2017. Cross continental increase in methane ebullition under climate change. *Nat. Commun.* **8**: 1682. doi:10.1038/s41467-017-01535-y
- Algesten, G., S. Sobek, A. K. Bergström, A. Ågren, L. J. Tranvik, and M. Jansson. 2003. Role of lakes for organic carbon cycling in the boreal zone. *Glob. Chang. Biol.* **10**: 141–147. doi:10.1046/j.1529-8817.2003.00721.x
- Almeida, R. M., and others. 2016. High primary production contrasts with intense carbon emission in a eutrophic tropical reservoir. *Front. Microbiol.* **7**: 717. doi:10.3389/fmicb.2016.00717
- Balmer, M. B., and J. A. Downing. 2011. Carbon dioxide concentrations in eutrophic lakes: Undersaturation implies atmospheric uptake. *Inland Waters* **1**: 125–132. doi:10.5268/IW-1.2.366
- Bastviken, D. 2009. Methane. *In* G. E. Likens [ed.], *Encyclopedia of inland waters*. Elsevier.
- Bastviken, D., J. J. Cole, M. L. Pace, and M. C. Van de Bogert. 2008. Fates of methane from different lake habitats: Connecting whole-lake budgets and CH₄ emissions. *J. Geophys. Res. Biogeosci.* **113**: G02024. doi:10.1029/2007JG000608
- Bastviken, D., L. J. Tranvik, J. A. Downing, P. M. Crill, and A. Enrich-Prast. 2011. Freshwater methane emissions offset the continental carbon sink. *Science* **331**: 50. doi:10.1126/science.1196808
- Bates, D., M. Mächler, B. Bolker, and S. Walker. 2015. Fitting linear mixed-effects models using lme4. *J. Stat. Softw.* **67**: 1–48. doi:10.18637/jss.v067.i01.
- Bolund, P., and S. Hunhammar. 1999. Ecosystem services in urban areas. *Ecol. Econ.* **29**: 293–301. doi:10.1016/S0921-8009(99)00013-0
- Brothers, S., G. Kazanjian, J. Köhler, U. Scharfenberger, and S. Hilt. 2017. Convective mixing and high littoral production established systematic errors in the diel oxygen curves of a shallow, eutrophic lake. *Limnol. Oceanogr.: Methods* **15**: 429–435. doi:10.1002/lom3.10169
- Casper, P., S. C. Maberly, G. H. Hall, and B. J. Finlay. 2000. Fluxes of methane and carbon dioxide from a small productive lake to the atmosphere. *Biogeochemistry* **49**: 1–19. doi:10.1023/A:1006269900174
- Champion, T. 2001. Urbanization, suburbanization, counterurbanization and reurbanization, p. 143–161. *In* R. Paddison [ed.], *Handbook of urban studies*. London: Sage.
- Cole, J. J., N. F. Caraco, G. W. Kling, and T. K. Kratz. 1994. Carbon dioxide supersaturation in the surface waters of lakes. *Science* **265**: 1568–1570. doi:10.1126/science.265.5178.1568
- Cole, J. J., M. L. Pace, S. R. Carpenter, and J. F. Kitchell. 2000. Persistence of net heterotrophy in lakes during nutrient addition and food web manipulations. *Limnol. Oceanogr.* **45**: 1718–1730. doi:10.4319/lo.2000.45.8.1718
- Dean, W. E., and E. Gorham. 1998. Magnitude and significance of carbon burial in lakes, reservoirs, and peatlands. *Geology* **26**: 535–538. doi:10.1130/0091-7613(1998)026<0535:MASOCB>2.3.CO;2
- Del Giorgio, P. A., and P. J. I. B. Williams. 2005. *Respiration in aquatic ecosystems*. Oxford Univ. Press.
- DelSontro, T., L. Boutet, A. St-Pierre, P. A. del Giorgio, and Y. T. Prairie. 2016. Methane ebullition and diffusion from northern ponds and lakes regulated by the interaction between temperature and system productivity. *Limnol. Oceanogr.* **61**: S62–S77. doi:10.1002/lno.10335
- Downing, J. A. 2010. Emerging global role of small lakes and ponds: Little things mean a lot. *Limnetica* **29**: 9–24.
- Downing, J. A., and others. 2006. The global abundance and size distribution of lakes, ponds, and impoundments. *Limnol. Oceanogr.* **51**: 2388–2397. doi:10.4319/lo.2006.51.5.2388
- Finlay, K., P. Leavitt, A. Patoine, A. Patoine, and B. Wissel. 2010. Magnitudes and controls of organic and inorganic carbon flux through a chain of hard-water lakes on the

