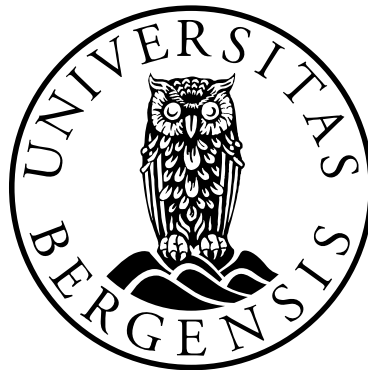


Components of the choline oxidation pathway in relation to acute myocardial infarction, type 2 diabetes and mortality

Prospective observational studies among patients with suspected or verified coronary heart disease in Norway

Gard Frodahl Tveitevåg Svingen



Dissertation for the degree of philosophiae doctor (PhD)
at the University of Bergen

2016

Dissertation date: 7 April

© Copyright Gard Frodahl Tveitevåg Svingen

The material in this publication is protected by copyright law.

Year: 2016

Title: Components of the choline oxidation pathway in relation to acute myocardial infarction, type 2 diabetes and mortality.
Prospective observational studies among patients with suspected or verified coronary heart disease in Norway

Author: Gard Frodahl Tveitevåg Svingen

Print: AiT Bjerch AS / University of Bergen

ISBN 978-82-308-3360-5

Bergen, Norway 2016

LIST OF SELECTED ABBREVIATIONS

ACE – Angiotensin converting enzyme
ACS – Acute coronary syndrome
ACVD– Atherosclerotic cardiovascular disease
AMI – Acute myocardial infarction
Apo – Apolipoprotein
BADH – Betaine aldehyde dehydrogenase
BHMT – Betaine-homocysteine *S*-methyl transferase
BMI – Body mass index
CABG – Coronary artery bypass graft
CAD – Coronary artery disease
CHDH – Choline dehydrogenase
CBS – Cystathionine- β -synthase
CI – Confidence interval
CoR – Coefficient of reliability
CRP – C-reactive protein
CVD – Cardiovascular disease
DMG – Dimethylglycine
DDH – Dimethylglycine dehydrogenase
GAM – Generalized additive model
GFR – Glomerular filtration rate
GNMT – Glycine-*N*-methyltransferase
HDL-C – High density lipoprotein cholesterol
ICC – Intraclass correlation coefficient
IDI – Integrated discrimination index
HR – Hazard ratio
LDL-C – Low density lipoprotein cholesterol
LME – Linear mixed effects model
LVEF – Left ventricular ejection fraction
MMA – Methylmalonic acid

MS – Methionine synthase
NAFLD – Non-alcoholic fatty liver disease
NRI – Net reclassification improvement
NORVIT – Norwegian Vitamin Trial
PCI – Percutaneous coronary intervention
PPAR – Peroxisome proliferator-activated receptor
ROC-AUC – Receiver operating characteristics-area under the curve
RCT – Randomized controlled trial
SAH – *S*-adenosylhomocysteine
SAM – *S*-adenosylmethionine
SAP – Stable angina pectoris
SARDH – Sarcosine dehydrogenase
SD – Standard deviation
SSH – Sarcosine dehydrogenase
SMC – Smooth muscle cell
SNP – Single nucleotide polymorphism
STROBE-ME – STrengthening the Reporting of OBServational studies in
Epidemiology-Molecular Epidemiology
tHcy – Total homocysteine
TMAO – Trimethylamine *N*-oxide
T2D – Type 2 diabetes
UAP – Unstable angina pectoris
VLDL – Very low density lipoprotein
WECAC – Western Norway Coronary Angiography Cohort
WENBIT – Western Norway B vitamin Intervention Trial

SCIENTIFIC ENVIRONMENT

The current thesis is based upon studies of two Norwegian cohorts, made up of patients with suspected and/or verified coronary heart disease (CHD); The Western Norway Coronary Angiography Cohort (WECAC) and the Norwegian B-vitamin Intervention Trial (NORVIT). Although the NORVIT was a multicentre national intervention study, the scientific environment of the current thesis has mainly been localized to the University of Bergen, with Professor Ottar Kjell Nygård as main supervisor and Professor Per Magne Ueland and PhD Eva Ringdal Pedersen as co-supervisors. However, the work has been a joint collaboration with other local and national research environments, including the Western Norway Cardiovascular Registry (WENOCARD), the Cardiovascular Disease in Norway (CVDNOR) project, the KG Jebsen Center for Diabetes Research, and Bevital AS, as well as scientific co-workers at the Department of Heart Disease, Haukeland University Hospital and Stavanger University Hospital, and the Norwegian University of Science and Technology.

Funding of the current project has mainly been provided by the University of Bergen in terms of a four-year full-time PhD scholarship, but also by the Western Norway Regional Health Authority and the Foundation to Promote Research into Functional Vitamin B12 Deficiency.

ACKNOWLEDGEMENTS

I first met Dr Ottar Nygård in 2000, when working as an assistant in the Department of Heart Disease at Haukeland University Hospital. Little did I know of his many projects at that time, but soon I found myself in front of a computer plotting WENBIT patient data into the CORDOBA database. At first this meant little more to me than some extra and much appreciated money, but I soon gained more interest into the field of cardiology and continued to work on the database also after graduating in 2003. Upon returning to Bergen in 2006 I felt that everyday clinical practice left little room for academic work. However, my recent hospital internship had ignited an interest in academia, and despite much reluctance from my side, the then appointed professor Nygård convinced me into starting the current research project in 2009, thereby introducing me to scientific work. For that I am extremely grateful. Ottar, your philosophy of supervision is based upon the necessity for your students to work independently, and without you having the need for a detailed control of every aspect of their work. This economic exertion of leadership reflects trust in your employees, and for me this has been essential for being able to carry out the work upon which this thesis rests. Your open-mindedness and truly academic approach, the way you promote your students, and how you unselfishly share your vast knowledge as well as extremely hard earned data is nothing but inspirational.

I am also most grateful to my co-supervisors professor Per Magne Ueland and PhD Eva Ringdal Pedersen for their time and effort spent in both critically reviewing my manuscripts and sharing their expertise within the fields of biochemistry, epidemiology and science in general. To be given the chance to work with international top scientists, I cannot be anything but humble. Per, I know PhD supervision may feel as a slow-going phlebotomy to you, but I hope I have not left you critically anaemic.

Further, I thank all my co-authors for their contributions in all aspects of the scientific work, not least because of their hard effort with the WECAC and NORVIT databases, the latter being chaired by professor Kaare Harald Bønnaa at the University of Tromsø. A special tribute goes to professor Grethe Tell at the Department of Global

Public Health and Primary Care, University of Bergen and to Tomislav Dimoski at The Norwegian Knowledge Centre for the Health Services, Oslo, for their efforts made in the CVDNOR project. I feel honoured of being allowed to use these exceptionally fine data in my work. Not least, I thank the study participants in the WECAC and NORVIT cohorts for their invaluable contribution to the current work.

I also wish to thank the scientific environment both within our research group at the Department of Preventive Cardiology, Haukeland University Hospital, and our extended group, comprising scientists and staff at Bevital AS. Apart from your important scientific input, you have provided many laughs and good times, and the annual vitamin meetings abroad definitively have served as highlights. A special tribute is also paid to Britt Gjellefall, for your devoted and professional attitude towards work, for your great humour and for the sincere care you take of your colleagues. During the last four years I have also shared office with my colleagues and co-authors Hall, Reinhard, Kjetil and Jonas. Our microenvironment has consisted of not only low oxygen tension and tons of teenage humour, but also important discussions on politics, history, statistics, science, and not least the hits in the face that life provides. Thank you.

Although my interest for cardiology was sparked on early during my student years, it was consolidated when carrying out my hospital internship under the guidance of Dr Ulf Hurtig. As a brilliant clinician and firm believer in evidence based medicine, you are an inspiration for carrying out both clinical and scientific work.

Although highly inspirational and rewarding, this journey has at times also felt strenuous and frustrating. I simply would not have been able to make it without the support from my closest family; to my mother Grethe and father Jan Erik, thank you for your unconditional love, confidence and support that you have showed me throughout my life. Thanks also to my brother Kjetil, my sister-in-law Anna, to my uncle Svein and aunt Grethe, and to my family-in-law Karen, Terje and Espen.

Finally, Renathe, this work is dedicated to my true life companions: you, Tobias, Vebjørn and Håvard. I love you.

Gard Frodahl Tveitevåg Svingen,
Bergen, January 2016

ABSTRACT

BACKGROUND

The choline oxidation pathway comprises the sequential metabolism of choline into betaine, dimethylglycine (DMG), and sarcosine. In addition, dietary choline and betaine can be turned into trimethylamine *N*-amino oxide (TMAO). Alterations in choline metabolism may relate to cardiovascular disease (CVD) and type 2 diabetes (T2D). Several investigations have focused on systemic concentrations of choline and betaine; however large-scale prospective data are scarce. There is therefore a need for more comprehensive assessments of choline metabolites in relation to incident CVD, T2D and mortality, in addition to investigating any potential benefit in risk prediction from such biomarkers.

AIM

We carried out observational cohort studies of the prospective relationships between plasma DMG and incident acute myocardial infarction (AMI) and mortality, as well as the association between systemic and urinary choline metabolites with the risk of incident T2D. The biomarkers' impact on model discrimination and reclassification of patients at risk was also assessed, as were their test-retest reliabilities and temporal trends according to B-vitamin treatment.

