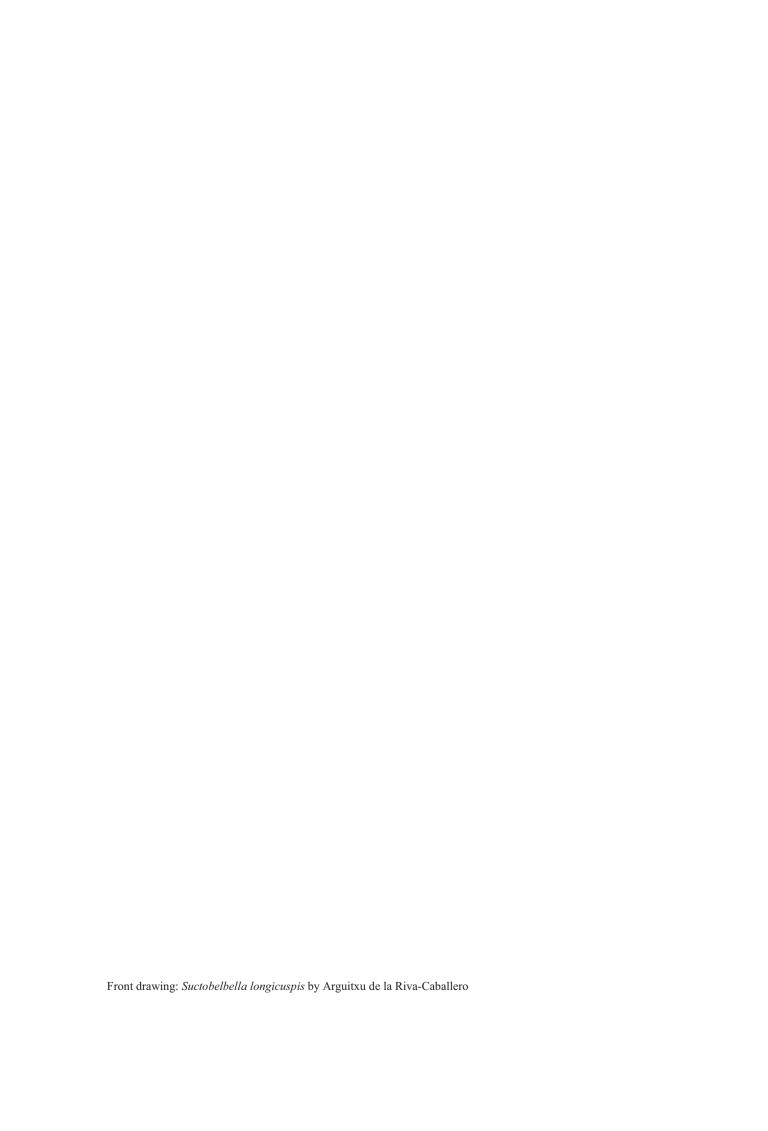
Oribatid mites in a changing world

Arguitxu de la Riva Caballero



Dissertation for the degree of philosophiae doctor (PhD)
University of Bergen, Norway
2011





To Adriana and Matteo

Supervisors:

Main supervisor: Torstein Solhøy

Co-supervisor: Hilary H. Birks

Co-supervisor: Lita G. Jensen

Contents

Abstract	3
Acknowledgments	5
List of papers	7
Declaration	9
Synthesis Oribatid mites in a changing world	
Introduction	11
Archives and Proxies	11
What are oribatid mites?	13
Oribatids in palaeoecology	15
Aims of this thesis	18
Study Areas	19
Methods	20
Field sampling	21
Laboratory methods	22
Data analysis	23
Outline of main results in the four papers	24
Background fauna characterisation	
Oribatid mite representation in lake sediments and best coring point	
Palaeolimnological and archaeological reconstructions using oribatid mites	27 ke in fossils
Paper IV: Neolithic impact on local vegetation at Kvitevoll, Halsnøy Island, western Norway reconstructed from oribatid mites and pollen analysis	
Conclusions, problems, and further work	31
Acknowledgements	32
Defenence	22

Oribatid Mites in a Changing World

De la Riva-Caballero, A. 2011. Oribatid Mites in a Changing World. PhD thesis, Bergen Museum and Faculty of Mathematics and Natural Sciences, University of Bergen, Norway

Abstract

The main scope of this thesis is to illustrate the validity of oribatid mites as for palaeoecological tools reconstructions. Palaeoecology studies the responses of past organisms to past environmental changes. This can be accomplished through the use of biological proxies, which are indicators of past conditions. The search for additional means of distinguishing climate change has only recently led to the use of other commonly found biological proxies such as tiny oribatid mites known as moss-mites. Oribatid mites are among the most numerous biological remains in anoxic sediments, yet until now oribatids have not been widely used due to the uncertainties about their present distribution and the lack of expertise to identify them to species level. This thesis contains four papers which provide evidence about how oribatid mites, when they are properly identified to species level and their background distribution adequately known, can give useful additional and supporting information for reconstructing past habitat and environmental conditions.

Paper I studied oribatid preferences and ecology in different habitats, mainly forested, in western Norway. One hundred and ninety two species were found of which 64 were

new records for Norway. The species Chamobates borealis, Oppiella nova, *Moritzoppia* neerlandica, and Rhinoppia subpectinata characterised the oribatid communities of Betula, mixed, and Picea forest subsets. Deciduous forest oribatid communities characterised by Achipteria coleoptrata, Acrotritria ardua. Ceratozetes gracilis, and Oribatella calcarata. Hemileius initialis. Nanhermannia dorsalis, C. borealis, Tectocepheus velatus, and Atropacarus striculus characterised wet habitats. In water-logged habitats, Limnozetes ciliatus, Mucronothrus nasalis, Trimalaconothrus glaber dominated. Carabodes labyrinthicus, C. marginatus, Melanozetes mollicomus, and T. velatus characterised the oribatid community of the lichen and moss subset. The tree-line ecotone was dominated by the euryceous species H. T. velatus, and Oribatula initialis, This tibialis. study represents survey of oribatid thorough communities in western Norway, and the insights it gives are an important tool for habitat reconstructions, as they provide the background knowledge about modern oribatid fauna needed to identify the type of past community and past environments represented in Quaternary sediments.

Paper II studied the oribatid communities at the tree-line in western Norway and compared them with the oribatid fossil assemblages found in Lake Trettetjørn. The modern oribatid assemblage provided a guide to the reliability of the fossil assemblages to reconstruct ecological environmental changes and, in addition, to find the most favourable coring point within the small lake. Results showed that the core retrieved from the middle of Lake Trettetjørn basin represented the oribatid fauna from the catchment area. Aquatic oribatids were the best group represented in the lake sediments, followed by oribatids from the habitats adjacent to the lake. This constitutes good evidence that oribatids excellent indicators of local habitats. Comparison of the oribatid fauna found in the lake traps with the oribatid assemblages from paper III illustrated the importance of identifying the mites to species level, as this increased the ecological indicator value and. reliability therefore, the of the palaeoreconstructions.

In Paper III, sub-fossil oribatid mites, pollen, plant macrofossils, and diatoms from a lake sediment core from western Norway were studied. This multi-proxy study attempted reconstruct tree-line fluctuations and their impact on Lake Trettetjørn's environment. Evidence from pollen, macrofossils, and oribatids complemented and corroborated each other in the reconstruction of the vegetational development. A semi-open grassland developed into forest. Mires began to replace forested areas on the landscape as a more oceanic climate began to prevail. All proxies indicated increasingly intensive human land-use as the Upsete settlement grew to accommodate the construction of the Bergen-Oslo railway.

In Paper IV, oribatid mites and pollen were used to reconstruct the local habitat at an archaeological excavation. The study aimed to identify the start of cereal cultivation at Kvitevoll farm, on Halsnøy island, western Norway. The number of oribatid remains identified to species level and the close match to pollen stratigraphy led to a palaeoenvironmental detailed reconstruction. Oribatids and pollen indicated the development of a moist forest followed by vegetation openings and mire expansion over the site. At the top of the sequence, the presence of oribatids such as Tectocepheus velatus and the increase in members of the family Oppiidae indicated a higher degree of disturbance, probably from grazing. Pollen of Cerealia indicated the start of cultivation around the same time.

Acknowledgments

It is difficult to thank all the people who have helped me with this thesis. I am certain I will forget mentioning some, but please do not be offended. I am grateful to all of you.

Torstein Solhøy deserves a special thanks, not only because he has been my supervisor, but also because he was the one who introduced me to oribatid mites and gave me the chance to fall in love with them during that first summer of 2001. He is an extraordinary person in many ways. He knows when he is really needed, and then he is there. John Birks also especially deserves to be thanked. He has helped me through the process, and not only with the numerical analyses!

Hilary Birks has contributed, especially in the final steps, to make this thesis a reality. She is the perfectionist I needed at my side to show me how to do things properly. Lita G. Jensen, my supervisor at Bergen Museum, has always been there. Thank you all!!

In addition, I would like to thank Jorunn Larsen, Anne E. Bjune, and Lene S. Halvorsen, all fellow authors in one of the papers comprised in this thesis, for their encouragement and support through the process. Working with you has been wonderful.

My colleagues at the Department of Biology: Marianne P. Heggen, Ingelinn Aarnes, Kristine Fjordheim, Linn C. Kruger, Bjørn Arild Hatteland, and Arild Breistøl. It has been great to share so many coffee breaks, conversations about theses and other matters, parties, etc., with you. Your support through the process has been especially invaluable.

Marianne P. Heggen through the years became not only the best colleague one could dream of, but also a good friend. She has been encouraging, enlightening, patient, funny and serious, helping me any time I needed it. Thanks Marianne! I look forward to the next time we share an office!

I am in debt to Anna Seniczak, with whom I shared my first summer working with mites at Acarilaben. She is an exceptional teacher and was able to make the task of learning to identify those wonderful but tricky creatures funny, challenging, and fascinating. Definitively, I could not have started my PhD thesis without that first summer.

Luis S. Subías has been an invaluable help. He was able to pass on his love for oribatid mites, especially for oppiids, and I suddenly discovered that I was completely fascinated by those golden little creatures. He has shared knowledge about not only mites but also life in our coffee breaks and meetings whenever I visited Madrid, and he patiently listened to my thesis or everyday life problems and tried to help me out. In other words, he became not only a guide in the oribatid world but also a friend. Thank you Luis!!

At Bergen Museum I would especially like to thank Ingrid Herø, who is now retired. She has been the most effective and helpful woman in administration that I have ever met. Moreover, she was always in a good mood and smiling regardless of the "strange" situations I came to explain in my funny Norwegian.

The last year and a half I have been living in Milan. I do not have working colleagues there, but I have a wonderful mother-in-law, mamma Vale, who has made my everyday work a pleasure. She has not only invited me into her house, but also cooked for me! She has been my coffee break "colleague" and has listened patiently to all of my achievements and complaints. In addition, she has always helped at home not only with cooking and cleaning but also, and most importantly, by playing with Adriana. Thank you very much Vale!

Thank you to Hanneke, Usman, Danyal, and Ishaq, unconditional friends despite the distance. They are our Norwegian family, and they always made our weekends something to look forward to, always supported me, and have offered not only their house whenever I was in Bergen during this last year and a half, but also made me feel like one of their family.

Susi, thank you very much, for being more than a friend, no matter the distance, for your enthusiasm, your good mood, the coffees, the laughs, dinners, lunches, vacations, and your help when needed.