- northern Great Plains. *Limnol. Oceanogr.* **55**: 1551–1564. doi:[10.4319/lo.2010.55.4.1551](https://doi.org/10.4319/lo.2010.55.4.1551)
- Gledhill, D. G., P. James, and D. H. Davies. 2008. Pond density as a determinant of aquatic species richness in an urban landscape. *Landsc. Ecol.* **23**: 1219–1230. doi:[10.1007/s10980-008-9292-x](https://doi.org/10.1007/s10980-008-9292-x)
- Grasshoff, K., and H. Johannsen. 1972. A new sensitive and direct method for the automatic determination of ammonia in sea water. *ICES J. Mar. Sci.* **34**: 516–521. doi:[10.1093/icesjms/34.3.516](https://doi.org/10.1093/icesjms/34.3.516)
- Hanson, P. C., S. R. Carpenter, J. A. Cardille, M. T. Coe, and L. A. Winslow. 2007. Small lakes dominate a random sample of regional lake characteristics. *Freshwater Biol.* **52**: 814–822. doi:[10.1111/j.1365-2427.2007.01730.x](https://doi.org/10.1111/j.1365-2427.2007.01730.x)
- Holgerson, M. A. 2015. Drivers of carbon dioxide and methane supersaturation in small, temporary ponds. *Biogeochemistry* **124**: 305–318. doi:[10.1007/s10533-015-0099-y](https://doi.org/10.1007/s10533-015-0099-y)
- Holgerson, M. A., and P. A. Raymond. 2016. Large contribution to inland water CO₂ and CH₄ emissions from very small ponds. *Nat. Geosci.* **9**: 222–226. doi:[10.1038/NNGEO2654](https://doi.org/10.1038/NNGEO2654)
- Inard, C., D. Groleau, and M. Musy. 2004. Energy balance study of water ponds and its influence on building energy consumption. *Build. Serv. Eng. Res. Technol.* **25**: 171–182. doi:[10.1191/0143624404bt1060a](https://doi.org/10.1191/0143624404bt1060a)
- Jonsson, A., J. Karlsson, and M. Jansson. 2003. Sources of carbon dioxide supersaturation in clearwater and humic lakes in northern Sweden. *Ecosystems* **6**: 224–235. doi:[10.1007/s10021-002-0200-y](https://doi.org/10.1007/s10021-002-0200-y)
- Kamphake, L., S. Hannah, and J. Cohen. 1967. Automated analysis for nitrate by hydrazine reduction. *Water Res.* **1**: 205–216. doi:[10.1016/0043-1354\(67\)90011-5](https://doi.org/10.1016/0043-1354(67)90011-5)
- Kelly, C. A., and D. P. Chynoweth. 1981. The contributions of temperature and of the input of organic matter in controlling rates of sediment methanogenesis. *Limnol. Oceanogr.* **26**: 891–897. doi:[10.4319/lo.1981.26.5.0891](https://doi.org/10.4319/lo.1981.26.5.0891)
- Kosten, S., F. Roland, D. M. L. Da Motta Marques, E. H. Van Nes, N. M. L. d. S. L. Sternberg, M. Scheffer, and J. J. Cole. 2010. Climate-dependent CO₂ emissions from lakes. *Global Biogeochem. Cycles* **24**: 24. doi:[10.1029/2009GB003618](https://doi.org/10.1029/2009GB003618)
- Martinez-Cruz, K., R. Gonzalez-Valencia, A. Sepulveda-Jauregui, F. Plascencia-Hernandez, Y. Belmonte-Izquierdo, and F. Thalasso. 2017. Methane emission from aquatic ecosystems of Mexico City. *Aquat. Sci.* **79**: 159–169. doi:[10.1007/s00027-016-0487-y](https://doi.org/10.1007/s00027-016-0487-y)
- Martinez-Cruz, K., A. Sepulveda-Jauregui, P. Casper, K. W. Anthony, K. A. Smemo, and F. Thalasso. 2018. Ubiquitous and significant anaerobic oxidation of methane in freshwater lake sediments. *Water Res.* **144**: 332–340. doi:[10.1016/j.watres.2018.07.053](https://doi.org/10.1016/j.watres.2018.07.053)
- Martínez-García, S., E. Fernández, D. A. del Valle, D. M. Karl, and E. Teira. 2013. Experimental assessment of marine bacterial respiration. *Aquat. Microb. Ecol.* **70**: 189–205. doi:[10.3354/ame01644](https://doi.org/10.3354/ame01644)