MATERIALS AND METHODS

Analyses on the association between plasma DMG and incident AMI, as well as the relationships between choline metabolites and incident T2D were performed among patients evaluated for suspected stable angina pectoris (SAP). The risk assessment between plasma DMG and mortality included SAP patients and patients with AMI from an independent cohort. Clinical endpoint data were obtained from regional and national health registries. Endpoint analyses on incident AMI and mortality were carried out by Cox regression, whereas analyses on incident T2D were performed by logistic regression. Model discrimination and reclassification were explored by calculating the C-statistics, the integrated discrimination index (IDI), and the

continuous net reclassification improvement (NRI>0), respectively. Mixed linear modelling was used for assessing temporal trends in metabolite concentrations.

RESULTS

Higher plasma DMG was associated with several traditional risk factors for coronary heart disease (CHD). After about four and a half years of follow-up, plasma DMG showed linear relationships with incident AMI among 4154 patients with suspected SAP (age, gender and fasting adjusted hazard ratio (HR) for the fourth vs. first quartile (95% confidence intervals (CI)) 1.95 (1.42-2.68); $P < 0.001$). The relationship between plasma DMG and incident AMI was particularly strong among non-smokers and patients with lower serum apoB and triglyceride levels (P for interaction ≤ 0.03).

Among essentially the same patients, as well as among 3733 patients with AMI who were followed for 7 years, higher plasma DMG was also associated with increased risk of all-cause mortality (age and gender adjusted HRs (95% CIs) for the fourth vs. the first quartile 1.72 (1.21–2.46) and 1.76 (1.42–2.18) among SAP and AMI patients, respectively) and CVD mortality (HRs (95% CIs) 1.94 (1.21–3.11) and 1.97 (1.50–2.59) among SAP and AMI patients, respectively). The associations were only slightly attenuated when adjusting for established CHD risk factors, to which adding plasma DMG also improved risk prediction for both AMI and all-cause mortality. Moreover, plasma DMG showed good to excellent within-person reproducibility throughout repeated measurements among patients not receiving supplementation with folic acid + vitamin B12.

In general, higher plasma choline and lower plasma betaine and serum sarcosine levels were associated with an adverse risk profile of T2D. In urine, most choline metabolites were positively related to an adverse diabetes risk profile. After an average follow-up of 7.5 years, 233 (6.4%) out of 3621 non-diabetic patients were registered with new-onset T2D. Incident T2D was strongly inversely associated with plasma betaine and positively related to urine betaine, DMG and sarcosine (age, gender and fasting adjusted odds ratios (95% CIs) per 1 SD increment 0.72 (0.62-0.83), 1.25 (1.09-1.43), 1.22 (1.06-1.40), and 1.30 (1.13-1.49), respectively). We did not find any relationship between choline or TMAO and incident T2D. The estimates were not

materially altered when adjusting for a range of traditional T2D risk factors and potential confounders, and were similar in sensitivity analyses. Among the choline metabolites associated with new-onset T2D in univariate analyses, plasma betaine and urine sarcosine were most strongly related to incident T2D, and both indices also enhanced risk prediction when added to the multivariate model.

After 1 year, as compared to placebo treatment, supplementation with folic acid + vitamin B12 lowered plasma DMG and sarcosine, but increased plasma betaine and choline. No alterations in plasma TMAO were observed. In urine, we observed similar responses to supplementation as to those seen in blood.

CONCLUSION AND IMPLICATIONS

Among patients with suspected or verified SAP, high plasma DMG was related to increased risk of AMI, as well as all-cause and CVD mortality, the latter endpoints being validated among patients with AMI. Moreover, plasma DMG improved risk prediction of both AMI and mortality.

Lower plasma betaine and higher urine betaine, DMG and sarcosine concentrations were related to incident T2D. Plasma betaine and urine sarcosine also improved reclassification of patients at risk. Plasma DMG and betaine, as well as urine sarcosine showed good to excellent within-subject reproducibility among patients not supplemented with folic acid + vitamin B12, justifying one-time assessment of biomarker status.

Our observational findings suggest novel pathophysiological pathways involved in conditions heavily impacting the global burden of disease, warranting more research into the field of one-carbon and choline metabolism in relation to life-style related diseases.

LIST OF PUBLICATIONS

The current thesis is based on the following original articles published in international peer-review journals and referred to by their roman numerals:

PAPER I

Svingen GF, Ueland PM, Pedersen EK, Schartum-Hansen H, Seifert R, Ebbing M, Løland KH, Tell GS, Nygård O. Plasma dimethylglycine and risk of incident acute myocardial infarction in patients with stable angina pectoris. *Arterioscler Thromb Vasc Biol.* 2013;33(8):2041-8. (Appendix I)

PAPER II

Svingen GFT, Schartum-Hansen H, Ueland PM, Pedersen EP, Seifert R, Ebbing M, Bønaa KH, Mellgren G, Nilsen DWT, Nordrehaug JE, Øyen J, and Nygård O. Elevated plasma dimethylglycine is a risk marker of mortality in patients with coronary heart disease. *Eur J Prev Cardiol* 2015;22(6):743-52. (Appendix II)

PAPER III

Svingen GFT, Schartum-Hansen H, Pedersen EKR, Ueland PM, Tell GS, Mellgren G, Njølstad PR, Strand E, Karlsson T, Seifert R, Nygård O. The prospective associations of systemic and urinary choline metabolites with incident type 2 diabetes. Submitted, as per January 2016. (Appendix III)

Reprints were made with permission from Wolters Kluwer Health Lippincott Williams & Wilkins (paper I) and SAGE Publications Ltd (paper II). All rights reserved.

TABLE OF CONTENTS

ABBREVIATIONS	3
SCIENTIFIC ENVIRONMENT	5
ACKNOWLEDGEMENTS	6
ABSTRACT	8
LIST OF PUBLICATIONS	11
TABLE OF CONTENTS	12
1 INTRODUCTION	16
1.1 CARDIOVASCULAR DISEASE	16
1.1.1 DEFINITION, EPIDEMIOLOGY AND IMPLICATIONS	16
1.1.2 PATHOPHYSIOLOGY AND CLINICAL PRESENTATION OF IHD	17
1.2 DIABETES MELLITUS	19
1.2.1 DEFINITION, EPIDEMIOLOGY AND IMPLICATIONS	19
1.2.2 PATHOPHYSIOLOGY AND CLINICAL PRESENTATION OF T2D	20
1.2.3 T2D AND CHD ARE STRONGLY RELATED	21
1.3 THE CHOLINE OXIDATION PATHWAY	22
1.3.1 A BRIEF OVERVIEW	22
1.3.2 CHOLINE AND BETAINE CAN BE PRODUCED ENDOGENOUSLY OR OBTAINED FROM THE DIET	22
1.3.3 RAMIFICATIONS OF CHOLINE METABOLISM	24
1.3.4 CHOLINE AND BETAINE METABOLISM IN THE GUT – THE FORMATION OF TRIMETHYLAMINE- <i>N</i> -OXIDE	27
1.3.5 CHOLINE AND BETAINE IN RELATION TO HEPATIC LIPID METABOLISM	28
1.3.6 HCY AS A CARDIOVASCULAR RISK FACTOR – A LONG STORY MADE SHORT	28
1.3.7 CHOLINE METABOLISM IN RELATION TO CVD AND DIABETES	29
1.3.8 OPEN RESEARCH QUESTIONS	31
2 AIMS OF THE THESIS	32

3 MATERIALS AND METHODS	33
3.1 PATIENT POPULATION	33
3.1.1 THE WENBIT, NORVIT AND WECAC COHORTS	33
3.1.2 FOLLOW-UP AND CLINICAL ENDPOINTS	34
3.1.3 LABORATORY ANALYSES	35
3.1.4 DATA ON DIETARY CHOLINE AND BETAININE INTAKE	39
3.2 STATISTICAL METHODS	39
3.2.1 SOFTWARE	39
3.2.2 BASELINE DATA	40
3.2.3 ENDPOINT ANALYSES	41
TIME-TO-EVENT ANALYSES, CENSORING AND THE	41
PROBLEM OF COMPETING RISKS	
ENDPOINT DATA WITHOUT INFORMATION ON TIME TO	42
EVENT	
SUBGROUP ANALYSES	42
MODEL DISCRIMINATION AND RECLASSIFICATION	43
3.2.4 REPEATED MEASUREMENTS	44
TEMPORAL CHANGES IN BIOMARKER CONCENTRATIONS	44
WITHIN-SUBJECT REPRODUCIBILITY	45
3.3 THE STROBE GUIDELINES – ENHANCING TRANSPARENCY	45
4 SUMMARY OF THE RESULTS	47
4.1 PLASMA DIMETHYLGLYCINE AND RISK OF INCIDENT ACUTE	47
MYOCARDIAL INFARCTION IN PATIENTS WITH STABLE ANGINA	
PECTORIS (PAPER I)	
4.2 ELEVATED PLASMA DIMETHYLGLYCINE IS A RISK MARKER	49
OF MORTALITY IN PATINTS WITH CORONARY HEART DISEASE	
(PAPER II)	
4.3 THE PROSPECTIVE ASSOCIATIONS OF SYSTEMIC AND	51
URINARY CHOLINE METABOLITES WITH INCIDENT TYPE 2	
DIABETES (PAPER III)	
5 DISCUSSION	53