My family, mamá, papá, Pilar, Fernando, María, Isabel, and Txema, though not always close physically, are always close to me. Their support has been crucial through this process. They have always understood me and guided me. Thank you very much! You are probably the strangest but the best family in the world. I love you all.

And last but not least I am extremely grateful to my husband, Matteo, and my daughter, Adriana. Matteo is, without a doubt, my better half. He understands me and supports me unconditionally. Adriana is still too young to understand what mummy was doing but her laughs, her first words, her first steps, her hugs, and her kisses made this last year much easier. I do not think that I will ever have words to tell you how grateful I am and how much I love you both.

List of papers

This thesis is based on four individual papers, which will be referred to by their Roman numbers in the synthesis.

Paper I: de la Riva-Caballero, A. Oribatid mite communities in western Norway. To be submitted to *Norwegian Journal of Entomology*

Paper II: de la Riva-Caballero, A., H. J. B. Birks, A. E. Bjune, H. H. Birks, and T. Solhøy. Oribatid mites assemblages across the tree-line in western Norway and their representation in lake sediments. *Journal of Paleolimnology* **44**: 361-374

Paper III: Larsen, J., A. E. Bjune, and A. de la Riva-Caballero. 2006 Holocene Environmental and Climate History of Trettetjørn, a Low-alpine Lake in Western Norway, Based on Subfossil Pollen, Diatoms, Oribatid Mites, and Plant Macrofossils. *Arctic, Antarctic and Alpine Research* **38**:571-583.

Paper IV: de la Riva-Caballero, A. and L. S. Halvorsen. Neolithic impact on local vegetation at Kvitevoll, Halsnøy Island, western Norway reconstructed from oribatid mites and pollen analysis. To be submitted to *Journal of Archaeological Science*.

All reprints have been made with the permission of the editors.

Journal of Paleolimnology is published under Springer (http://s100.copyright.com/CustomerAdmin/PLF.jsp?lID=2011090_1314958885973)

Arctic, Antarctic, and alpine research is published under University of Colorado

Order Detail ID: 56131228

Arctic, antarctic, and alpine research by University of Colorado, Boulder.Institute of Arctic and Alpine Research Copyright 2006 Reproduced with permission of INSTITUTION OF ARCTIC & ALPINE RESEARCH, UNIVERSITY OF COLORADO - BIOONE in the format Dissertation via Copyright Clearance Center.



Declaration

Papers II to IV are co-authored, and the nature of the contributions by the different authors are listed below.

Paper II

de la Riva-Caballero, A., H. J. B. Birks, A. E. Bjune, H. H. Birks, and T. Solhøy. Oribatid mites assemblages across the tree-line in western Norway and their representation in lake sediments. *Journal of Paleolimnology* **44**: 361-374

A. de la Riva-Caballero: Acari identification and analyses, numerical analyses, writing. HJB. Birks: Numerical analyses, editing.

AE. Bjune: Experimental design, field sampling, editing.

HH. Birks: Project management, experimental design, field sampling, editing.

T. Solhøy: Acari identification, editing.

Paper III

Larsen, J., A. E. Bjune, and A. de la Riva-Caballero. 2006 Holocene Environmental and Climate History of Trettetjørn, a Low-alpine Lake in Western Norway, Based on Subfossil Pollen, Diatoms, Oribatid Mites, and Plant Macrofossils. *Arctic, Antarctic and Alpine Research* **38**:571-583.

J. Larsen: Project management, field sampling, diatom analysis, writing AE. Bjune: Field sampling, pollen and plant macrofossils analyses, writing. A. de la Riva-Caballero: Oribatid mites analysis, writing.

Paper IV

de la Riva-Caballero, A. and L. S. Halvorsen. Neolithic impact on local vegetation at Kvitevoll, Halsnøy Island, western Norway reconstructed from oribatid mites and pollen analysis. Submitted to *Journal of Archaeological Science*.

A. de la Riva-Caballero: Field sampling, oribatid mite analysis, numerical analysis, writing.

LS. Halvorsen: Field sampling, pollen analysis, writing

Oribatid Mites in a Changing World

De la Riva-Caballero, A. 2011. Oribatid Mites in a Changing World. PhD thesis, Bergen Museum and Faculty of Mathematics and Natural Sciences, University of Bergen, Norway

Introduction

Archives and Proxies

Palaeoecology is the study of responses of past organisms to past environments. This includes the study of past climatic changes which are reflected in the change of abundance and occurrence of different biological proxy indicators time. Palaeoecological through reconstructions enable the comparison of past and present ecosystems, which reveal possible causes mechanisms of biological change over time (Birks and Birks 1980, Berglund 1986, Willis and Birks 2006, Douglas 2007, Gamble 2007).

biological proxy biological indicator ofpast environmental conditions (Smol et al. 2001, Douglas 2007). Among a wide variety of biological proxies, the most used are pollen, widely plant macrofossils, chironomids (non-biting midges) (Fig 1), and diatoms (Douglas 2007). The search for additional means of distinguishing climate change has only recently lead to the use of other also commonly found biological proxies such as the tiny oribatid mites known as moss-mites (Solhøy and Solhøy 2000, Erickson and Platt 2007). Remains of animals and plants are deposited and preserved in sediments of lakes, mires

and other anoxic locations, which become archives of past environments. One can reconstruct the development of the environment in and around the lake or mire using continuous sequences of the sediments and the biological proxies contained in them. The use of only one palaeoenvironmental in a proxv reconstruction will always be less satisfactory than reconstructions which several proxies are combined, as the strengths and weaknesses of the individual proxies then are counterbalanced (Birks and Wright 2000, Birks and Birks 2006).

palaeoecology one can reconstruct the organism assemblages as presence or absence data, populations as numbers of individuals of the taxa, and communities as a combination of the assemblages and the individual abundances. This thesis concentrates on the reconstruction of communities, which allows one to draw inferences about the environment and habitat of past ecosystems (Birks and Birks 1980). Each species in a community has an optimum and a niche within its range of environmental tolerance. There is also a community niche, which is the range of environmental factors common to all the the species that comprise community. Identifying a complete past community of oribatid mites is almost impossible because life assemblages are modified by taphonomic processes.

These are the processes the deceased mites go through from the moment they die until they are finally deposited into the sediments. They include transportation from the source to the lake or mire, predation, preservation, and possible redeposition (Birks and Birks 1980, Erickson 1988, Erickson and Platt 2007). There are three main approaches used to reconstruct past habitats: (1) modern analogue, which compares modern communities with the fossil assemblage; numerical (2)

analysis of fossil data to identify possible gradients that reflect environmental change; and (3) analysis of the abiotic component of the sediment (Birks and Birks 1980, Birks 2003). In this study, the first and second approaches are used. These approaches assume that little or no change in the ecological requirements of the species found in Holocene sediments has occurred since the deposition of the fossil assemblage.

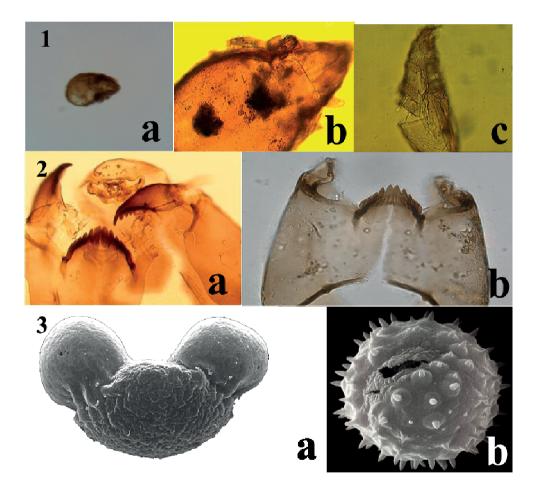


Figure 1. Some examples of biological proxies. **1.** Oribatid mites. a. *Tectocepheus velatus*. b. *Dometorina plantivaga*. c. *Cosmochthonius lanatus* (Picture 1 A. De la Riva-Caballero, pictures 2 and 3. J. Arroyo). **2**. Chironomids a. *Chaetocladius cf. piger* b. *Limnophyes* sp. (pictures G. Velle) **3.** Pollen a. *Pinus*. b. *Valeriana sambucifolia*. (pictures J. Berge).

What are oribatid mites?

The word oribatid comes from the Greek words oreos - mountain and batein - travel (i.e. those that travel around in the mountains). They are also known as moss or box mites. Oribatids are among the most abundant inhabitants of the soil system (Travé et al. 1996, Walter and Proctor 1999). There are ca. 11000 known species, and their local diversity can be high, for example with up to 150 species may coexist in a small area of temperate forest floor (Hansen and Colleman 1998, Solhøy and Solhøy 2000, Norton 2007, Schatz and Behan-Pelletier 2008, Norton and Behan-Pelletier 2009).

Oribatid mites are tiny, flightless arthropods belonging to the subphylum Chelicerata with sizes ranging from 0.13 to 1.2 mm (Walter and Proctor 1999, Norton and Behan-Pelletier 2009). As chelicerates they have two pairs of chelicerae and palpi in the oral region, a body divided in two parts, prodorsum and notogaster (Fig. 2), and three pairs of legs in the larval stages and four in the nymphal and adult stages (Fig. 3). Except for some primitive families, oribatids have a hard. darkly pigmented (chitin) exoskeleton as adults but are usually soft-bodied as immatures (Norton and Behan-Pelletier 2009).

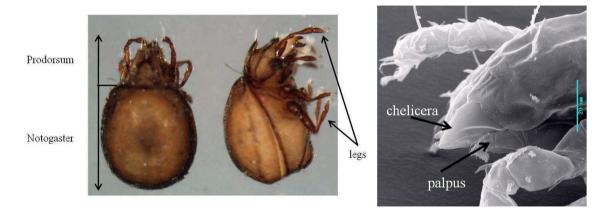


Figure 2. Mite morphology: prodorsum, notogaster, and legs are indicated on a picture of *Xenillus tegeocranus* (left picture, by Stefan Hewig – X. *tegeocranus* average length 720-1100 μ m, Weigmann 2006), chelicerae and palpi are indicated on a SEM picture of an *Hydrozetes* sp. juvenile (right picture, by Anna Seniczak); blue line on the right indicates the measure scale for *Hydrozetes* sp. (20 μ m).

Among oribatids, there are both sexual and asexual species. Generally, ancestral groups have asexual reproduction through thelytoky (production of females, from females, keeping the diploid condition), while most derivative groups tend to show sexual reproduction instead (Travé et al. 1996, Walter and Proctor 1999, Norton Behan-Pelletier 2009). and These minute creatures have a life cycle that

usually takes one or two years in temperate to boreal regions (Norton and Behan-Pelletier 2009), but it can take up to seven years in extreme habitats (Søvik et al. 2003). Their developmental sequence is typical of Chelicerata, with egg, pre-larva (immovable), larva, three nymphal stages (proto-nymph, deuto-nymph and trito-nymph) and the adult, which is the reproductive stage. They do not have

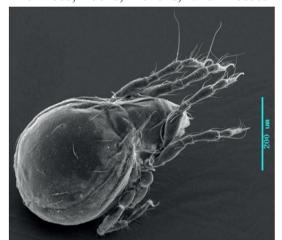
parental care, but they carefully choose where to lay their eggs (Norton 1990).