- Mendonça, R., S. Kosten, S. Sobek, J. J. Cole, A. C. Bastos, A. L. Albuquerque, S. J. Cardoso, and F. Roland. 2014. Carbon sequestration in a large hydroelectric reservoir: An integrative seismic approach. *Ecosystems* **17**: 430–441. doi:[10.1007/s10021-013-9735-3](https://doi.org/10.1007/s10021-013-9735-3)
- Mendonça, R., R. A. Müller, D. Clow, C. Verpoorter, P. Raymond, L. J. Tranvik, and S. Sobek. 2017. Organic carbon burial in global lakes and reservoirs. *Nat. Commun.* **8**: 1694. doi:[10.1038/s41467-017-01789-6](https://doi.org/10.1038/s41467-017-01789-6)
- Moss, B., and others. 2011. Allied attack: Climate change and eutrophication. *Inland Waters* **1**: 101–105. doi:[10.5268/IW-1.2.359](https://doi.org/10.5268/IW-1.2.359)
- Myhre, G., and others. 2013. Anthropogenic and natural radiative forcing, p. 659–740. *In* T. F. Stocker et al. [eds.], *Climate change 2013: The physical science basis. Contribution of Working Group I to the fifth assessment report of the intergovernmental panel on climate change*. Cambridge Univ. press.
- Natchimuthu, S., I. Sundgren, M. Gålfalk, L. Klemedtsson, P. Crill, Å. Danielsson, and D. Bastviken. 2016. Spatio-temporal variability of lake CH₄ fluxes and its influence on annual whole lake emission estimates. *Limnol. Oceanogr.* **61**: S13–S26. doi:[10.1002/lno.10222](https://doi.org/10.1002/lno.10222)
- Novotny, V. 1999. Integrating diffuse-nonpoint pollution control and water body restoration into watershed management. *J. Am. Water Resour. Assoc.* **35**: 717–727. doi:[10.1111/j.1752-1688.1999.tb04169.x](https://doi.org/10.1111/j.1752-1688.1999.tb04169.x)
- O'Reilly, C. M., and others. 2015. Rapid and highly variable warming of lake surface waters around the globe. *Geophys. Res. Lett.* **42**: 10,773–710,781. doi:[10.1002/2015GL066235](https://doi.org/10.1002/2015GL066235)
- Pacheco, F. S., F. Roland, and J. A. Downing. 2013. Eutrophication reverses whole-lake carbon budgets. *Inland Waters* **4**: 41–48. doi:[10.5268/IW-4.1.614](https://doi.org/10.5268/IW-4.1.614)
- Raymond, P. A., and others. 2013. Global carbon dioxide emissions from inland waters. *Nature* **503**: 355–387. doi:[10.1038/nature12760](https://doi.org/10.1038/nature12760)
- Roland, F., L. O. Vidal, F. S. Pacheco, N. O. Barros, A. Assireu, J. P. H. B. Ometto, A. C. P. Cimleris, and J. J. Cole. 2010. Variability of carbon dioxide flux from tropical (Cerrado) hydroelectric reservoirs. *Aquat. Sci.* **72**: 283–293. doi:[10.1007/s00027-010-0140-0](https://doi.org/10.1007/s00027-010-0140-0)
- Sand-Jensen, K., and P. A. Staehr. 2009. Net heterotrophy in small Danish lakes: A widespread feature over gradients in trophic status and land cover. *Ecosystems* **12**: 336–348. doi:[10.1007/s10021-008-9226-0](https://doi.org/10.1007/s10021-008-9226-0)
- Scheffer, M., and others. 2015. Creating a safe operating space for iconic ecosystems. *Science* **347**: 1317–1319. doi:[10.1126/science.aaa3769](https://doi.org/10.1126/science.aaa3769)
- Scherer, N. M., H. L. Gibbons, K. B. Stoops, and M. Muller. 1995. Phosphorus loading of an urban lake by bird droppings. *Lake Reservoir Manage.* **11**: 317–327. doi:[10.1080/07438149509354213](https://doi.org/10.1080/07438149509354213)
- Schwarz, J. I., W. Eckert, and R. Conrad. 2008. Response of the methanogenic microbial community of a profundal Lake