5.1 SUMMARY	53
5.2 METHODOLOGICAL CONSIDERATIONS	53
5.2.1 BIAS	53
SELECTION BIAS	53
INFORMATION BIAS	56
REGRESSION-DILUTION BIAS	58
5.2.2 CONFOUNDING	59
5.2.3 REVERSE CAUSALITY	60
5.2.4 MENDELIAN RANDOMIZATION – IMPROVING THE SHORTCOMINGS OF OBSERVATIONAL STUDIES?	63
5.2.5 GENERALIZABILITY	64
5.2.6 MULTIPLE COMPARISONS	64
5.2.7 SUBGROUP ANALYSES	65
5.2.8 MODEL DISCRIMINATION AND RECLASSIFICATION	66
5.2.9 BIOSAMPLE PROCESSING	70
5.2.10 STATISTICAL ANALYSES	70
5.3 ETHICAL CONSIDERATIONS	71
5.3.1 LEGAL ASPECTS	71
5.3.2 FINANCIAL ISSUES	72
5.4 DISCUSSION OF THE MAIN FINDINGS	72
5.4.1 PUTTING THE FINDINGS INTO AN EPIDEMIOLOGICAL CONTEXT	72
PAPERS I AND II	72
PAPER III	73
5.4.2 POSSIBLE MECHANISMS	74
A GENERAL DISCLAIMER	74
MITOCHONDRIAL FUNCTION AND ENERGY AND LIPID HOMEOSTASIS	74
RAMIFICATIONS TO METHYL METABOLISM, NUCLEOTIDE PRODUCTION, CELL TURNOVER AND EPIGENETIC REGULATION	78

CHOLINE METABOLITES IN THE URINE	79
CAN DIETARY FACTORS EXPLAIN OUR FINDINGS?	81
6 CONCLUSIONS	83
7 FUTURE PERSPECTIVES	85
8 REFERENCES	86
9 APPENDIX	110

1 INTRODUCTION

1.1 CARDIOVASCULAR DISEASE

1.1.1 DEFINITION, EPIDEMIOLOGY AND IMPLICATIONS

Cardiovascular disease (CVD) comprises all diseases of the circulatory system (i.e. heart and blood vessels). CVD related mortality has dropped in several countries, including Norway (Figure 1), during the last decades,¹ and this decline has largely been attributed to the identification and improvement of CVD risk factors.² Yet, CVD is still the number one cause of mortality globally, accounting for about one third of all deaths,¹ and ischemic heart disease (IHD) is considered the main cause of years lost due to illness and death.³ In Europe, about 4 out of every 10 deaths before the age of 75 years are due to CVD.⁴ Accordingly, in 2012, 13 010 and 4 852 deaths out of totally 41 913 deaths in Norway were attributed to CVD (International Classification of Diseases (ICD) 10 codes I00-I99) and IHD (ICD 10 codes I20-25), respectively, emphasizing CVD as the number one cause of death nationwide.⁵

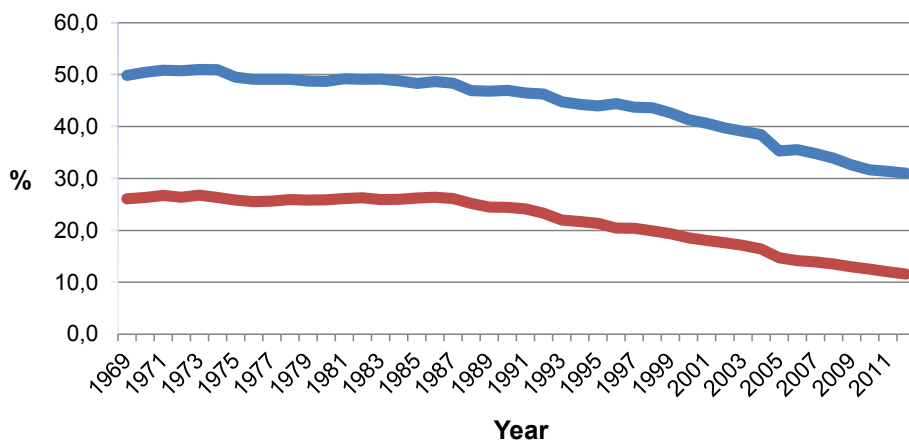


Figure 1. Death from cardiovascular disease (blue line) and ischemic heart disease (red line) as percentages of death from all causes in Norway, during the period 1969-2012.

Data obtained from Statistics Norway.⁵

Most incident CVD events do not immediately cause death; hence CVD has a major impact on overall morbidity. Reliable data on the occurrence of non-lethal CVD

among individuals in Norway has only recently emerged via the Norwegian Cardiovascular Disease Registry (NCDR),⁶ contrasting other Scandinavian countries which have had similar national registries for many years.^{7,8} On the other hand, The Western Norway Cardiovascular Registry (WENOCARD; <http://cvdnor.b.uib.no/wenocard>) has served as a precursor of the NCDR, by recording regional data on CVD hospitalizations in Western Norway public hospitals from 1972 and throughout 2006.⁹ Likewise, the Cardiovascular Disease in Norway (CVDNOR; <https://cvdnor.b.uib.no/>) project collected data on prevalent and incident CVD, as well as diabetes mellitus, from 42 Norwegian public hospitals in the period 1994-2009.

According to the NCDR, approximately 330 000 patients received at least one CVD diagnosis at Norwegian public hospitals in 2013, and almost 60% had incident disease.⁶ About 15 000 cases of AMI (ICD-10 I210-I229) were registered and over 23 000 admissions to public hospitals and out-patient clinics were classified as due to angina pectoris (ICD-10 I200-I209). These data correspond to previous reports from regional Norwegian surveys, indicating that the prevalence of non-fatal CVD was approximately 9% among subjects older than 40 years, and mostly related to IHD.¹⁰

Apart from individual human repercussions, CVD also has a major impact on societal health expenditures and lost productivity. According to a recent report from the European Heart Network and the European Society of Cardiology, the annual overall economic cost from CVD in Europe as per 2009 was estimated to approximately €196 billion. Of these expenses, IHD constitutes about 1/3, with over 60% being related to direct health care expenditures and costs related to lost productivity.¹¹

1.1.2 PATHOPHYSIOLOGY AND CLINICAL PRESENTATION OF IHD

Most cases of IHD are caused by atherosclerosis in the coronary arteries (i.e. coronary artery disease (CAD)). The pathophysiological mechanisms involve deposition of apolipoprotein (apo) B and cholesterol containing low-density lipoproteins (LDLs) inside the vessel wall at one or several locations in the coronary artery tree. This is accompanied by low-grade cellular inflammation, smooth muscle-cell proliferation, and fibrosis, forming an atherosclerotic plaque. The plaque may grow both outwards

(positive remodeling) and into the vessel lumen (negative remodeling). An inward growing plaque can potentially obliterate blood flow, giving rise to clinical features of coronary heart disease (CHD) on physical or emotional exertion. Such lesions can be visualized by coronary angiography and quantitatively graded according to luminal obstruction. Symptoms of CHD may vary, but typically include chest pain (i.e. angina pectoris), and result from insufficient blood supply to the distal myocardium to accommodate increased need of oxygen (i.e. ischemia).¹² In patients with stable angina pectoris (SAP) due to reversible ischemia, symptoms will typically arise on a certain level of physical or psychological exertion, and relieved once the precipitating factor is removed or when taking medications that lessen myocardial oxygen expenditure. However, coronary blood flow may be obliterated, either gradually by a growing plaque itself, or abruptly by a thrombus generated on the plaque surface (i.e. atherothrombosis), to such a degree that myocardial cell death occurs distal to the lesion. Myocardial necrosis due to ischemia is the very definition of an acute myocardial infarction (AMI),¹³ and puts the patient at risk of both immediate and long-term complications, such as systolic heart failure and cardiac arrest. It is noteworthy, however, that even the presence of non-obstructing coronary atherosclerotic lesions carries a significant risk of atherothrombotic events.¹⁴

The Framingham Heart Studies have established high age, male gender, dyslipidemia, smoking and hypertension to predict CVD events with an accuracy of approximately 80% among presumably CVD naïve subjects.¹⁵ Although causality between established risk markers and endpoints has been heavily debated, the hypothesis of LDL-cholesterol (LDL-C) deposition in atheromatosis has been strengthened by the favorable results on CVD events and mortality from LDL-C lowering therapy by statins.¹⁶ Statins act primarily by inhibiting the rate-limiting step of the endogenous cholesterol synthesis, thereby enhancing the hepatic clearance of circulating LDLs by increasing the amount of LDL receptors on surface of the hepatocytes.¹⁷ Furthermore, intravascular ultrasound studies have suggested reductions in focal atherosclerotic plaque volume, as well as altered plaque composition by intensive statin treatment,¹⁸ indicating that statin therapy might affect the atherosclerotic plaque more directly. This highlights that atherosclerosis is dynamic,

and that reverse cholesterol transport, as facilitated by high-density lipoproteins (HDLs), might also be influenced by statin treatment. Although some argues against their alleged pleiotropic effects,¹⁹ statins may have beneficial effects on CVD beyond those of lipid modification.²⁰ On the other hand, statins do not eliminate atherosclerosis, as an incident major cardiovascular event will still occur in a substantial proportion of patients who receive statin therapy.¹⁶ This residual risk may be because of inadequate improvement of lipid status,²¹ but also due to the presence of other known²¹ and unknown factors influencing risk of atherothrombosis.