Oribatids are mostly particle-feeding detritivores (*i.e.* obtain nutrients from decomposing organic matter) and mycophages (*i.e.* feed on members of the fungus kingdom) that live in soil and litter (Luxton 1972, Walter and Proctor 1999, Norton 2007). They play an active role in ecosystems, for example in the decomposition of organic matter, nutrient cycling, and soil formation. In addition, they are good indicators of moisture and pH in both terrestrial and aquatic ecosystems (Behan-Pelletier 1999).

Oribatid mite community composition changes with vegetation composition. The composition of the mite community is controlled partly by food availability, soil type, and moisture (Walter and Proctor 1999, Solhøy and Solhøy 2000). Certain oribatids have rather narrow niches and are obligatory or facultative inhabitants of substrates like trees, rocks, lichens, and mosses

(Norton 1990), which makes them ideal palaeo-indicators of such habitats.

Moss mites are generally not considered to be phoretic organisms; they do not use other organisms for transportation mean. Both Berthet (1964) and Hammer (1965) state that oribatids do not show long dispersal, in part because of their flightless condition and gregarious nature. However, several investigators have shown that dispersal over more than the few meters that oribatids can cover in their lifespans is possible through some kind of phoresy on insects (Norton 1980, Coulson 2009), mammals (Miko and Stanko 1991), and even on birds (Lebedeva and Krivolutsky 2003, Lebedeva Lebedev 2008). However, none of these studies discusses whether this common within oribatid mites. To date, only one family (Mesoplophora) has been shown to have body modifications enable attachment other to organisms (Norton 1980).



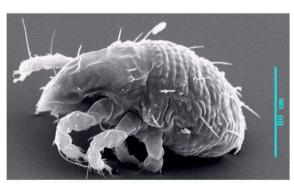


Figure 3. Adult (left) and larva (right) of *Hydrozetes* sp. (pictures by Anna Seniczak). Blue lines on the right of both pictures indicate a scale of 200µm for the adult (left) and 80µm for the larva (right).

Oribatids in palaeoecology

Brief history - Oribatid mites are known from the early Ordovician (Bernini *et al.* 2001) and the Devonian (Norton *et al.* 1988), with some other evolved genera dating back to the Jurassic (Krivolutsky and Drouk 1986) (Fig. 4).

According to Hammer (1965) and Hammer and Wallwork (1979), most extant species had already arisen sometime between the Eocene –

Oligocene and the Pleistocene. Labandeira *et al.* (1997) gives an overview of pre-Pleistocene records. Krivolutsky and Drouk (1986) state that the major trends in the evolution of oribatids were established before the Jurassic, and they suggest that oribatid mites have very low morphogenetic rates. I, therefore, assume that oribatid fossils from the Holocene do not differ in their morphology and behaviour from those of today.

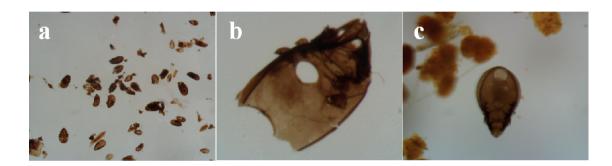


Figure 5. Oribatid subfossils. **a.** general view of the washed out oribatid fragments from a sample **b.** *Liacarus keretinus* (average measure for *L. coracinus* 650-1100μm (Weigmann 2006)). **c.** *Oribatula tibialis* (average measure 410-530μm (Weigmann 2006)) (pictures A. de la Riva-Caballero).

Their flightless condition, hard chitinous exoskeleton (well preserved in anoxic sediments) and narrow niches make oribatid mites very suitable indicators of local past habitats (Erickson 1988, Drouk 1997, Solhøy Solhøy 2000, Solhøy 2001, Erickson and Platt 2007). However, the use of fossil oribatid mites as palaeoindicators is a relatively new discipline (Erickson 1988, Solhøy and Solhøy 2000, Solhøy 2001, Erickson and Platt 2007, Elias 2010). Nordenskiöld (1901) was the first author who identified fossil oribatid mites in his work. After him, many others in the first half of the 20th century mentioned fossil oribatid mites and listed them in their works, but they

did not interpret them ecologically. One of the main problems they faced was that the sediments/layers were not radiocarbon dated, as the technique was not developed until the 1950s by Libby (Gosse 2007). During the second half of the 20th century, the use of fossil oribatids started to expand. In Europe, Knülle (1957a), Karppinen et al. (1979), and Golosova et al. (1985) presented records of fossil oribatids from northern Europe, Greenland, and northern Siberia. Schelvis (1989) was the first researcher to use fossil oribatid mites as indicators of past ecosystems, the interpreting fossil oribatid assemblages found in archaeological sediments from an ecological point of

view and even implementing a semiquantitative method (Schelvis 1989, Schelvis and van Geel 1989, Schelvis 1990a, Schelvis 1990b, Schelvis 1992, Schelvis 1997a, Schelvis 1997b). The most recent works published using oribatid mites fossil from sediments are from Solhøy and Solhøy (2000), Hodgson and Convey (2005), Larsen et al. (2006), and Heggen et al. (2010). The work from Solhøy and Solhøy (2000) is outstanding because of the high resolution of the sediment core, abundant radiocarbon dates. identification to species level of all the adults found, and good documentation of the species' present-day distributions and habitat requirements (Solhøy 2001). Recently, oribatids have been used in palaeolimnology and in archaeology, for example by Schatz et al. (2002), Arroyo et al. (2007), Chepstow-Lusty et al. (2007), and Wild et al. (2007). In North America, Erickson (Erickson 1988, Erickson 1997, Erickson et al. 2003, Erickson and Solod 2008) was the first researcher to use fossil oribatids retrieved from lake sediments to infer climatic/habitat changes.

Sampling sites and identification- The use of oribatid mites as indicators of past habitats relies on their good preservation in anoxic sediments (Fig. 5). Thus, sediments from lakes and mires and other waterlogged environments are usually optimal. Figure 6 illustrates the process of oribatid deposition in a lake. The oribatids most commonly found in lake sediments are aquatic and wetland species living on aquatic plants and other vegetation on the lake shore. However, terrestrial oribatids reached the lake mainly through surface

water run-off and streams or by falling from the nearby vegetation can also be found. Taphonomic processes filter the species and number of oribatids that are finally incorporated into the sediments.

Million Years Ago	Eras	Mayor Divisions	Important Events
0.01	nary	Holocene	
1.64	Quaternary	Pleistocene	ICE AGES
5.2	Tertiary	Pliocene	
23.3		Miocene	
35.4		Oligocene	Mesostigmatans in amber
56.5		Eocene	Fossil ticks
65		Palaeocene	End of dinosaurs
145.6	Mesozoic	Cretaceous	
208		Jurassic	Evolved fossil oribatid genera – Krivolutsky and Drouk 1986
245		Triassic	Mass extintion
290	c	Permian	
362.5	aleozoic	Carboniferous	
408.5	Upper Paleozoic	Devonian	Fossil endeostigmatans and oribatids – Norton et al 1988, Norton et al 1989
438.1	Lower Palaeozoic	Silurian	_
505		Ordovician	Fossil oribatid mites from Sweden – Bernini et al. 2001
545	Lowel	Cambrian	First potential chelicerate
2500	Precambrian	Proterozoic	-
3500	Precar	Archaean	

Figure 4. Acarocentric point of view of the history of life. Modified after Walter and Proctor 1999. Only references related to oribatid mites are indicated.

Identification to species level and a good knowledge of their present-day ecology is of extreme importance for obtaining a reliable reconstruction of the past environment. This knowledge allows oribatid species to be grouped according to their present-day ecology. As with plant macrofossils (Birks 2001), oribatids are usually used to reconstruct local habitat changes as their occurrence and distribution are highly dependent on local substrate

type. In addition, oribatid remains are normally not transported far from their area of origin (Fig. 6) as, in contrast to pollen, they are not wind transported.

In general, knowledge about the broad-scale geographical distribution of oribatid mites is imperfect (Drouk 1997, Behan-Pelletier 1999, Weigmann 2006). This limits detailed reconstruction of past climate using only fossil oribatid assemblages. Oribatid distribution patterns not correspond do biogeographic boundaries and are not limited by known climatic factors, and oribatid's species composition's withinzone variability is as high as its amongzone variability (Drouk 1997). These factors limit the use of oribatid mites to studies. qualitative Nonetheless. Schelvis (1989, 1990b) and Erickson and Platt (2007) have demonstrated the application of oribatid mites in semiquantitative methods (Schelvis 1990a,

Erickson and Solod 2008). Oribatid mite analysis requires patience and good knowledge of not only mite identification from fragments but also the ecology of the species found (Schelvis 1997a, Solhøy 2001, Erickson and Platt 2007). It is far more informative to find a set of species indicating the same type of vegetation than to find many individuals of the same species (Schelvis 1997a). In some instances there are very few remains of mites found in the sediments or too many representatives of generalist species, restricting the interpretation of the past environment and vegetation (Wild et al. 2007). It is, therefore, optimal to combine the study of fossil mites with other proxies that will aid in obtaining a comprehensive understanding of the palaeoenvironment (Birks and Birks 2006).

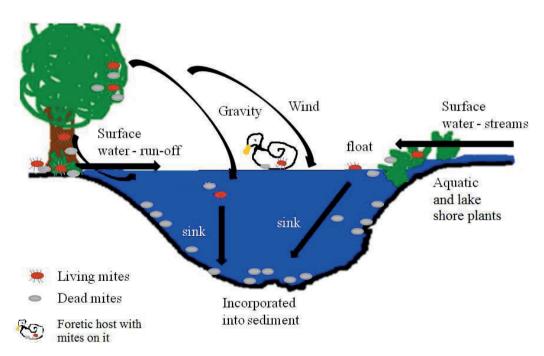


Figure 6. Processes leading to the deposition of terrestrial and aquatic oribatid mites in lake sediments.

Aims of this thesis

My work deals with extant and past oribatid communities. The study of extant oribatid communities improves the knowledge on the ecology and biology of oribatid mites, which is of extreme importance in order to infer habitat changes through oribatid mites. The main scope of the entire thesis is to demonstrate the usefulness of oribatid mites as tools for inferences about past habitats paleoecology in archaeology and to address questions related to the use of oribatid mites as proxies palaeoecology archaeology. This has resulted in 4 papers that address different research questions:

Paper I - Are there distinct oribatid mite communities characteristic of different vegetation types in western Norway? Can we detect characteristic oribatid communities in the tree-line ecotone? The oribatid communities associated with mixed (coniferous and deciduous), birch, pine, and deciduous lichens and mosses, forests. habitats, and the tree-line ecotone are characterized. This type of study is invaluable for palaeoecological it reconstructions, as helps identifying the type of plant community and environment the fossil oribatid assemblages might reflect.

<u>Paper II -</u> Which oribatid ecological groups are best represented in lake sediments? Which is the best coring point within Lake Trettetjørn basin? In this paper I study the representation of oribatid communities at the tree-line ecotone in western Norway and compare it with the

oribatid fossil assemblages found in the Holocene sediments of Lake Trettetjørn, a small lake situated at the present treeline. The modern oribatid fauna from different vegetation zones along an elevational gradient across the tree-line was characterised as a basis comparison with the fossil oribatid assemblages found in sediments from a lake at tree-line within the altitudinal gradient. The ecological information acquired, in addition to that from Paper I, will also be of use in future investigations. The modern assemblages provide a guide to the reliability of the fossil assemblages to reconstruct ecological and environmental changes and, in addition, to finding the most favourable point to take a sediment core within the small lake.