- sediment (Lake Kinneret, Israel) to algal deposition. *Limnol. Oceanogr.* **53**: 113–121. doi:[10.4319/lo.2008.53.1.0113](https://doi.org/10.4319/lo.2008.53.1.0113)
- Sepulveda-Jauregui, A., J. Hoyos-Santillan, K. Martinez-Cruz, K. M. Walter Anthony, P. Casper, Y. Belmonte-Izquierdo, and F. Thalasso. 2018. Eutrophication exacerbates the impact of climate warming on lake methane emission. *Sci. Total Environ.* **636**: 411–419. doi:[10.1016/j.scitotenv.2018.04.283](https://doi.org/10.1016/j.scitotenv.2018.04.283)
- Smith, V. H. 1979. Nutrient dependence of primary productivity in lakes. *Limnol. Oceanogr.* **24**: 1051–1064. doi:[10.4319/lo.1979.24.6.1051](https://doi.org/10.4319/lo.1979.24.6.1051)
- Smolders, A., L. Lamers, E. Lucassen, G. Van der Velde, and J. Roelofs. 2006. Internal eutrophication: How it works and what to do about it—a review. *Chem. Ecol.* **22**: 93–111. doi:[10.1080/02757540600579730](https://doi.org/10.1080/02757540600579730)
- Sobek, S., T. DelSontro, N. Wongfun, and B. Wehrli. 2012. Extreme organic carbon burial fuels intense methane bubbling in a temperate reservoir. *Geophys. Res. Lett.* **39**: L01401. doi:[10.1029/2011GL050144](https://doi.org/10.1029/2011GL050144)
- Staehr, P. A., D. Bade, M. C. Van de Bogert, G. R. Koch, C. Williamson, P. Hanson, J. J. Cole, and T. Kratz. 2010. Lake metabolism and the diel oxygen technique: State of the science. *Limnol. Oceanogr.: Methods* **8**: 628–644. doi:[10.4319/lom.2010.8.628](https://doi.org/10.4319/lom.2010.8.628)
- Theeuwes, N., A. Solcerová, and G. Steeneveld. 2013. Modeling the influence of open water surfaces on the summertime temperature and thermal comfort in the city. *J. Geophys. Res.: Atmos.* **118**: 8881–8896. doi:[10.1002/jgrd.50704](https://doi.org/10.1002/jgrd.50704)
- Toffolon, M., S. Piccolroaz, B. Majone, A.-. M. Soja, F. Peeters, M. Schmid, and A. Wüest. 2014. Prediction of surface temperature in lakes with different morphology using air temperature. *Limnol. Oceanography* **59**: 2185–2202. doi:[10.4319/lo.2014.59.6.2185](https://doi.org/10.4319/lo.2014.59.6.2185)
- Tranvik, L. J., and others. 2009. Lakes and reservoirs as regulators of carbon cycling and climate. *Limnol. Oceanogr.* **54**: 2298–2314. doi:[10.4319/lo.2009.54.6_part_2.2298](https://doi.org/10.4319/lo.2009.54.6_part_2.2298)
- Trolle, D., P. A. Staehr, T. A. Davidson, R. Bjerring, T. L. Lauridsen, M. Søndergaard, and E. Jeppesen. 2012. Seasonal dynamics of CO₂ flux across the surface of shallow temperate lakes. *Ecosystems* **15**: 336–347. doi:[10.1007/s10021-011-9513-z](https://doi.org/10.1007/s10021-011-9513-z)
- Vähätalo, A. V., M. Salkinoja-Salonen, P. Taalas, and K. Salonen. 2000. Spectrum of the quantum yield for photochemical mineralization of dissolved organic carbon in a humic lake. *Limnol. Oceanogr.* **45**: 664–676. doi:[10.4319/lo.2000.45.3.0664](https://doi.org/10.4319/lo.2000.45.3.0664)
- Verpoorter, C., T. Kutser, D. A. Seekell, and L. J. Tranvik. 2014. A global inventory of lakes based on high-resolution satellite imagery. *Geophys. Res. Lett.* **41**: 6396–6402. doi:[10.1002/2014GL060641](https://doi.org/10.1002/2014GL060641)
- Waajen, G. W., E. J. Faassen, and M. Lüring. 2014. Eutrophic urban ponds suffer from cyanobacterial blooms: Dutch examples. *Environ. Sci. Pollut. Res.* **21**: 9983–9994. doi:[10.1007/s11356-014-2948-y](https://doi.org/10.1007/s11356-014-2948-y)
- Waschbusch, R. J., W. R. Selbig, and R. T. Bannerman. 1999. Sources of phosphorus in stormwater and street dirt from two urban residential basins in Madison, Wisconsin, 1994–95, p. 51. Water-Resources Investigations Report.
- West, W. E., J. J. Coloso, and S. E. Jones. 2012. Effects of algal and terrestrial carbon on methane production rates and methanogen community structure in a temperate lake sediment. *Freshwater Biol.* **57**: 949–955. doi:[10.1111/j.1365-2427.2012.02755.x](https://doi.org/10.1111/j.1365-2427.2012.02755.x)
- West, W. E., K. P. Creamer, and S. E. Jones. 2016. Productivity and depth regulate lake contributions to atmospheric methane. *Limnol. Oceanogr.* **61**: S51–S61. doi:[10.1002/lno.10247](https://doi.org/10.1002/lno.10247)
- Weyhenmeyer, G. A., S. Kosten, M. B. Wallin, L. J. Tranvik, E. Jeppesen, and F. Roland. 2015. Significant fraction of CO₂ emissions from boreal lakes derived from hydrologic inorganic carbon inputs. *Nat. Geosci.* **8**: 933–936. doi:[10.1038/ngeo2582](https://doi.org/10.1038/ngeo2582)
- Wik, M., B. F. Thornton, D. Bastviken, S. MacIntyre, R. K. Varner, and P. M. Crill. 2014. Energy input is primary controller of methane bubbling in subarctic lakes. *Geophys. Res. Lett.* **41**: 555–560. doi:[10.1002/2013GL058510](https://doi.org/10.1002/2013GL058510)
- Wik, M., R. K. Varner, K. W. Anthony, S. MacIntyre, and D. Bastviken. 2016. Climate-sensitive northern lakes and ponds are critical components of methane release. *Nat. Geosci.* **9**: 99–105. doi:[10.1038/NNGEO2578](https://doi.org/10.1038/NNGEO2578)
- Wilkinson, G. M., C. D. Buelo, J. J. Cole, and M. L. Pace. 2016. Exogenously produced CO₂ doubles the CO₂ efflux from three north temperate lakes. *Geophys. Res. Lett.* **43**: 1996–2003. doi:[10.1002/2016GL067732](https://doi.org/10.1002/2016GL067732)
- Wong, T., P. F. Breen, and S. D. Lloyd. 2000. Water sensitive road design: Design options for improving stormwater quality of road runoff. Cooperative Center for Catchment Hydrology.
- Yvon-Durocher, G., A. P. Allen, D. Bastviken, R. Conrad, C. Gudasz, A. St-Pierre, N. Thanh-Duc, and P. A. del Giorgio. 2014. Methane fluxes show consistent temperature dependence across microbial to ecosystem scales. *Nature* **507**: 488–491. doi:[10.1038/nature13164](https://doi.org/10.1038/nature13164)