Taken together, CVD in general, and CHD in particular, are major determinants of morbidity and death, with huge individual and socioeconomic costs. Intense focus on preventive measures has likely reduced the burden of disease; however, there is still considerable residual risk, warranting research into novel pathophysiological pathways.

1.2 DIABETES MELLITUS

1.2.1 DEFINITION, EPIDEMIOLOGY AND IMPLICATIONS

The term diabetes mellitus comprises diseases characterized by hyperglycemia. Although highly simplified, diabetes can generally be divided into states of either a primary and absolute insulin deficiency (type 1 diabetes), or relative insulin insufficiency due to insulin resistance and pancreatic β -cell dysfunction (type 2 diabetes (T2D)), as well as gestational diabetes and rarer etiologies primarily arising from disorders of the pancreas.²² The current thesis will mainly deal with T2D.

Diabetes is on the rise worldwide, and the International Diabetes Federation has estimated the global prevalence among adults to 8.3% (382 million people) in 2013, with an expected rise of more than 50% in the next 25 years. Most prevalent and incident cases are due to T2D, and the increase will likely have a particularly strong impact on younger people in developing countries.²³

The Norwegian Diabetes Register estimates the prevalence of diabetes among adults in Norway to be approximately 4%,²⁴ which is comparable to our neighboring Scandinavian countries.^{25,26} Accordingly, the Norwegian Prescription Database reported that 170 000 Norwegian citizens used antidiabetic drugs (The Anatomical

Therapeutic Chemical (ATC) classification system and the Defined Daily Dose A10) in 2014,²⁷ constituting about 3% of the current total Norwegian population. In addition, several patients with T2D do not receive pharmacological antidiabetic treatment, and the International Diabetes Federation states that almost half of all prevalent cases of T2D in the world are likely not diagnosed.²³ Hence, the real prevalence of T2D in Norway is almost certainly much higher than officially claimed.

According to the Global Burden of Disease project, diabetes ranks 14 in terms of causing disability-adjusted life years worldwide,³ and diabetes was estimated to account for about 11% of total global health care expenditures in 2013.²³ In Norway, a diagnosis of diabetes was registered in 2.5% of the consultations in general practice in 2014,²⁸ and 2.4% of the admissions to public somatic hospitals and out-patient clinic visits in 2014.²⁹ In addition, diabetes may be the underlying cause or accompanying condition of a range of other illnesses, and in particular CVD.²³ Diabetes therefore has a huge impact on individuals, public health care and the society as a whole, and motivates research into novel pathophysiological mechanisms in order to prevent disease development and its complications.

1.2.2 PATHOPHYSIOLOGY AND CLINICAL PRESENTATION OF T2D

The pathophysiology of T2D is complex and has yet to be fully delineated; however the main focus has been on insulin resistance, which is defined as the need of supernormal insulin excretion from pancreatic β -cells to keep blood sugar levels within the normal range,³⁰ and being strongly linked to increasing bodyweight and physical inactivity.³¹ However, the β -cells may eventually be unable to compensate for the insulin resistant state, hence hyperglycemia ensues.³¹ T2D pathophysiology is also associated with factors other than insulin resistance and β -cell failure per se, such as impaired pancreatic α -cell function, genetic and epigenetic regulation, factors related to the diet and the gastrointestinal system, lipotoxicity, and alterations in the kidneys, adipose tissue and the nervous system.^{31, 32}

The clinical presentation of diabetes varies and symptoms may have slow onset. Typical symptoms of hyperglycemia include polydipsia and blurred vision, whereas the inability to utilize glucose as an energy source may lead to fatigue and weight

loss;³³ however, many patients with T2D have little or no symptoms, contributing to the delay of diagnosis.

1.2.3 T2D AND CHD ARE STRONGLY RELATED

The clinical implications of T2D are severe, and excessive mortality is closely related to CVD,³⁴ and in particular increased risk of CHD.³⁵ T2D and atherosclerotic CVD (ACVD) seem to act additively in terms of adverse prognosis, and T2D may be regarded as a CHD and stroke equivalent.³⁶ Conversely, patients diagnosed with CHD are at higher risk of developing T2D, but it is not known whether this association is explained by already impaired glucose homeostasis inherent to CHD in the first place.³⁷

T2D is further linked to atherosclerosis by multiple common risk factors, which may be related to insulin resistance and hyperinsulinemia,^{31, 38} but could also involve other elements, such as alterations in amino acid metabolism.^{39, 40} In addition, several pharmacological treatment options for CVD (i.e. beta-blockers, thiazides, niacin and statins) are associated with adverse glucose control, as well as increased risk of incident T2D,⁴¹ and the use of several glucose lowering drugs (including thiazolidinediones and sulfonylureas) have been associated with higher cardiovascular risk.^{42, 43}

The close associations between T2D and CVD suggest that these two non-communicable diseases may not only share some common soil,⁴⁴ but also have complementary relationships yet to be resolved. Furthermore, although the diabetes epidemic has been suggested to curtail the steadily declining CHD mortality rates observed during recent decades,² there are evidence that increasing rates of obesity and diabetes have not proportionately impacted CVD mortality rates.⁴⁵ Although this discrepancy may be influenced by methodological shortcomings,⁴⁵ actions to identify specific diabetic phenotypes particularly prone to CHD seem pertinent.

1.3 THE CHOLINE OXIDATION PATHWAY

1.3.1 A BRIEF OVERVIEW

Choline is a quaternary ammonium compound which probably enters the mitochondrion via a specific carrier-mediated transport mechanism.⁴⁶ It is metabolized into betaine via a two-step reaction, catalyzed by the enzymes choline dehydrogenase (CHDH) and betaine aldehyde dehydrogenase (BADH), respectively (Figure 2).⁴⁷ Betaine diffuses out into the cell cytosol, and is demethylated into dimethylglycine (DMG), as it donates one methyl group to homocysteine (Hcy), which is converted into methionine. This irreversible reaction is catalyzed by the enzyme betaine-homocysteine-*S*-methyl transferase (BHMT).⁴⁸ DMG then enters the mitochondrion and is demethylated into sarcosine and glycine via the enzymes dimethylglycine dehydrogenase (DDH) and sarcosine dehydrogenase (SDH), respectively, with tetrahydrofolate (THF) being the methyl group acceptor in both reactions. Whereas DMG can only be produced from betaine, sarcosine may also be obtained by the methylation of glycine in the cell cytosol, catalyzed by glycine-*N*-methyltransferase (GNMT).⁴⁹

1.3.2 CHOLINE AND BETAINE CAN BE PRODUCED ENDOGENOUSLY OR OBTAINED FROM THE DIET

In the human body most choline is present as the phospholipid phosphatidylcholine (PC),⁴⁷ which can be converted to free choline by phospholipases.⁵⁰ PC production takes place in the liver mainly from choline via the three-step Kennedy (CDP) pathway, or alternatively from phosphatidylethanolamine (PE) by the enzyme phosphatidylethanolamine *N*-methyltransferase (PEMT);⁵¹ however this endogenous production is insufficient for biological choline demands,⁴⁷ making choline an essential nutrient. According to the US Institute of Medicine, the average recommended daily choline intake is 550 mg and 425 mg/day for men and women, respectively,⁵² but the intake varies extensively, with certain populations probably having insufficient intakes.⁵³ Nordic authorities have not specifically implemented dietary choline intake in their current nutrition guidelines.⁵⁴ Among dietary factors explored in a Norwegian general population sample, only eggs and cholesterol intake

significantly predicted plasma choline, although various other food items are rich in choline, as well.⁵⁵

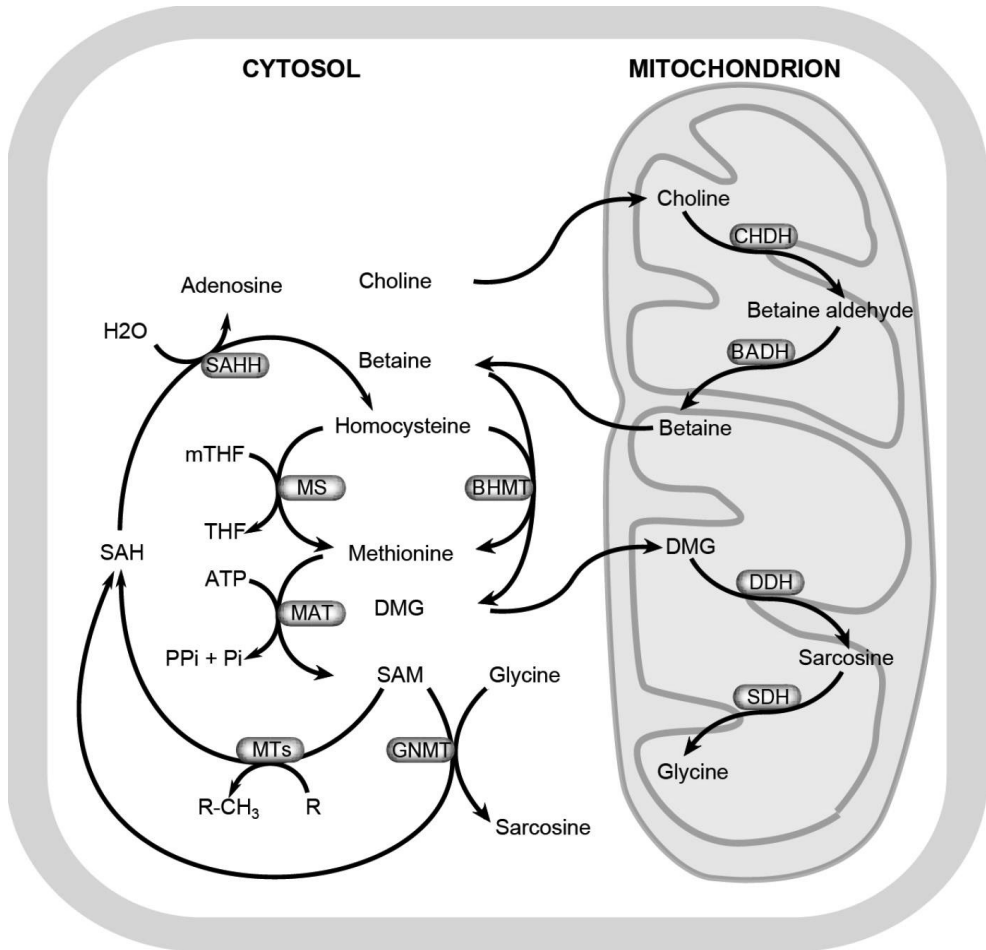


Figure 2. The coinciding choline and methionine cycle pathways.