Paper III – Does the mite fauna in lake sediments of Trettetjørn reflect Holocene environmental changes? What does the addition of oribatid mites to a multiproxy study add to the overall environmental interpretations? In this paper I use oribatid mites as proxies to reconstruct the Holocene environmental history of Trettetjørn, the low-alpine (800 m a.s.l.) small lake in Paper II. This is a multi-proxy study combining mites with pollen, plant macrofossils, and diatoms analyses in an attempt to reconstruct the fluctuations in tree-line and their impact on the lake environment.

<u>Paper IV -</u> Can oribatid mites be used to detect Neolithic or later impact on the surroundings of the settlement? In this study I use oribatid mites in an archaeological context. Mite data are combined with pollen data to reconstruct the environment surrounding a human settlement from

the Norwegian Bronze Age. The study also aimed to identify the start of cereal cultivation at Kvitevoll farm, on Halsnøy Island, western Norway.

Study Areas

The climate in Norway is determined by its northerly location in Europe on the east side of the Atlantic Ocean and the marked relief of the country (Moen 1999). The world climate is divided in 5 major regions based on precipitation and temperature, and Norway falls into 3 of them: the arctic and alpine climate, found in the mountains and the areas far north; the cool temperate climate, found in most of the land below the tree-line; and the warm temperate climate, found in the lowlands in southern and western Norway (Moen 1999).

The study areas of this thesis are mainly situated in western Norway, in the counties of Hordaland and Sogn og Fjordane (papers I, II, III, IV). Thus, I focus on the climate of western Norway. According to Moen (1999), coastal regions of western Norway have an oceanic climate influenced by the sea and characterised by relatively low summer (12°C in average) and high winter (>0°C) temperatures and high precipitation and atmospheric humidity (eKlima 2009). Both precipitation and temperature vary widely not only geographically but also annually. In general, coastal districts receive most of the precipitation during autumn and early winter, while inland precipitation is higher during summer (Moen 1999).

The geology in Norway is complex, but Moen (1999) divides it into two main categories: basement rock and Caledonian mountain chain. The

basement rocks date from the Precambrian and basically are unaffected by later events. They are mainly gneiss, granite, and quartzite and are found in northern and southern The Caledonian Norway. orogeny developed during the Silurian and periods. The Devonian resulting mountain chain stretches southwest to northeast in Scandinavia, and Norway from Rogaland to Finnmark. In addition to the bedrock, the superficial deposits are also important for the vegetation cover that will eventually develop. These deposits vary greatly over Norway. In western Norway there are mainly three types: till, deposited by glaciers; mass-movement deposits, which are rocks, stones, and earth that have slid down a slope and accumulated at its foot; and exposed bedrock with a thin cover of superficial deposits (Moen 1999). The third type dominates the study area of this thesis. The type of soil that develops on top of the superficial depends on deposits climate bedrock. In western Norway, which has an oceanic climate, podsolic soils are typical in forests producing acid litter. Peaty soils are also common in areas with high ground water and where the production of plant material exceeds its decomposition.

Elevation above sea level of the study area varies in each of the studies comprising this thesis. Thus, in paper I there are different transects which vary in elevation from 300 to over 1000 m a.s.l. Lake Trettetjønn is situated at 810 m a.s.l. (paper II and III), but the study area in paper II ranges from 663 m a.s.l. to 1120 m a.s.l. Kvitevoll farm (paper IV) is situated at 55 m a.s.l. Figure 7 shows the position of the study sites.

The study sites are situated in the boreonemoral, southern boreal, middle boreal, northern boreal, and low alpine zones (Moen 1999). Coastal areas and areas along the fjords are in the boreonemoral and southern boreal zones, which are characterised by coniferous and broad-leaved woodlands. Ombrotrophic bogs and dry grasslands are common along the coast and inland along the fjords (Fremstad 1997). Scots pine (*Pinus sylvestris*) is common on

swallow soils and dry areas, while Norway spruce (Picea abies) grows in richer soils. Downy birch (Betula pubescens) is a constant element of the woodlands in this part of the country. Usually, the woodland field layer is dominated by members of Ericaceae, such as Calluna, Empetrum, and Vaccinium. At higher altitudes, the field layer is often dominated by bilberry (Vaccinium *myrtillus*) (Fremstad 1997).

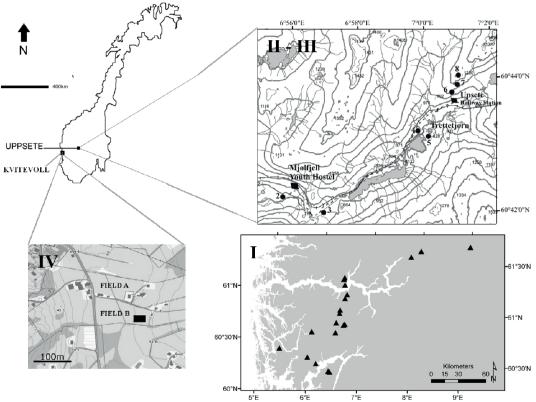


Figure 7. Map of the study areas. Numbers I- II-III-IV on the upper left corner correspond to the different papers. Triangle marks on map I correspond to the sampling sites in paper I from Hardanger northwards to Sognefjord. Scale of map II-III is about 1:100000, see paper II.

Mires are a consistent component of the landscape in western Norway. They are frequently ombrotrophic bogs with dwarf shrubs such as *Calluna*. The tree layer is not common in bogs unless they have dried out. The ground layer is usually dominated by a carpet of *Sphagnum* species (Fremstad 1997). In areas where

the groundwater level is periodically high, swamp woodlands might be found. There are several swamp types in western Norway. Species-poor swamp woodland and grey alder – willow woodland are common in the lowlands (Fremstad 1997, Moen 1999). Soligenous mires, often dominated by Cyperaceae, and lake-margin fens

dominated by Cyperaceae and *Equisetum*, are also common.

Methods

Field sampling

Different sampling methods were selected to match the purpose and scope of the different projects.

<u>Paper I –</u> This project studied the composition of oribatid mite

communities in different habitats in western Norway. Soil samples of 25 x 25 x 5 cm were collected from different vegetation types in the selected habitats. After collection, the samples were extracted using Berlese-Tullgren funnels (Fig. 8) until they were completely dry (ca. one week) and the extracted material was stored in 70% ethanol until sorted and identified.

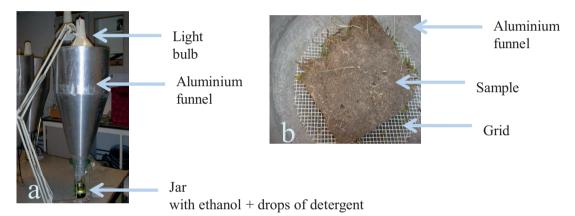


Figure 8. a. Berlese-Tullgren funnel as used at the Department of Biology, UiB. **b.** Inverted sample placed inside the Berlese-Tullgren funnel, viewed from above.

Paper II – In order to study the plant and animal debris that is deposited on the ground and potentially transported into a lake, terrestrial traps were set out in the vegetation types the elevation along gradient. A terrestrial trap consisted of a plastic AstroTurf© doormat, 50x80 cm, which was laid out in the field for a season, winter (September – June) or summer (June - September), and then collected and replaced to collect another season. The sampling was done during four seasons (two years). In addition, to evaluate which oribatid ecological groups were best represented in the modern lake sediments, three traps were set in the lake (Trettetjørn), one at the inlet, one in the middle, and one near the outlet. These traps were large plastic containers weighted down with a large rock and attached to a float to mark their exact location (Birks and Bjune 2010).

<u>Paper III –</u> The sampling followed the standard methodology of palaeoecological studies (Douglas 2007). During winter 2001 a sediment core from the deepest part of the lake (Trettetjørn) was retrieved, using a 110-mm modified piston corer (Nesje 1992).

<u>Paper IV</u> – The sampling followed the standard methodology in archaeology and in the study of Quaternary insects in general (Elias 1994, Elias 2010), in which the size of

the samples analysed tends to be much larger than in palaeolimnology. Samples at different depth intervals (every 3 cm) in the archeological trench were taken and sealed in labelled plastic bags. In addition, a peat block was retrieved from a trench dug in the mire beside the archaeological excavation. The block was wrapped in heavy-gauge aluminium foil in order to preserve it as well as possible until it was sub-sampled in the laboratory.

Laboratory methods

Preparation of samples - Depending on the nature of the project and where the samples were taken, different techniques were needed to prepare the samples so that they could be examined under the stereo microscope to extract the oribatid specimens. Therefore, I describe briefly what was done in each study. Mites were identified following mainly, Gilyarov (1975), Subías and Arillo (2001), and Weigmann (2006). In addition, special papers and reference collections were used in some cases. Specialist were consulted when needed. Nomenclature afterwards, was, standardized to Subías (2004, 2010).

Paper I – After collection, the samples were transported to the Department of Biology at UiB. Mites were extracted from the soil samples using modified Berlese-Tullgren funnels into 70% ethanol with a few drops of detergent (as illustrated in figure 8). After extraction, which usually took one week, the sample was inspected for mites under a stereo microscope. Soil samples might contain more than a thousand individuals of oribatid mites, resulting in tedious and time consuming lab work. Thus, I used

a subsampling method which allows for extrapolation. The methodology was developed by Southwood (1978) and modified by Johannessen (2002) and is described in detail in paper I.

Some oribatid species can be identified to species under a stereo microscope, but most need to be examined under a compound light microscope, following treatment in concentrated lactic acid that renders the chitinous cuticle of most oribatids clear. Dorsal and ventral characteristics of oribatid mites are important to identify them to species. Therefore, they are mounted on non-permanent cavity slides in some drops of concentrated lactic acid which allows them to be turned so that the desired character can be observed at the correct angle.

Paper II – The samples discussed in this paper were collected in doormats or submerged boxes. The doormats were washed using a shower head and the water was passed through a sieve of 125 µm mesh. The residues were stored in water with a little phenol to prevent fungal growth. The rest of the laboratory procedure was similar to many palaeo-studies. Mites were picked out under a stereo microscope and identified to species under the stereo microscope and compound microscope. Because these samples were not palaeosamples, most oribatids still retained their hard chitinous cuticles. These specimens were treated with concentrated lactic acid to make their cuticles clear.

<u>Paper III and IV –</u> These two papers provide practical examples of how oribatids can be used in palaeoreconstructions. In paper III, a sediment core (350 cm long) was retrieved from

Trettetjørn under a water depth of 6.8 m, using a 110-mm modified piston corer (Nesje 1992). Samples for plant macrofossil analysis were taken from 26 levels. The residues of those samples were used for mite analysis.

In paper IV, samples were cut every three cm from the peat-block (83.6 cm long and 16 cm wide) retrieved from a trench excavated in the mire next to the archaeological excavation.

Displacement of water in a measuring cylinder was used to estimate the volume of each sediment sample. In both papers, a sample was usually ca. 30 cm³. It is also possible to measure the weight instead of the volume in order to estimate the total number of oribatid mites found, but I find it less time consuming to measure the volume.

Palaeo-samples are usually disaggregated in dilute sodium pyrophosphate or, if necessary, in 10% KOH (Birks 2001, Solhøy 2001, Elias 2010) and rinsed through two sieves, 125 μm and 250 μm mesh size, in order to divide the sample into two fractions. This facilitates finding the mites when examining the samples under the stereo microscope. Some authors who work in palaeo-studies use flotation techniques to separate the mites from the palaeosamples (see Erickson and Platt 2007 for examples). However, afterwards the residues should also be inspected for mites under the stereo microscope. Therefore, I found it less consuming to pick out the mites from the complete sieved samples under the stereo microscope.