Acknowledgments

We thank K. F. Ettwig for her advice and discussions. We thank Paul van de Ven, Germa Verheggen, Stefan Weideveld, Haijung Wang, Maarten Schimmel, Carlos Henrique Duque Estrada and Felipe Rust for their assistance during the experiment and for helping with analyses. We also thank Jan Schaart (HSV Heumen) for providing an estimation of fish biomass and species in the pond and for logistic support on site. Additionally, we thank Roel van Veen from water authority Rivierenland for providing us groundwater seepage estimates and Simone Cardoso for advice on the statistical analyses. This study was conducted under the auspices of the CAPES (Brazil)/NUFFIC (The Netherlands), project 004/2008. N.B. was supported by Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES) process number 3934-13-6 and CAPES (Brazil)/NUFFIC (The Netherlands) project 045/2012; S.K. was supported by NWO-VENI (grant 86312012); and V.H. was partially supported by Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq, Brazil).

Conflict of Interest

None declared

majority of the manuscript writing. T.J.H.M.v.B., N.B., R.M., R.A., I.H. J.A., V.H., L.P.M.L., M.L., F.R. and S.K. contributed to revisions of the manuscript.

Author Contribution Statement

V.H., M.L., and S.K. initiated the overarching project. N.B., R.M. and S.K. initiated the study present in the paper. N.B., R.M., T.J.H.M. v.B and I.H.J.A. conducted the fieldwork. T.J.H.M.v.B and N.B. analysed the data. I.H.J.A. and N.B. conducted the sediment incubation. R.A. contributed to the statistical analyses. T.J.H.M.v.B. did the

Submitted 29 July 2018

Revised 31 January 2019

Accepted 14 March 2019

Associate editor: Kimberly Wickland