BADH, betaine aldehyde dehydrogenase; BHMT, betaine-homocysteine-S-methyltransferase; CHDH, choline dehydrogenase; DDH, dimethylglycine dehydrogenase; DMG, dimethylglycine; GNMT, glycine-N-methyltransferase; MAT, methionine adenosyltransferase; MS, methionine synthase; MTs, various methyltransferases; SAH, S-adenosylhomocysteine; SAHH, S-adenosylhomocysteine hydrolase; SAM, S-adenosylmethionine.

Betaine may also be obtained directly through the diet, and plasma betaine has shown strong positive relations with the intake of high-fiber bread,⁵⁵ a common food

item in Nordic countries.⁵⁶ However, no recommendations on betaine intake currently exist. Besides, no population surveys on either choline or betaine intake have hitherto been carried out in the Nordic countries, thus representative intake and status of choline and related metabolites are largely unknown for subjects immediately comparable to those investigated in this thesis.

1.3.3 RAMIFICATIONS OF CHOLINE METABOLISM

Apart from serving as the source of downstream metabolites, choline is important for the synthesis of neurotransmitters, and PC is crucial in the assembly of cell membranes and lipoproteins,⁴⁷ as well as the production of bile.⁵⁰ Hence, choline species are found in various tissues, but downstream metabolism of free choline seem most abundant in the liver and the kidney,^{47, 50, 57} which also have highest activities of both CHDH⁵⁸ and BADH.^{59, 60}

CHDH is located to the inner mitochondrial membrane, and in humans the greatest CHDH activity has actually been found in the kidney, followed by the liver.⁵⁸ Moreover, CHDH uses flavin adenine dinucleotide (FAD) as a cofactor, and the electron is transferred to the respiratory chain via coenzyme Q10.⁵⁸

BADH is located both in the cell cytosol and inside the mitochondrial matrix.⁵⁹ It uses NAD⁺ (and NADP⁺) as a cofactor in the irreversible oxidation of betaine aldehyde,⁶¹ and is inhibited by its substrate, betaine aldehyde. BADH is found in several different tissues^{47, 62}, but betaine is mainly found in the liver and kidney of mammals.⁵⁷ This probably reflects the role of betaine not only as a methyl donor, but also as a regulator of cell volume (i.e. as an osmolyte), which is considered of particular importance in the extremely hypertonic environment of the renal cortex.^{47, 57}

As expected, the main tissue distributions are similar for betaine as for BHMT, a Zn-containing enzyme which makes up approximately 0.6-1.6% of the total hepatic protein content,⁴⁸ the vast amount highlighting the important role for betaine in liver metabolism. The BHMT pathway is involved in regulating levels of methionine and the non-protein amino acid, homocysteine (Hcy). It therefore tightly links betaine to folate metabolism, since methionine also can be formed via folate- and cobalamin-dependent remethylation of Hcy by methionine synthase (MS). Methionine is

important in protein synthesis, but also for the production of the ubiquitous methyl donor *S*-adenosylmethionine (SAM). SAM is necessary for a whole range of transmethylation reactions, including the production of creatine, PC and polyamines, as well as in DNA methylation and epigenetic regulation,⁶³ the latter referring to the modification of genetic expression without altering the genomic sequence per se.⁶⁴ The demethylation of SAM yields *S*-adenosylhomocysteine (SAH), which can be further hydrolyzed into Hcy, hence completing the so-called methionine cycle. Accordingly, the BHMT and MS pathways are complementary to each other. Although the latter pathway traditionally has received the greater attention regarding Hcy remethylation, BHMT might be more important in regulating Hcy levels than previously appreciated.^{48,65} The significance of the BHMT pathway in terms of methylation status has been emphasized by *in vitro* and *in vivo* studies where impaired BHMT flux leads to lower hepatic SAM levels.⁶⁵⁻⁶⁷

The relationship between the MS and the BHMT pathways is further illustrated by several studies suggesting that BHMT flux seems to be inversely related to folate status.^{48,68} BHMT activity is also inhibited by DMG and SAM, and insulin, growth hormone, thyroxin and testosterone, as well as hypertonicity may reduce BHMT activity and/or transcription.⁴⁸ Methionine also inhibits BHMT,⁶⁹ and it follows that BHMT is up-regulated when dietary intake of methionine is low and intake of labile methyl groups is high.^{70,71} However, dietary methionine excess may also up-regulate BHMT,⁷² reflecting dual roles of the BHMT pathway in terms of methionine and Hcy homeostasis.⁷³ In contrast, increased intakes of choline and betaine up-regulate BHMT flux, as do diabetes,^{74,75} as well as treatment with glucocorticoids.⁴⁸

Hcy is degraded via the vitamin B6-dependent transsulfuration pathway, which is initiated by the condensation of Hcy and serine, catalyzed by cystathionine- β -synthase (CBS). The transsulfuration pathway is essential for the making of cystathionine and cysteine, being pivotal in the production of the intracellular antioxidant glutathione, as well as for taurine and protein synthesis.⁶⁹ It follows that inborn errors of CBS lead to Hcy accumulation, and traditionally, betaine has been used in the treatment of such patients.⁷⁶ Not only does betaine supplementation increase the remethylation of Hcy into methionine, but increased BHMT flux also

stimulates the transsulfuration pathway,⁷⁷ most probably due to increased SAM levels which activate CBS.⁶⁹

DDH and SDH are vitamin B2-dependent enzymes⁷⁸, and assumed to have tight connections with the mitochondrial respiratory chain, which utilizes the electrons provided during DMG and sarcosine catabolism.⁷⁹ Methyl groups from DMG and sarcosine are transferred to THF, forming 5, 10-methylenetetrahydrofolate. Hence, folate availability may influence the metabolism of DMG and sarcosine, although experimental studies suggest that folate status does not determine the passage of electrons to the electron transfer proteins.⁸⁰ Besides generating methyl groups and electrons, DMG has no other known direct metabolic role, although DMG supplementation has been associated with altered immune responses,⁸¹⁻⁸³ as well as having potentially favorable effects on glucose and lipid homeostasis, oxidative stress, and lung vasculature in poultry.⁸⁴ Sarcosine may be a scavenger of excess methyl groups via increased GNMT flux, in particular among patients with impaired SDH.⁸⁵ Further, demethylation of sarcosine via SDH yields glycine, which is involved in glucose metabolism, the synthesis of nucleotides, proteins and antioxidants, as well as in cellular signaling.⁸⁶

The concentrations of DMG and sarcosine in tissues other than the liver and the kidneys have been investigated only to a limited degree, as most studies have reported on their blood or urinary levels. DMG was found in rectal cancer cells,⁸⁷ and sarcosine has been reported in prostate cancer cells⁴⁹ and in the rat cerebral cortex.⁸⁸ Nonetheless, studies in animals and/or humans indirectly suggest the presence of DMG and/or sarcosine also in various other tissues, due to their contents of DDH and/or SDH and/or GNMT (Table 1). Since DMG can only be obtained via the BHMT reaction, tissues containing DDH, but not BHMT, may therefore depend upon the import of DMG.

Determinants of circulating DMG and sarcosine concentrations have not been extensively investigated, but DMG levels increase according to higher intake of metabolic precursors and especially betaine,^{89, 90} as well as in patients with chronic renal disease.⁹¹ The minor A allele on the single nucleotide polymorphism (SNP)

BHMT 742 G>A (R239Q),⁹² is inversely related to plasma DMG,⁹³ although not associated with plasma total Hcy (tHcy) levels.⁹² Polymorphisms of the *sdh* gene influence plasma DMG levels as well,⁹⁴ and even genetic traits coding for impaired

Table 1. Tissue specificity of enzymes taking part in downstream choline metabolism

Enzyme		Tissue	Species
CDH	<i>Protein</i>	Liver and kidney ⁵⁸	Human
BADH	<i>Protein</i>	Liver ⁹⁵	Rat
		Kidney ⁹⁶	Pig
BHMT	<i>Protein</i>	Liver, kidney ⁴⁸	Human
		Pancreas ⁴⁸	Sheep
		Optical lens ⁴⁸	Rhesus monkey
DDH	<i>mRNA</i>	Liver, kidney, heart, brain, lung ⁹⁷	Rat
	<i>Protein</i>	Liver, kidney, lung, heart, spleen, brain ⁹⁷	Rat
SDH	<i>mRNA</i>	Liver, kidney, lung, thymus, oviduct, prostate, seminal vesicle, heart and brain. ⁹⁸	Rat
	<i>Protein</i>	Breast cancer tissue ⁹⁹	Human
		Liver, pancreas and adrenal gland. ⁹⁸	Rat
GNMT	<i>Protein</i>	Liver, kidney, pancreas, salivary gland, jejunum ¹⁰⁰	Various mammals, mainly rats.