Detailed identification of mite species is done by placing the specimens in some drops of concentrated lactic acid in cavity slides and examining them under a compound light microscope. The identified mites were counted and the numbers normalised to a constant volume (100 cm³) as concentrations.

Data analysis

Depending on the research questions addresses different analytical methods were used.

Paper I – TWINSPAN (Two-way Indicator Species Analysis, Hill 1979) and DISCRIM (simple discriminant analysis, ter Braak 1986) were used to explore the large and noisy data-set and uncover patterns in oribatid distribution. Canonical Correspondance Analysis (CCA) was used to organise the data in relation to the environmental variables. Forward selection and Monte Carlo permutation test were also performed to detect those variables that significantly explain, in a statistical sense, the variance in the Variation data. partitioning was used to assess the relative importance of the spatial variables and habitat conditions in influencing the oribatid communities. The program CANOCO 4.5 Windows (ter Braak and Šmilauer 2002) was used to perform CCA and Variation partitioning.

Paper II – Diagrams were made using TILIA and TGView (Grimm 1993, 2004). CANOCO 4.5 for Windows (ter Braak and Šmilauer 2002) was used to perform Principal Components Analysis (PCA) to summarise the patterns within the modern assemblages, to explore the relationship to the different habitats along the transect, and to compare the modern oribatid fauna along the altitudinal gradient to the fossil oribatid

assemblages in the Trettetjørn Α sediments. simple percentage approach was used to assess how well is of oribatid fauna the represented in the Trettetjørn sediments. Paper III - TILIA and TGView (Grimm 1993, 2004) were used to make the stratigraphic diagrams. CANOCO 4.5 for Windows (ter Braak and Šmilauer 2002) was used to perform a Detrended Correspondence Analysis (DCA) to summarise the main trends in the mite data through the core.

<u>Paper IV</u> – TILIA and TGView (Grimm 1993, 2004) were used to make the stratigraphic diagrams. CANOCO 4.5 for Windows (ter Braak and Šmilauer 2002) was used to perform a PCA to summarise the main trends in the oribatid data in that part of the sequence where they were found.

Outline of main results in the four papers

Background fauna characterisation

Paper I - Oribatid mite communities in western Norway

Distinct oribatid mite communities are characteristic of different vegetation tvpes in western Norway. TWINSPAN (Hill 1979) and DISCRIM (ter Braak 1986) analysis (Fig. 9 and Paper I), and also the CCA. conveniently divided the whole dataset into subsets according to the main habitat. This facilitated the characterisation ofthe oribatid communities found in each of the modern habitat types included in the study.

This work comprises the most exhaustive study of oribatid mites in Norway since Karppinen's work in 1971. A total of 64 new oribatid records for Norway were documented. Numbers of single species vary between several thousand to a few hundred individuals, and total sample numbers are always several thousand individuals.

Chamobates borealis, Oppiella nova, Moritzoppia neerlandica, and Rhinoppia subpectinata characterised the oribatid communities from Betula, mixed, and Picea forest. Achipteria coleptrata, Acrotritria ardua, Ceratozetes gracilis, and Oribatella calcarata were the most common species in deciduous forest.

Hemileius initialis. Nanhermannia coronata, C. borealis, Tectocepheus velatus, and Atropacarus striculus were the most common species in wet habitats. Limnozetes ciliatus, Mucronothrus nasalis, Trimalaconothrus glaber appeared only in waterlogged habitats, which confirms their status as true aquatic species. The lichen and habitat moss characterised Carabodes by labyrinthicus, *C*. marginatus, Melanozetes mollicomus, and T. velatus. The forest species *Parachipteria* punctata (Schatz 1983, Weigmann 2006) was rather common in this habitat, which can be explained by the frequent presence of moss in the samples from this habitat type.

Can we detect characteristic oribatid communities in the tree-line ecotone? The reconstruction of the tree-line is a recurrent topic in palaeoecology (Bjune 2005, Larsen *et al.* 2006, Birks and Bjune 2010). The characterisation of the fauna inhabiting

this ecotone may help in identifying this ecotone in Holocene sediments, and this study of the oribatid communities in this ecotone in western Norway addresses this need. Generalist species such as T. velatus, H. initialis, and Oribatula tibialis dominated this habitat. forest species However. such Camisia biurus (Schatz 1983, Gjelstrup and Solhøy 1994), and lichen associated species such as Carabodes marginatus (Seyd and Seaward 1984) were also found at the tree-line ecotone, which exemplifies the boundary character of this habitat.

Several authors have underlined the importance of a good knowledge of the modern ecology of the oribatid fauna when using them in palaeostudies (Schelvis 1990a, Solhøy 2001, Erickson and Platt 2007,). Therefore, studies like this one, in which oribatid communities from different habitats are characterised, are extremely useful in palaeoecology. In particular, the study provides thorough knowledge of the oribatid species and the communities that occur in western Norway today.

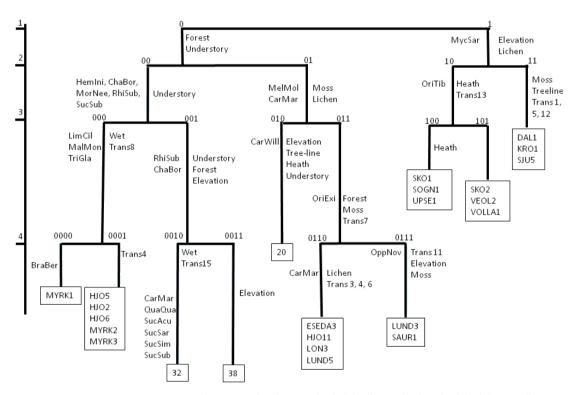


Figure 9. TWINSPAN-DISCRIM diagram. The line on the left indicate the level of division. Indicator environmental variables are on the right of the class. Indicator species are on the left. The abbreviations at the end of each nodule correspond to the samples as indicated in table ESM1 from paper I.

Oribatid mite representation in lake sediments and best coring point

Paper II - Oribatid mite assemblages across the tree-line in western Norway and their representation in lake sediments

The best represented oribatid ecological group in the lake traps were aquatic, followed by oribatids from the habitats adjacent to the lake (Fig. 10). Similar results have been obtained in other studies using lake sediments (Solhøy and Solhøy 2000, Larsen *et al.* 2006, Heggen *et al.* 2010), providing good evidence that oribatids are excellent proxies of local habitats.

In a multidisciplinary approach the same sampling point might not be optimal for all the proxies analysed. In palaeolimnology, cores are usually retrieved from the deepest part of the basin in order to obtain the least

disturbed and the longest, most complete sequence (Birks and Birks 1980, Berglund 1986, Glew and Last 2001). However, it seems that for oribatid mite analysis this is not always the optimal location (Erickson 1988, Solhøy and Solhøy 2000). The results of this paper show that within Lake Trettetjørn basin a core retrieved from the middle of the basin adequately reflects the oribatid fauna of the catchment area.

The comparison of the Holocene oribatid assemblages from paper III and the oribatid fauna found in the lake traps from paper II showed the importance of identifying the oribatid individuals to species level. This increases the ecological indicator value and consequently the reliability of any palaeo-reconstruction done using oribatid mites.

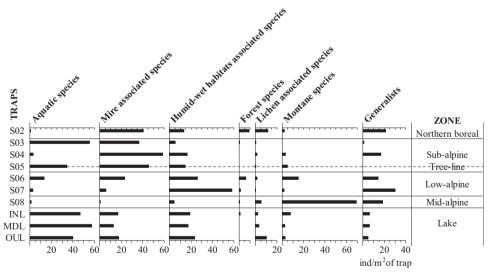


Figure 10. Diagram showing the different ecological groups found along the altitudinal gradient and in the Trettetjørn traps. The horizontal axis represents the calculated number of oribatid individuals found per m² of doormat trap over the collection period of 2 years. From left to right: aquatic species, mire associated species, moist-habitats associated species, forest species, lichen associated species, montane species and generalists. The vertical axis represents the different traps along the altitudinal gradients (S02 to S08) and in Trettetjørn (INL – inlet, MDL – middle, OUL – outlet). The dashed line through S05 indicates that this trap site is situated at the tree-line.

Palaeolimnological and archaeological reconstructions using oribatid mites

The use of oribatid mites as indicators of past habitats, as stated above, is a rather recent approach. The fact that many aspects of oribatids' biology and ecology are unknown makes it difficult to implement quantitative techniques using oribatid fossil data. However, this is not a problem when reconstructing past habitats, as outstanding studies oribatid mites have using been published, including one from Kråkenes in western Norway (Solhøy and Solhøy 2000). This thesis is also comprised of two reconstructions of past habitats oribatid mites habitat using as indicators, paper III and paper IV.

Paper III: Holocene environmental and climate history of Trettetjørn, a low-alpine lake in western Norway, based on subfossil pollen, diatoms, oribatid mites, and plant macrofossils

This study combined data on oribatids with pollen, plant macrofossil, and diatom data. The results obtained from the fossil oribatid analysis added reliability to this study, as they point to the same direction of environmental change as the other proxies involved in this project. In addition, the weaknesses and strengths of the different proxies are counterbalanced.

The Holocene oribatid mite sequence was divided into 6 phases, following the pollen zones (Figures 3 and 4 in paper III) with a time span between 8500 and 1000 cal. yr. BP. In the first or pioneer pollen zone (8575–8270 cal. yr. BP) no mites were

recovered. After deglaciation most probable few oribatids populated the landscape, and therefore few or no oribatid is likely to be incorporated into the sediments. The unstable landscape might also have an stronger physical effect on the fossil mites during deposition, *i.e.* in the processes of being washed from the catchment.

Aquatic oribatid species became abundant in the second zone with the development of aquatic and shore vegetation after 8578 cal yr BP. This might also indicate a wetter climate, with a higher number of oribatids being washed into the basin. The oribatid assemblage was a combination of open habitat and woodland species such as *Liebstadia similis* and *Dometorina plantivaga* (Grandjean 1950, Travè 1963, Solhøy 1976, Solhøy 1979, Maraun and Scheu 2000) during the second zone (8270–7760 cal. yr. BP).

The macrofossil record indicated that woodland of birch and pine became closed around Trettetjørn during the third phase (7760–5200 cal. yr. BP) associated with the occurrence of the oribatid species *Ophidiothricus tectus* and *D. plantivaga* that are typical woodland species (Weigmann 2006).

During the fourth phase (5200-4175 cal. yr. BP), there was a retreat of the forest probably due to a cooling of the climate. No oribatid species associated with woodland were found. This corresponded to the increase in oribatids indicating more open habitats such as Hypochthonius rufulus. This species is also associated with wet to habitats (Solhøy soaked According to the pollen record it seems that mires developed locally during this phase, but judging from the oribatid

record, mire formation had already started in the previous phase, in which the oribatid species *Trimalaconothrus* spp., *Malaconothrus* sp., *Moritzoppia neerlandica*, associated with wet habitats (Solhøy 1979, Weigmann 2006), increased.

Trees around Trettetjørn disappeared in the fifth phase (4175– 1555 cal. yr. BP). Forest oribatids were still absent from the sediments, and there was a slight increase of oribatids characteristic of mires and grassland such as Nanhermannia dorsalis (as N. coronata in the paper) and Banksinoma lanceolata (Karppinen 1955, Tarras-Wahlberg 1961, Solhøy 1976, Solhøy 1979). During this phase, members of the family Camisiidae, an oribatid taxon often associated with heath (Solhøy 1979), were also found. This increase of terrestrial oribatids might be a result of increased water transport to the lake by increased precipitation. This might also have resulted in the increase in fern spores as soil was washed in from the catchment.

During the last phase (1555 cal. yr. BP-AD 2000) there were few oribatid remains. The pollen record indicated a more intense use of the landscape through grazing and summer farming, which would result in an artificial lowering of the local tree-line. There was an increase in microscopic charcoal, which is further evidence for human impact in the area. Aquatic plants disappeared and also aquatic mites, only oribatids associated with mires and few others indicating heath were found. There were several remains of oribatids from the Oppiidae family. According to Behan-Pelletier (1999) and Iturrondobeitia et al. (2005) the number of oribatid species decreases with human disturbances. However, members of the Oppiidae family seem not to suffer as much as other oribatid taxa and are still present after habitat disturbance. Therefore, even if the remains found at Trettetjørn were not identified to species level, they could be an indication of a higher disturbance level in the area by sheep and goat grazing around the lake.

Paper IV: Neolithic impact on local vegetation at Kvitevoll, Halsnøy Island, western Norway reconstructed from oribatid mites and pollen analysis

This work is the first study in Norway in which oribatids are used as indicators environments of past in archaeological context. A peat sequence was analysed and the preservation of oribatid remains was exceptional. Therefore, it was possible to identify the oribatids to species level. This, in addition to the high abundance oribatid remains. increased the reliability and robustness of the palaeoenvironmental reconstruction.

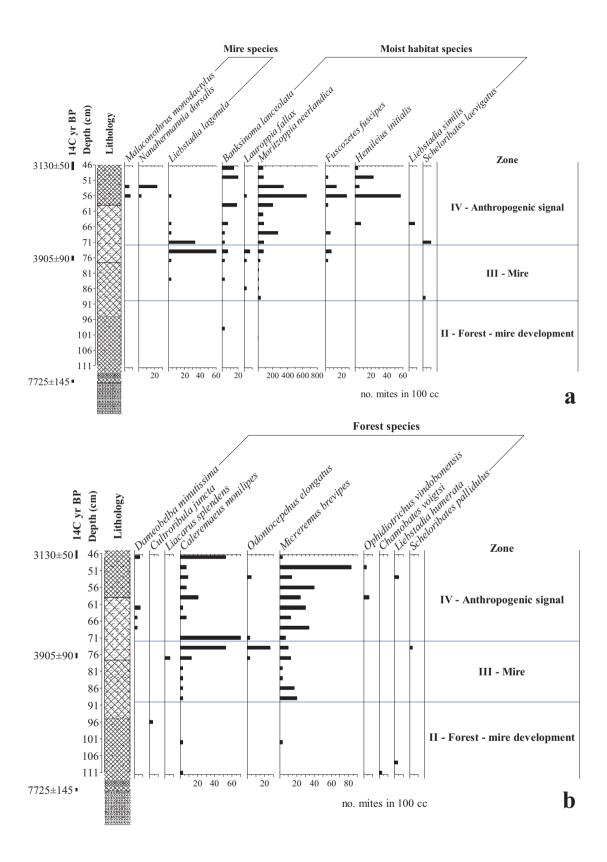
The time span ranges from the late Mesolithic to the Late Neolithic; 7725 ¹⁴C yr BP (6300 cal. yr. BP) at the bottom to 3130 ¹⁴C yr BP (1300 cal. yr. BP) at the top of the sequence. Zonation of the sequence followed major changes in the mite abundances (Fig. 11) and pollen followed the same zonation.

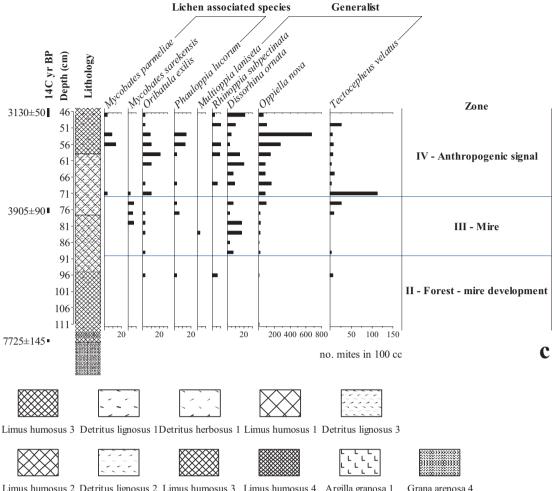
The bottom of the sequence was devoid of oribatid remains and pollen was scarce. It seemed that at this stage there was running water through the site, which was an open landscape dominated by shrubs and wet areas.

This was followed by a rapid increase in trees such as *Betula* and *Salix*, and an increase in *Sphagnum* moss, indicating the onset of mire succession (Fig.4 in Paper IV).

Fossil oribatids do reflect changes in the surroundings of the The oribatid settlement. sequence showed the steadily opening of the woodland around the site (Zone II Fig. 11). The oribatid assemblage was dominated by forest species such as Caleremaeus monilipes and Micreremus brevipes, Cultroribula juncta (Schatz 1989, Subías and Arillo 2001, T. Solhøy pers. comm.), throughout the peat sequence (Fig. 11), but steadily species associated with more open habitats appeared, indicating the opening of the woodland (Fig. 11a). Mire became a dominant element more of landscape. The oribatid fossil assemblage reflected the typical forestmire oribatid community (Zone III, Fig. 11a,b), as suggested by the presence of oribatids the *Fuscozetes* fuscipes, Moritzoppia neerlandica, Liebstadia lagenula all considered peat soil dwellers (Knülle 1957b, Weigmann Miko 2002, Schatz 2004, Weigmann 2006, Schatz and Fischer 2007, Schatz 2008) and forest species such as Liacarus splendens and Scheloribates pallidulus (Subías and Arillo 2001, Weigmann 2006). Towards the end of the sequence the mire got

drier. Oribatid species tolerating drier as conditions such Malaconothrus monodactylus and Nanhermannia dorsalis (Knülle 1957b, Popp 1962, Markkula 1986) replaced Liebstadia which lagenula, prefers wetter conditions. Mites species typical of more open and moist biotopes such as Scheloribates laevigatus, Liebstadia similis, Lauroppia fallax, Moritzoppia neerlandica, and Banksinoma lanceolata (Knülle 1957b, Wunderle et al. 1990, Subías and Arillo 2001, Weigmann 2006) indicated the further opening of the woodland (Zone IV, Fig.11a). However, the mite signal still showed an important presence of trees in the area (Fig. 11b). Species such as Damaeobelba minutissima. **Odontocepheus** elongatus, and Ophidiothricus vindobonensis indicated the presence of well-established (deciduous) forest (Pérez-Íñigo 1997, Weigmann 2006, Schatz and Fischer 2007). Oribatids cannot directly indicate human disturbances, but the increase in members of the family Oppiidae, considered common in disturbed areas (Behan-Pelletier 1999, Clapperton et al. 2002, Iturrondobeitía et al. 2005), as well as the generalist species Tectocepheus velatus, during zone IV, indicated higher disturbances in the area, although not necessarily anthropogenic.





Limus humosus 2 Detritus lignosus 2 Limus humosus 3 Limus humosus 4 Argilla granosa 1 Grana arenosa 4

Figure 11. a. Mite concentrations from the Kvitevoll peat sequence showing mire species, moist-habitat associated species, and lichen associated species. b. Mite concentrations from Kvitevoll showing forest species. c. Mite concentrations from Kvitevoll peat sequence showing lichen associated species and generalists. The data are shown on a depth basis with the three radiocarbon dates (7725±145, 3905±90, 3130±50) on the left-hand side on all diagrams. Lithology symbols follow of all diagrams follow top legend.

Conclusions, problems, and further work

The primary aim of this thesis is to investigate and evaluate the usefulness of oribatid mites as past habitat indicators and to characterise some present day oribatid communities relevant for palaeoecological reconstructions. The results indicate that fossil oribatid mites are valuable and useful indicators of past habitats. Despite the results shown in this and other works, there is still a long way to go before the usefulness of oribatids in palaeoecological studies fully developed. Among the main reasons are the lack of knowledge about oribatids' biology and ecology, about taphonomical processes controlling which species are incorporated into the sediments, and about oribatid mite's taxonomy. The four papers comprised in this thesis contribute to overcome these problems. Paper I does not provide new aspects of their biology, characterises oribatid but it the communities found in common

vegetation types in western Norway. This knowledge is of vital importance when using oribatids as proxies in palaeoecological studies, especially in western Norway. Further meticulously planned studies of community structure, habitat choice of oribatid mites, and changes due to anthropogenic disturbances will be of invaluable use in palaeoecology; they are definitely needed.

The abundance of oribatid remains found, especially in lake sediments, also constitutes interesting problem. There are many aspects of oribatid taphonomy that are still not known, yet they affect which species are incorporated and preserved sediments. could It hypothesised that better preserved and higher numbers of mites might be retrieved from sediment cores taken closer to the lakeshore. However, paper II shows that this is not always necessarily true. The problem is less analysing acute when mire-peat sediments, but in this case, species associated with mires will dominate the sequence. Paper IV shows that peat sequences may result in robust habitat reconstructions, and oribatids indicate the changes in the surrounding local environment.

There is no doubt that reliability and robustness of any palaeoreconstruction using oribatid mites increases with increasing taxonomic resolution. This is because within the same genus there might be species living in completely different habitats. Paper III shows that even if the mite remains are identified at a low level of detail, this taxonomic resolution is also

able to reflect habitat changes in the lake catchment.

Scientific knowledge is not built quickly. It may take years, decades, even centuries before a new concept or paradigm is accepted. Much work is still needed in the palaeo-acarological world, but it is, after all, only a century since oribatids were first discovered as fossils. Thus far, their use palaeoecological contexts has demonstrated that they can provide palaeoenvironmental interpretations. These generally cannot stand alone, but need to be seen in a multi-proxy context where they enhance habitat and environmental reconstructions made from the other proxies.

Acknowledgements

I am grateful to Torstein Solhøy, John Birks, Hilary Birks,Lita G. Jensen and Matteo Pedercini for their helpful comments on earlier versions of this synthesis. I would like to thank Brooke Wilkerson for proof-reading and for her valuable suggestions to improve this synthesis.

References

Arroyo, J., A. de la Riva-Caballero, J. C. Iturrondobeitia, J. M. B. Castro, E. Carbonell, J. L. Arsuaga, and C. Diez. 2007. Primera aproximación a la paleoentomología de los yacimientos de la sierra de Atapuerca (Burgos, España): la fauna subfósil de Oribatidos (Acari, Oribatida). Graellsia, 63: 27-34.

Behan-Pelletier, V. 1999. Oribatid mite biodiversity in agroecosystems: role for bioindication. Agriculture, Ecosystems and Environment, 74: 411-423.

Berglund, B. E. (editor) 1986. Handbook of Holocene Palaeoecology and Palaeohydrology. John Wiley and Sons, Chichester, 869 pp.

Bernini, F., G. Carnevale, G. Bagnoli, and S. Stouge. 2001. An early Ordovician oribatid mite from the island of Öland, Sweden. Pp. 41-49 *in* Proceedings 4th Symposium EURAAC. Kluwer Academic Publishers, Siena, Italy.