BADH, betaine aldehyde dehydrogenase; BHMT, betaine homocysteine S-methyltransferase; CHDH, choline dehydrogenase; DDH, dimethylglycine dehydrogenase; GNMT, glycine N-methyltransferase; SDH, sarcosine dehydrogenase.

methylenetetrahydrofolate dehydrogenase (MTHFD1) activity are associated with higher hepatic and plasma DMG concentrations, possibly due to a greater demand for one-carbon units obtained through the choline oxidation pathway.¹⁰¹

1.3.4 CHOLINE AND BETAINES METABOLISM IN THE GUT

– THE FORMATION OF TRIMETHYLAMINE-N-OXIDE

Dietary choline, and to a lesser extent betaine, may be transformed into trimethylamine (TMA) by bacteria in the gut, and TMA can be further oxidized into trimethylamine N-monoxide (TMAO) by hepatic flavine monooxidase 3 (FMO3).¹⁰² Systemic TMAO levels may therefore reflect choline intake, and the gut microbiome might also be regarded as a choline and betaine metabolizing organ.

1.3.5 CHOLINE AND BETAINE IN RELATION TO HEPATIC LIPID METABOLISM

Both choline and betaine are involved in hepatic lipid metabolism, and especially in the export of lipids from the liver. Not only is free choline used in the production of PC, but PEMT uses three molecules of SAM in the alternative production of PC from PE, thus highlighting the need of adequate methylation status for this reaction to occur. The BHMT pathway may also be more directly related to hepatic lipid metabolism as suggested by specific concomitant genetic transcriptions of hepatic *bhmt* and *apob* mRNA.¹⁰³ Accordingly, BHMT induction by betaine supplementation resulted in increased production and secretion of VLDL from the liver; however systemic apoB and triglyceride levels were not affected.¹⁰³ This was possibly due to increased hepatic clearance via the LDL receptor,¹⁰³ suggesting an overall increased transport of apoB containing lipoproteins and cholesterol between the liver and extrahepatic tissues. In humans high-dose betaine supplementation has been associated with a relative increase in serum LDL-cholesterol, and supplementation with both choline and betaine modestly increases serum triglycerides.¹⁰⁴ Accordingly, increased availability of choline and betaine has been inversely related to non-alcoholic fatty liver disease (NAFLD),⁴⁷ which is strongly associated with T2D and considered an independent CVD risk factor.¹⁰⁵

1.3.6 HOMOCYSTEINE AS A CARDIOVASCULAR RISK FACTOR – A LONG STORY MADE SHORT

Since the last half of the 20th century there has been a great focus on the strong relationship between high levels of tHcy (the sum of all Hcy species) in blood and urine, and risk of occlusive CVD (Figure 3). The associations are based upon firm data from numerous epidemiological and experimental studies, suggesting hyperhomocysteinemia to be a causal risk factor for ACVD beyond those identified by the Framingham Heart Studies.¹⁰⁶ Moreover, the solution to the problem seemed straightforward, as well as inexpensive and tolerable, because the administration of folic acid and/or vitamin B6 lowers circulating tHcy by increasing Hcy remethylation or degradation. Hence, several trials with such therapy in order to reduce CVD risk

were launched in the late 1990s and early 2000s; however, despite substantial tHcy lowering effects by the interventions, no improvement in prognosis were observed,¹⁰⁷ nor did later Mendelian randomization studies immediately support elevated tHcy as a causal risk factor for ACVD.^{108, 109} In fact, treatment with folic acid and vitamin B12 in the WENBIT was associated with accelerated growth of atherosclerotic plaques.¹¹⁰ Consequently, the limelight has faded on the “homocysteine theory” (Figure 3), indicating that increased plasma tHcy is merely an epiphenomenon of other pathways responsible for increased CVD risk.

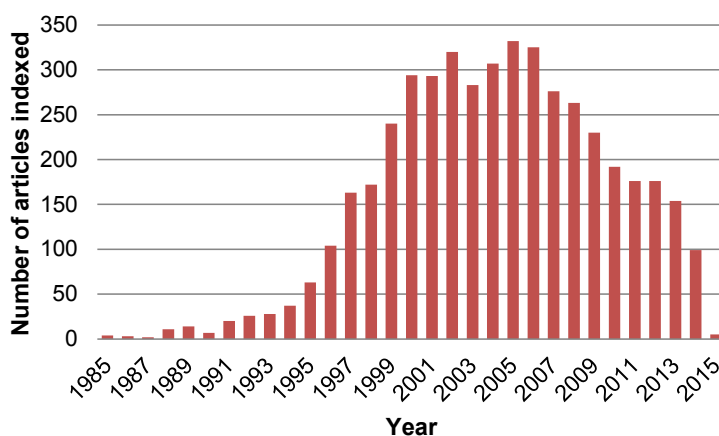


Figure 3. Yearly number of articles indexed in the Pubmed/Medline database by the medical subheadings “homocysteine” + “cardiovascular disease” during the last 30 years. Data obtained from www.pubmed.com

1.3.7 CHOLINE METABOLISM IN RELATION TO CVD AND DIABETES

Small population studies have related higher systemic choline levels to adverse cardiovascular risk, and even suggested that choline in whole blood and serum may yield incremental prognostic information beyond that conferred by conventional risk markers among patients with suspected acute coronary syndrome (ACS; i.e. either unstable angina pectoris or AMI).¹¹¹ Accordingly, observational data from a Norwegian general population sample showed that high plasma choline and low betaine were related to an adverse CVD and T2D risk profile, including high BMI,

elevated blood pressure and serum triglycerides, and low serum high-density lipoprotein-cholesterol (HDL-C).¹¹² On the other hand, a study among about 500 patients with a recent ACS found U-shaped associations between plasma and urine betaine with risk of incident AMI and hospitalizations for heart failure, respectively.¹¹³ Notably, the same study also reported as a secondary finding that increased plasma DMG levels were related to CVD events and mortality.¹¹³ A small study found higher urine DMG, but not betaine, in patients with than without premature vascular disease,¹¹⁴ and another reported the BHMT 742 G>A minor allele to be inversely related to the extent of angiographic CAD among elderly patients.⁹² In summary, these observations indicate that higher circulating choline and DMG levels might reflect adverse CVD risk, whereas the risk relationships with betaine seem a bit more complex. In addition, a recent series of preclinical and human studies has suggested plasma TMAO to play a role in ACVD, and that reducing choline intake might be a means of risk reduction,^{102, 115} despite earlier general population based studies reporting no associations between intakes of neither choline nor betaine and CVD risk.^{116, 117}

Circulating choline levels according to diabetes status are not well established;⁴⁷ however a recent small cross-sectional study from China suggested lower serum choline and betaine concentrations in association with prevalent diabetes and diabetic complications.¹¹⁸ An inverse association between systemic betaine and existing T2D has been supported by some,¹¹⁸⁻¹²⁰ but not all⁵⁷ studies, and experimental investigations have suggested that increased betaine availability may reduce adiposity and improve insulin resistance.⁵³ A recent Swedish study found robust, but relatively weak, associations between genetic traits coding for lower plasma DMG and increased risk of developing diabetes during long-term follow-up.⁹⁴ Among a US male general population lower serum sarcosine levels were observed among those with than without T2D.¹²¹

Higher dietary intakes of PC were related to increased risk of incident T2D,¹²² and higher plasma TMAO levels have been observed among patients with than without T2D.^{120, 123} This potentially links choline intake and the gut microbiome to T2D risk,

and the relationship is possibly also modified by the inhibiting actions of insulin on FMO3.¹²⁴

Substantially higher urinary betaine levels have been observed among patients with T2D.⁵⁷ Our group showed that urine betaine was highly correlated with urine DMG and sarcosine, and that urine betaine predicted new-onset diabetes within the WENBIT follow-up period;¹²⁵ however, no longer-term prospective studies of urinary choline metabolites on incident T2D risk have hitherto been reported.

1.3.8 OPEN RESEARCH QUESTIONS

In summary, there are preclinical and epidemiological evidence of significant alterations in choline metabolism according to both CVD and T2D. However, most prospective epidemiological studies have so far focused on the CVD risk associated with systemic choline and betaine concentrations, in addition to circulating TMAO levels, and relatively little attention has been paid to relationships with DMG and sarcosine. Moreover, with the exception of betaine, any prospective relationships between choline metabolites in the urine and incident disease have yet to be investigated. There is therefore a need for a more comprehensive assessment of several choline metabolites in relation to CVD and T2D in large populations.

2 AIMS OF THE THESIS

The primary goals of the project were to investigate the relationship between plasma DMG levels and risk of incident AMI and mortality, as well as exploring potential associations between circulating and urinary choline metabolites with incident T2D. Secondary aims were to investigate potential improvements in risk prediction and to assess temporal trends of the biomarkers investigated.

SEPARATE PROJECT AIMS

2.1 Paper I: To investigate the association between plasma DMG and incident AMI among patients with suspected SAP.

2.2 Paper II: To explore the relationships between plasma DMG with all-cause, cardiovascular and non-cardiovascular mortality in two independent cohorts of patients with suspected SAP and AMI, respectively.