Berthet, P. L. 1964. Field study of the mobility of Oribatei (Acari) using radioactive tagging. Journal of Animal Ecology **33**:443-449.

Birks, H. H. 2003. The importance of plant macrofossils in the reconstruction of Lateglacial vegetation and climate: examples from Scotland, western Norway, and Minnesota, USA. Quaternary Science Reviews 22: 453-473.

Birks, H. H. and H. J. B. Birks. 2006. Multi-proxy studies in palaeolimnology. Vegetation History and Archaeobotany 15: 235-251.

Birks H. H. and A. E. Bjune A. E. 2010. Can we detect a west Norwegian tree line from modern samples of plant remains and pollen? Results from the DOORMAT project. Vegetation History and Archaeobotany 19:325-340.

Birks, H. H. and H. E. Wright. 2000. Introduction to the reconstruction of the late-glacial and early-Holocene aquatic ecosystems at Kråkenes Lake, Norway. Journal of Paleolimnology 23:1-5.

Birks, H. H. 2001. Plant Macrofossils. Pp. 50-74 *in* J. Smol, H. J. B. Birks, and W. Last editors. Tracking Environmental Change Using Lake Sediments. Volume 4. Zoological Indicators. Kluwer Academic Publishers, Dordrecht, The Netherlands.

Birks, H. J. B. and Birks H. H. 1980. Quaternary Palaeoecology. Edward Arnold, London. 285 pp.

Bjune, A. E. 2005. Holocene vegetation history and tree-line changes on a north-

south transect crossing major climate gradients in southern Norway - evidence from pollen and plant macrofossils in lake sediements. Review of Palaeobotany and Palynology **133**: 249-275.

Chepstow-Lusty, A. J., M. R. Frogley, B. S. Bauer, M. J. Leng, A. B. Cundy, K. P. Boessenkool, and A. Gioda. 2007. Evaluating socio-economic change in the Andes using oribatid mite abundances as indicators of domestic animal densities. Journal of Archaeological Science 34: 1178-1186.

Clapperton M. J., D. A. Kanashiro, and V. M. Behan-Pelletier. 2002. Changes in abundance and diversity of microarthropods associated with Fescue Prairie grazing regimes. Pedobiologia **46**:496-511.

Coulson, S.J. 2009. Association of the soil maite Diapterobates notatus (Thorell, 1871) (Acari, Oribatidae) with Cynomya mortuorum (Linneaus, 1761) (Calliphoridae, Calliphorinae): implications for the dispersal of oribatid mites, International Journal of Acarology 35: 175-177.

Douglas, M. S. V. 2007. Paleolimnology - Overview. Pp. 2020-2029 *in* Elias, S. A. editor. Encyclopedia of Quaternary Science. Elsevier, Oxford.

Drouk, A. Y. 1997. Acarological Analysis: Problems of Paleoecological Reconstructions. Pp. 91-97 *in* M. E. Edwards, A. V. Sher, and R. D. Guthrie editors. Terrestrial paleoenvironmental studies in Beringia. The Alaska Quaternary Center, University of Alaska Fairbanks, Fairbanks.

Eklima, 2009. Free access to weather- and climate data from Norwegian Meteorological Institute from historical data to real time observations, http://sharki.oslo.dnmi.no/page?_pageid=73
39035.73
add=portal&_schema=P
ORTAL

Elias, S.A. (editor) 1994. Quaternary Insects and their Environments.

Smithsonian Institution Press, Washington DC. 271 pp.

Elias, S.A. 2010. Advances in Quaternary Entomology. Pp. 1-280 in: van der Meer, J.J.M. editor, Developments in Quaternary Sciences series Volume 12, Elsevier, Amsterdam.

Erickson, J. M. 1988. Fossil oribatid mites as tools for Quaternary paleoecologists: preservation quality, quantities, taphonomy. Pp. 207-226 *in* R. S. Laub, N. G. Miller, and D. W. Steadman editors. Late Pleistocene and early Holocene paleoecology and archeology of the eastern Great Lakes region. 33. The Buffalo Society of Natural Sciences.

Erickson, J. M. 1997. Can paleoacarology contribute to global change research? Pp. 533-537 *in* Mitchell, R., D. J. Horn, G. R. Needham, and W. C. Welbourn editors. Proceedings of the 9th International Congress of Acarology. Ohio Biological Survey, Columbus, Ohio.

Erickson, J. M. and R. B. Platt . 2007. Oribatid mites. Pp. 1547-1566 *in* S. A. Elias editor. Encyclopedia of Quaternary Science. Elsevier, Oxford.

Erickson, J.M. and A.M. Solod. 2008. Recognition of postglacial cold intervals by quantitative biozonation of fossil oribatid mites. Pp. 9-16 *In* J. B. Morales-Malacara, V. Behan-Pelletier, E. Ueckermann, T. M. Pérez, E. Estrada, C. Gispert, and M. Badii editors. Acarology XI: Proceedings of the International Congress. Instituto de Biología, UNAM; Facultad de Ciencias, UNAM; Sociedad Latinoamericana de Acarología. México.

Erickson, J. M., R.B. Platt, and D.H. Jennings. 2003. Holocene fossil oribatid mite biofacies as proxies of paleohabitat at the Hiscock site, Byron, New York. Bulletin of the Buffalo Society of Natural Sciences 37:176-189.

Fremstad, E. 1997. Vegetasjonstyper i Norge [Vegetation types in Norway]. Norsk Institutt for Naturforskning, Trondheim. 279 pp. Gamble, C. 2007. Archaeological records - Overview. Pp. 52-63 in Elias, S. A. Encyclopedia of Quaternary Science. Elsevier. Oxford.

Gilyarov, M.S. 1975. A key to the soil-inhabiting mites. Sarcoptiformes. Nauka, Moscow, USSR. 364 pp.

Gjelstrup, P. and T. Solhøy. 1994. Oribatid mites. Pages 1-78 *in* P. M. Jönasson editor. The Zoology of Iceland. Carlsberg-Fond Icelandic Litterature Society, Copenhagen.

Glew, J.R. and W. M. Last. 2001. Sediment core collection and extrusion. Pp. 73-105 *in* W. M. Last, H. J. B. Birks, and J. P. Smol editors. Tracking Environmental Change Using Lake Sediments. Volume 1. Basin analysis, Coring and Chronological Techniques. Kluwer Academic Publishers, Dordrecht, The Netherlands.

Golosova, L., A. Y. Druk, E. Karppinen, and S. V. Kiselyov. 1985. Subfossil oribatid mites (Acarina, Oribatei) of Northern Siberia. Annales Entomologici Fennici **51**:3-18.

Gosse, J.C., 2007. Cosmogenic nuclide dating – Overview. Pp. 409-411 *In*: Elias,S. A.editor, Encyclopedia of Quaternary Science, Elsevier, Oxford.

Grandjean, F. 1950. Sur deux espéces du genre *Dometorina* N. G. et les M'urs de *D. plantivaga* (Berl.) (Acariens, Oribates). Bulletin de la Société Zoologique de France **75**: 224-242.

Grimm, E.C. 1993. Tilia. 2.0.b4. Illinois State Museum, research and collections center. Illinois State Museum

Grimm, E.C. 2004. TGView. 2.0.2. Illinois State Museum, research and collections center. Illinois State Musuem

Hammer, M. 1965. Are Low Temperatures a Species Preserving Factor? Illustrated by the Oribatid Mite Mucronothrus nasalis (Willm). Acta Universitatis Lundensis **2**:2-10.

Hammer, M., Wallwork, J.A. 1979. A review of the world distribution of oribatid mites (Acari: Cryptostigmata) in relation to continental drift, Det Kongelige Danske Videskabernes Selskab, Biologiske Skrifter **22**: 5-31.

Hansen, R. A. and D. C. Coleman. 1998. Litter complexity and composition are determinants of the diversity and species composition of oribatid mites (Acari: Oribatida) in litterbags. Applied Soil Ecology, 9: 17-23.

Heggen, M. P., H. H. Birks, and N. J. Anderson. 2010. Long-term ecosystem dynamics of a small lake and its catchment in west Greenland. The Holocene, **20**: 1207-1222.

Hill, M. O. 1979. TWINSPAN - A FORTRAN program for arranging multivariate data in an ordered two-way table by classification of the individuals and attributes. Ecology and Systematics, Cornell University. Ithaca, New York.

Hodgson, D. A. and P. Convey. 2005. A 7000-year Record of Oribatid Mite Communities on a Maritime-Antarctic Island: Responses to Climate Change. Arctic, Antarctic and Alpine Research, 37: 239-245.

Iturrondobeitía, J.C., A. I. Caballero, and J. Arroyo. 2005. Avances en la utilización de los ácaros oribátidos como indicadores de las condiciones edáficas. Pp. 3-28 *in* A. de Castro editor. Biodiversidad y arácnidos: los invertebrados y la estrategia ambiental vasca de desarrollo sostenible. Munibe, San Sebasatian, Spain.

Johannessen, R. A. 2002. Oribatid mite diversity and abundance in relation to ecological factors along a snow bed to ridge top to snow bed transect in an alpine environment at Finse, Norway. Master thesis unpublished. Department of Zoology, University of Bergen.

Karppinen, E. 1955 Ecological and Transect Survey Studies on Finnish Camisiids. Annales Zoologici Societatis "Vanamo", 17: 3-80.

Karppinen, E. 1971 Studies on the Oribatei (Acari) of Norway. Annales Entomologia Fennoscandica, 37: 30-53.

Karppinen, E., D. A. Krivolutsky, M. Koponen, L. S. Kozlovskaja, L. M. Laskova, and M. Viitasaari. 1979. List of subfossil oribatid mites (Acarina, Oribatei) of northern Europe and Greenland. Annales Entomologia Fennoscandica, **45**: 103-108.

Knülle, W. 1957a. Ocurrence and indicator value of oribatids (Acari: Oribatei) in postglacial deposits. Zoologischer Anzeiger, **159**: 215-229.

Knülle, W. 1957b. The distribution of Acari: Oribatei in the soil. Zeitschrift für Morphologie und Ökologie der Tiere, **46**: 397-432.

Krivolutsky, D. A. and A. Y. Druk. 1986. Fossil oribatid mites. Annual Reviews of Entomology, **31**: 533-545.

Labandeira, C. C., T. L. Phillips, and R. A. Norton. 1997. Oribatid mites and the decomposition of plant tissues in Palaeozoic coal-swamp forest. Palaios, **12**: 319-353.

Larsen, J., A. E. Bjune, and A. de la Riva-Caballero. 2006 Holocene environmental and climate history of Trettetjørn, a low-alpine lake in western Norway, based on subfossil pollen, diatoms, oribatid mites, and plant macrofossils. Arctic, Antarctic and Alpine Research, 38: 571-583.

Lebedeva, N. V. and D. A. Krivolutsky. 2003. Birds spread soil microarthropods to Arctic islands. Doklady Biological Sciences, **391**: 329-332.

Lebedeva, N. V. and V. D. Lebedev. 2008. Transport of oribatid mites to the polar areas by birds. Pp. 359-367 *in* M. Bertrand, S. Kreiter, K. D. McCoy, A. Migeon, M. Navajas, M.-S. Tixier, and L. Vial editors. Integrative Acarology. Proceedings EURAAC 6th Congress. European Association of Acarologists, Montpellier, France.

Luxton, M. 1972. Studies on the oribatid mites of a Danish beech wood soil. Pedobiologia, **12**: 434-463.