2.3 Paper III: To study the associations between systemic and urinary choline metabolites with incident T2D among patients with suspected SAP.

3 MATERIALS AND METHODS

3.1 PATIENT POPULATION

3.1.1 THE WENBIT, NORVIT AND WECAC COHORTS

The WENBIT (ClinicalTrials.gov Identifier: NCT00354081) and NORVIT (ClinicalTrials.gov Identifier: NCT00266487) were large, Norwegian, randomized, controlled trials initiated and carried out in the late 1990's and early 2000's to investigate the clinical effect of tHcy lowering therapy with folic acid + vitamin B12 and/or vitamin B6 against placebo in a 2x2 factorial design. The WENBIT¹²⁶ enrolled 3090 patients from the Western Norway, the majority (98.6%) having angiographically verified significant CAD. The patients were recruited from Stavanger and Haukeland University Hospitals in Stavanger and Bergen, Norway. During the study enrollment from April 1999 to April 2004, both hospitals carried out percutaneous coronary intervention (PCI) whereas only Haukeland University Hospital carried out open heart surgery. The multicenter NORVIT¹²⁷ studied 3749 patients hospitalized for an AMI in all Norwegian health regions, and enrolled patients from December 1998 to March 2002. The patients in the NORVIT were recruited from both local, regional and university hospitals, and did not routinely undergo coronary angiography, as is recommended by current guidelines.¹²⁸

Participants in the NORVIT provided blood specimens at the baseline visit, and study visits after 1 year and at the end of study, whereas participants in the WENBIT provided blood samples at baseline, and at study visits after 1-3 months, 1 year and at the end of study. Most WENBIT study participants additionally provided urine samples at baseline and after 1 year of follow-up.

During the WENBIT study enrollment period, 2119 patients who underwent cardiac catheterization at Haukeland University Hospital, but who were not enrolled in the WENBIT trial, were included in an extended cohort of patients. These patients provided baseline blood and urine samples, and were followed up according to clinical endpoints, but did not attend later study visits. Together with WENBIT participants, these patients totaled 5209 patients, and made up the Western Norway Coronary Angiography Cohort (WECAC) (Figure 3).

In the current work, we chose to focus on WECAC patients with suspected or verified SAP (n=4164), thereby excluding 1045 patients with other indications for angiography (ACS, n=519; valvular disease, n=331; other indications, n=195). Our rationale was to study a rather homogenous group of patients, being less prone to changes in metabolites and biomarkers secondary to the acute phase (e.g. inflammation), as well as having less propensity to adapting life-style changes¹²⁹ which could potentially influence risk relationships between baseline values and outcome. Besides, WENBIT patients with ACS had their biosampling done partly before, during, or even after coronary angiography and intervention, without such information being registered in the database, making it possible that such procedures might have influenced biomarker status.

A total of 42 WECAC patients with suspected SAP rejected extended follow-up, and were thus censored in terms of endpoints beyond 31 Dec 2006. We did not have access to follow-up data on patients who moved out of Norway.

3.1.2 FOLLOW-UP AND CLINICAL ENDPOINTS

The WENBIT and the NORVIT cohorts were granted long-term follow-up on incident cardiovascular events, and all-cause, cardiovascular and non-cardiovascular mortality, extending the original trial durations. The rest of the patients making up the WECAC were granted inclusion in the combined WENBIT-NORVIT cohort after approval from the Regional Ethics Committee (Regional Ethics Approval number 2010/1880), which also approved the collection of information on incident diabetes from the CVDNOR project (Regional Ethics Approval number 2013/2324). The Regional Ethics Committee also specifically approved the projects included in the current work; “*DMG og coronarsykdom*” (Regional Ethics Approval number 2010/1881-8) and “*Betaine and choline excretion in urine*” (Regional Ethics Approval number 2010/1747-8).

All endpoints were linked to each patient’s unique Norwegian 11-digit person identification number. Incident AMI (paper I) in the WECAC was classified according to American and European guidelines of 2000,¹³⁰ but also included patients diagnosed with either “sudden cardiac death” or “sudden death” (ICD-10 codes I46 and R96, respectively),¹³¹ being in line with the definition used in previous Scandinavian

epidemiological surveys.¹³² Endpoint information was obtained from the WENOCARD and validated by the WENBIT endpoint committee as previously described.¹²⁶ Procedure-related AMI was defined as those occurring within 24 hours after coronary revascularization, and were not included.

Mortality data in both the NORVIT and the WECAC cohorts (paper II) were obtained from the Death Registry at Statistics Norway (<http://www.ssb.no>). The information on incident AMI and mortality, including time to events, was collected throughout 31 December 2006.

Information on incident T2D was retrieved from the CVDNOR project, assessing public hospital discharge diagnoses ICD E110-E119 throughout 31 December 2009. These latter endpoints were not validated further, nor did we obtain reliable time-to-event data. This was mainly because we had access to detailed hospitalization data for only a limited number of patients. Moreover, for the majority of cases (~90 %), T2D was recorded as a secondary diagnosis and not the main reason for contacting the health care system.

3.1.3 LABORATORY ANALYSES

Baseline study sampling was carried out usually 1-3 days prior to the angiographic procedure among patients enrolled at Haukeland university hospital, whereas patients recruited from Stavanger university hospital usually had their biosampling done at the day of cardiac catheterization. Plasma samples were immediately prepared and stored in tubes either containing ethylenediaminetetraacetic acid (EDTA) or sodium citrate, whereas serum samples were stored in gel-containing tubes. Baseline urine samples were collected by the patients at home on the morning of angiography. Biosampling at later study visits was carried out as described above, and the patients were not instructed to fast. All biosamples were frozen at -80°C until thawed and analyzed by laboratory staff blinded to the clinical outcomes of the patients. The results from analyses on plasma TMAO first became available during the spring of 2015, and

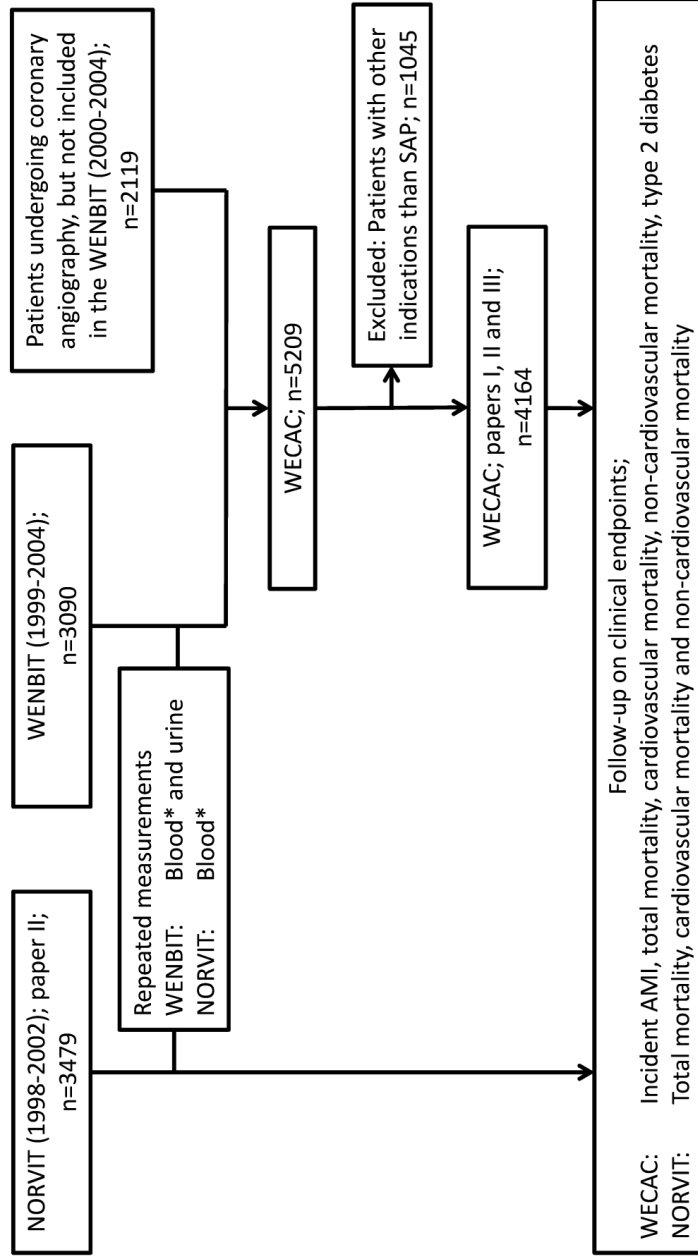


Figure 4. Flowchart depicting the inclusion and exclusion of study patients.

AMI indicates acute myocardial infarction; NORVIT; Norwegian Vitamin Trial; SAP, stable angina pectoris; WECAC, Western Norway Coronary Angiography Cohort; WENBIT; Western Norway B-Vitamin Intervention Trial.

*Plasma and serum samples

therefore we were not able to include this variable in papers I and II.