Maraun, M. and S. Scheu. 2000. The structure of oribatid mite communities (Acari, Oribatida): patterns, mechanisms, and implications for future research. Ecography, 23: 374-383.

Markkula, I. 1986. Comparison of the communities of the oribatids (Acari: Cryptostigmata) of virgin and forest-ameliorated pine bogs. Annales Zoologici Fennici, 23: 33-38.

Moen, A. 1999. National Atlas of Norway: Vegetation. Norwegian Mapping Authority, Hønefoss.

Miko, L. and M. Stanko. 1991. Small mammals as carriers of non-parasitic mites (Oribatida, Uropodina). Pp. 395-402 *in* F. Dusbábek, and V. Bukva editors. Modern Acarology. 1. Academia and SPB Academic Publishing by, Prague and The Hague.

Nesje, A. 1992. A piston corer for lacustrine and marine sediments. Arctic and Alpine Research, **24**: 257-259.

Nordenskiöld, E. 1901. Zur Kenntnis der Oribatiden Fauna Finnlands. Acta Societatis pro Fauna et Flora Fennica, **21**: 1-34.

Norton, R. A. 1980. Observation on phoresy by oribatid mites (Acari: Oribatei). International Journal of Acarology, **6**: 121-130.

Norton, R. A. 1990. Acarina: Oribatida. Pp. 779-792 in D. L. Dindal editor. Soil Biology Guide. A. Wiley-Interscience Publication, New York.

Norton, R. A. 2007. Holistic Acarology and ultimate causes: examples from the Oribatid mites. Pp. 3-20 *in* J. B. Morales-Malacara, V. Behan-Pelletier, E. Ueckermann, T. M. Perez, E. G. Estrada-Venegas, and M. Badii editors. Acarology XI: Proceedings of the International Congress. Universidad Nacional Autónoma de México Sociedad

Latinoamericana de Acarología, Merida, México.

Norton, R. A. and V. M. Behan-Pelletier. 2009. Suborder Oribatida. Pp. 430-564 in G. W. Krantz and D. E. Walter editors. A Manual of Acarology. Volume 3. Texas Tech University Press, Lubbock.

Norton, R. A., P. M. Bonamo, J. D. Grierson, and W. A. Shear. 1988. Oribatid mite fossils from a terrestrial Devonian deposit near Gilboa, New York. Journal of Paleontology, **62**: 259-269.

Pérez-Íñigo, C. 1997. Acari, Oribatei Gymnonota I. Pp. 1-374 *in* M. A. Ramos Sánchez, J. Alba Tercedor, X. Bellés i Ros, J. Gosálbez i Noguera, A. Guerra Sierra, E. Macpherson Mayol, F. Martín Piera, J. Serrano Marino, and J. Templado González editors. Fauna Ibérica. Museo Nacional de Ciencias Naturales CSIC, Madrid, Spain.

Popp, E. 1962. Semiaquatile Lebensräume (Bülten) in Hoch- und Niedermooren. [Semi-aquatic biotopes (hummocks) in raised bogs and fens. II. The mite fauna]. Internationale Revue der gesamten Hydrobiologie und Hydrographie, **47**: 533-579.

Schatz, H. 1983. U.-Ordn: Oribatei, Hornmilben. *In* Catalogus Faunae Austriae, Teil IXi. Österreichische Akademie der Wissenschfaten, Wien. 118 pp.

Schatz, H. 1989. Oribatida (Acari) aus dem Kalser Dorfertal (Osttirol, Hohe Tauern, Österreich) [Oribatid Mites (Acari: Oribatida) from the Kalser Dorfertal Valley (Central Alps, Eastern Tyrol, Austria)]. Berichte des naturwissenschaftlichmedizinischen Vereins in Innsbruck, 76: 107-125.

Schatz, H. 2004. Hornmilben (Acari, Oribatida) in Auwäldern an der Etsch und Talfer (Südtirol, Italien) [Oribatid mites (Acari, Oribatida) in riparian forest on the rivers Adige and Talvera (South Tyrol, Italy)]. Gredleriana, 4: 93-114.

Schatz, H. 2008. Hornmilben (Acari: Oribatida) im Naturpark Schlern-

Rosengarten (Südtirol, Italien). Gredleriana, **8**: 219-254.

Schatz, H. and V. M. Behan-Pelletier. 2008. Global diversity of oribatids (Oribatida: Acari: Arachnida). Hydrobiologia, **595**: 323-328.

Schatz, H. and B. Fischer. 2007. Hornmilben (Acari: Oribatida) von den Hundsheimer Bergen (Niedeösterreich, Österreich) (Oribatid mites (Acari, Oribatida) from the Hundsheim Mountains (Lower Austria, Austria)). Berichte des naturwissenschaftlich-medizinischen Vereins in Innsbruck, 94: 63-77.

Schatz, I., H. Schatz, F. Glaser, and A. Heiss. 2002. Subfossile Arthropodenfunde in einer bronzezeitlichen Grabungsstätte bei Radfeld (Tirol, Österreich) (Acari: Oribatida: Insecta: Coleoptera, Hymenoptera: Formicidae) [Subfosil] arthropod macro remains from a Bronze Age Site near Radfeld (Tirol, Austria)]. Berichte des naturwissenschaftlichmedizinischen Vereins in Innsbruck, 89: 249-264.

Schelvis, J. 1989. Mites (Acari) from the late Neolithic well at Kolhorn (The Netherlands). Palaeohistoria, **31**: 165-171.

Schelvis, J. 1990a. The reconstruction of local environments on the basis of remains of oribatid mites (Acari; Oribatida). Journal of Archaeological Science, **17**: 559-571.

Schelvis, J. 1990b. Mites from Medieval Oldeboorn. An environmental reconstruction. Proceedings Experimental and Applied Entomology, 1: 96-97.

Schelvis, J. 1992. Mammoths and mites. Proceedings Experimental and Applied Entomology, **3**: 140-141.

Schelvis, J. 1997a. Mites in the Background. Use and Origin of Remains of Mites (Acari) in Quaternary Deposits. Quaternary Proceedings, 5: 233-236.

Schelvis, J. 1997b. Remains of mites (Acari) as palaeo-ecological indicators: dwelling mounds and medieval

reclamations in The Netherlands *in* Environment and Subsistence in Medieval Europe - Medieval Europe 9th Conference. Institute for the Archaeological Heritage, The Netherlands.

Schelvis, J. and B. van Geel. 1989. A palaeoecological study of the mites (Acari) from a lateglacial deposit at Usselo (The Netherlands). Boreas, **18**: 237-243.

Seyd, E. L. and M. R. D. Seaward. 1984. The association of oribatid mites with lichens. Zoological Journal of the Linnean Society, **80**: 369-420.

Smol, J. P., H. J. B. Birks, and W. M. Last. (editors) 2001. Tracking Environmental Change Using Lake Sediments. Volume 4: Zoological Indicators. Kluwer Academic Publishers. Dordrecht. The Netherlands.

Solhøy, I. W. and T. Solhøy. 2000. The fossil oribatid mite fauna (Acari: Oribatida) in late-glacial and early-Holocene sediments in Kråkenes Lake, western Norway. Journal of Paleolimnology, **23**: 35-47

Solhøy, T. 1976. Species composition of the Oribatei (Acari) on oceanic mountain ground in western Norway. Norwegian Journal of Entomology, **23**: 17-22.

Solhøy, T. 1979. Oribatids (Acari) from an oligotrophic bog in western Norway. Fauna Norvegica, **26**: 91-94.

Solhøy, T. 2001. Oribatid mites. Pp. 81-104 *in* J. P. Smol, H. J. B. Birks, and W. M. Last editors. Tracking environmental change using lake sediments. Volume 4. Zoological Indicators. Kluwer Academic Publishers, Dordrecht, The Netherlands.

Southwood, T.R.E., P.A. 1978. Ecological Methods. With particular reference to the study of insect populations. Chapman and Hall. London. 524 pp.

Subías, L. S. 2004. Listado sistemático, sinonímico y biogeográfico de los ácaros oribátidos (Acariformes, Oribatida) del mundo (Excepto fósiles) (1758 - 2002). Graellsia **60** (special number): 3-305.

Subías, L. S. 2010. Listado sistemático, sinonímico y biogeográfico de los ácaros oribátidos (Acariformes, Oribatida) del mundo (Excepto fósiles) (1758 - 2002). http://www.ucm.es/info/zoo/Artropodos/Catalogo.pdf.

Subías, L. S. and A. Arillo. 2001. ACARI, Oribatei, Gymnonota II. Pp. 289 *in* Fauna Ibérica, eds M. A. Ramos Sánchez, J. Alba Tercedor, X. Bellés i Ros, J. Gosálbez i Noguera, A. Guerra Sierra, E. Macpherson Mayol, F. Martín Piera, J. Serrano Marino, and J. Templado González editors. Museo Nacional de Ciencias Naturales CSIC, Madrid, Spain.

Søvik G., H. P. Leinaas, R. A. Ims, and T. Solhøy. 2003. Population dynamics and life history of the oribatid mite Ameronothrus lineatus (Acari, Oribatida) on the high arctic archipelago of Svalbard. Pedobiologia, 47: 257-271.

Tarras-Wahlberg, N. 1961. The oribatei of a central Swedish bog and their environment. Doctoral Thesis. Zoological institute. University of Lund.

ter Braak, C.J.F. 1986. Interpreting a hierarchical classification with simple discriminant funtions: an ecological sample. In E. Diday editor. Data Analysis and Informatics IV. Elsevier Science. North-Holland.

ter Braak CJF, P. Šmilauer. 2002. CANOCO reference manual and CanoDraw for windows user's guide: software for canonical community ordination (version 4.5). Microcomputer Power, Ithaca, 500 pp.

Travé, J. 1963. Écologie et biologie des Oribates (Acariens) saxicoles et arboricoles. Publications du laboratoire Arago, Université de Paris, France 14: 1-267.

Travé, J., H. M. André, G. Taberly, and F. Bernini. 1996. Les Acariens Oribates. Wavre, Belgium. 110 pp.

Walter, D. and H. Proctor. 1999. Mites: ecology, evolution and behaviour. CABI Publishing, New York. 323 pp.

Weigmann, G. 2006. Acari. Actinochetida. Hornmilben (Oribatida). Pp. 520 *in*: Dahl. F. editor, Die Tierwelt Deutschland und der angrenzenden Meeresteile. 76 Teil. Goecke and Evers Keltern.

Weigmann, G. and L. Miko. 2002. Redescription of *Oribates lagenula* Berlese, 1904, the type of *Lagenobates* n. gen. (Acarina Oribatida). REDIA, **85**: 29-35.

Wild, V., I. Schatz, and H. Schatz. 2007. Subfossile Arthropodenfunde (Acari: Oribatida, Insecta: Coleptera) in Mooren bei der Schwarzensteinalm im Oberen Zemmgrund in den Zillertaler Alpen (Österreich). BFW-Berichte, **141**: 117-131.

Willis, K. J. and H. J. B. Birks. 2006. What is natural? The need for a long-term perspective in biodiversity conservation. Science, **314**: 1261-1265.

Wunderle, I., L. Beck, and S. Woas. 1990. Ein Beitrag zur Taxoinomie und Ökologie der Oribatulidae und Scheloribatidae (Acari, Oribatei) in Südwestdeutschland (A contribution to the taxonomy and ecology of the Oribatulidae and Scheloribatidae (Acari, Oribatei) in Southwest Germany). Andrias, 7: 15-60.