Routine biochemical analyses were carried out by the local laboratories in each recruiting hospital of the WECAC and NORVIT cohorts. Estimated glomerular filtration rate (eGFR) was calculated according to the Chronic Kidney Disease Epidemiology Collaboration formula.¹³³ Professor Rolf K. Berge and his research group at the University of Bergen analyzed serum apolipoprotein (apo) A1 and apo B 100 on the Hitachi 917 and 912 systems (Roche Diagnostics, GmbH, Mannheim, Germany), respectively. Study specific blood and urinary analyses were carried out by Bevitall As, Bergen, Norway (<http://www.bevital.no>), using automated methods based on mass spectrometry for investigating large numbers of samples, but requiring small individual sample volumes (Table 2).¹³⁴ Urine choline metabolites were measured by a modified liquid chromatography-tandem mass spectrometry (LC-MS/MS) method.¹³⁵

Table 2. Laboratory assays used for analyses of metabolites in plasma/serum and urine

Metabolite	Method
Plasma	
choline	LC-MS/MS ^{135, 136}
betaine	LC-MS/MS ^{135, 136}
dimethylglycine	LC-MS/MS ^{135, 136}
trimethylamine- <i>N</i> -monoxide	LC-MS/MS
total homocysteine	GC-MS ¹³⁷
riboflavin	LC-MS/MS ¹³⁸
5'-pyridoxal phosphate	LC-MS/MS ¹³⁸
Serum	
sarcosine	GC-MS ¹³⁴
folate	Microbiological assay ¹³⁹
cobalamin	Microbiological assay ¹⁴⁰
Urine	
choline	LC-MS/MS ¹³⁵
betaine	LC-MS/MS ¹³⁵
dimethylglycine	LC-MS/MS ¹³⁵
sarcosine	GC-MS/MS ¹⁴¹

DMG indicates dimethylglycine; GC-MS/MS, gas chromatography mass spectrometry; LC-MS/MS, liquid chromatography-tandem mass spectrometry.

Serum in citrate tubes had to be used instead of EDTA plasma when assessing sarcosine status in blood, as the EDTA tubes already contained a significant amount of sarcosine.¹⁴² The Bevital laboratory also measured serum C-reactive protein (CRP) by an ultrasensitive immuno- matrix-assisted laser desorption/ionization (MALDI)-MS method, with a detection limit of 0.17 mg/L.¹⁴³ Data on serum CRP, lipid fractions, plasma glucose and HbA1c were only available in the WECAC.

The BHMT single nucleotide polymorphism (SNP) BHMT 742 G>A,¹⁴⁴ as well as levels of glycosylated hemoglobin (HbA1c)¹⁴⁵ were assessed from EDTA whole blood samples, using matrix-assisted laser desorption/ionization-time-of-flight (MALDI-TOF) mass spectrometry.

In a subset of 1343 WENBIT participants, of whom the majority were fasting, we also obtained baseline serum insulin and C-peptide from citrate samples, using a solid phase, two-site chemiluminescent immunometric assay (Immulite 2000) from Siemens Healthcare Diagnostics. However, since EDTA plasma is usually used for insulin measurements, and the amount of citrate within each tube slightly varied, we compared EDTA and citrate samples from 40 patients, and found that the insulin concentrations were highly correlated (Spearman's rho 0.95; P<0.0001) (Figure 5); hence, the data obtained from citrate samples was extrapolated into those of EDTA plasma by linear mean regression modelling using a regression coefficient (β) of 1.494. Among fasting, non-diabetic patients with valid insulin and plasma glucose measurements, we calculated the computer-based homeostatic model assessment (HOMA2)¹⁴⁶ of insulin resistance, beta-cell function and insulin sensitivity. The HOMA2 parameters were used in articles II and III.

We also obtained serum levels of high-sensitive cardiac troponin T (hs-cTnT) among 4070 patients with SAP in the WECAC, using a Modular E170 from Roche Diagnostics, with a lower detection limit of 3ng/l. This parameter was not yet available when writing article I, nor was it available among NORVIT patients.

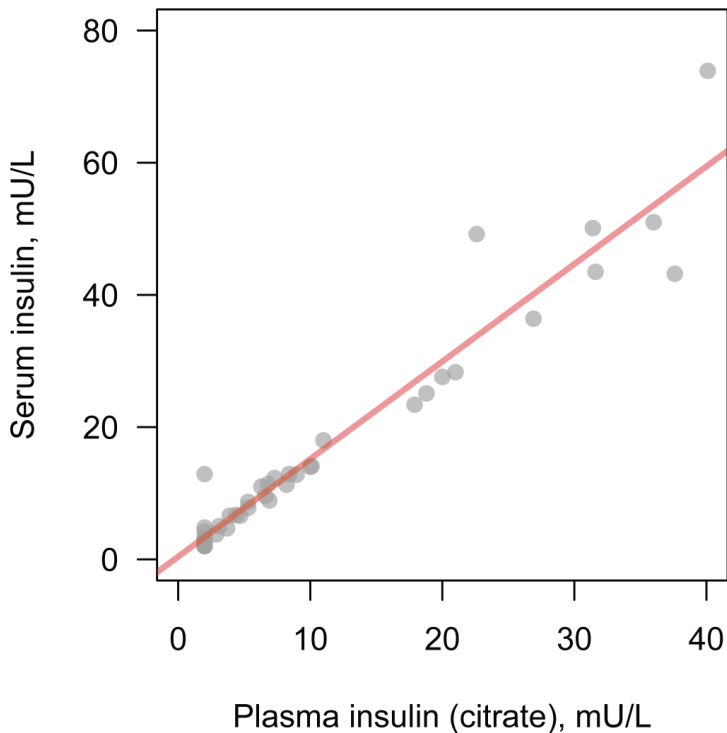


Figure 5. The relationship between insulin in citrate plasma and serum.

3.1.4 DATA ON DIETARY CHOLINE AND BETAININE INTAKE

Among WENBIT participants, 2412 patients provided information on dietary intake of macro- and micronutrients by completing a food frequency questionnaire (FFQ).¹⁴⁷

The daily intakes of total energy, choline and betaine were estimated among 1939 patients,¹⁴⁸ after excluding those with either particularly low (men and women <3300 kJ/day and <3000 kJ/day, respectively) or high (men and women >17500 kJ/day and > 15 000 kJ/day, respectively) estimated energy intake.

3.2 STATISTICAL METHODS

3.2.1 SOFTWARE

For all statistical analyses, we used SPSS for Windows versions 18-21 (SPSS Inc. Released 2009. PASW Statistics for Windows, Version 18.0. Chicago: SPSS Inc.; IBM Corp. Released 2010-2012. IBM SPSS Statistics for Windows, Version 19.0-

21.0. Armonk, NY: IBM Corp.), the free, open-source statistical software R for Windows versions 2.15.0-3.0.2 (The R Foundation for Statistical Computing, Vienna, Austria; packages *nlme*, *Hmisc*, *ppcor*, *mgcv*, *segmented*, *PredictABEL*, and *ICC*) and SPSS Sample Power Version 2.0 (SPSS, Inc., 2000, Chicago, IL, www.spss.com). Graphics were obtained by the various R packages, and slightly modified according to the standards of each journal by using the free image editing software Inkscape for Windows version 0.48 (www.inkscape.org).

3.2.2 BASELINE DATA

In articles I and II we assessed potential linear relationships between plasma DMG quartiles (i.e. categories according to the 25th, 50th and 75th percentiles) and relevant clinical, anamnestic and biochemical parameters. Traditionally, linear associations between continuous variables have been explored by linear least squares mean regression models and analysis of variance (ANOVA) for continuous and categorical variables, respectively. However, treating the mean as an average measure is problematic, especially in terms of biological data, due to their often extensively right-tailed (non-normal) distribution. In the case of least squares mean linear regression non-normal distribution may also severely influence the regression line.¹⁴⁹ Most commonly, such right-skewed continuous data have been log-transformed before analyses, followed by reporting trends across groups according to their geometric means or back-transformed values. One problem is, however, that such values are quite arbitrary and do not communicate well with the reader. Moreover, regarding linear trends across ordinal categories, ANOVA modeling assumes that data are normally distributed within each group. One way around this problem could be to apply the non-parametric Kruskal-Wallis one-way of variance test. However, this method does not allow for adjusting for covariates. Therefore, we used semi-parametric quantile regression modeling, as provided by the R package *Quantreg*. Quantiles refer to the number of equally distributed parts of a cumulative distribution. In short, quantile regression does not imply assumptions regarding distributions, and allows for covariate adjustment. The method may explore linear trends between any quantile (i.e. percentile) of two continuous variables; however we focused on the

trends according to the median (50 percentile) in all three papers. Quantile regression was also utilized when assessing differences in continuous variables according to two groups (e.g. patients with or without T2D). However, in paper III, we explored the relationships between incident T2D and not only one, but all metabolites downstream the choline oxidation pathway. Thus, it was not feasible to carry out baseline trend analyses according to each one of the independent variables, but rather making an overview over various interrelationships between choline metabolites and other variables of interest (i.e. correlation matrix). Once again, we wanted to adjust the correlations for covariates and we had to overcome the problem of non-normal distributions, the latter violating the assumptions when carrying out Pearson's correlations. We therefore solved this by performing partial ranked Spearman correlations, using the R package *ppcor*.

For categorical and ordinal dependent variables, we performed trend analyses by binary or ordinal logistic regression models, respectively.

3.2.3 ENDPOINT ANALYSES

Time-to-event analyses, censoring and the problem of competing risks. For articles I and II the occurrence of AMI or death was well defined, also in terms of time to events, the latter being of importance when following patients for several years. Hence, we used the well-established method of Cox proportional hazard modeling, where we censored patients who did not experience an event during follow-up, or for whom such information was not available. However, censoring of patients is problematic, because it does not allow us the information on what may have happened to those patients afterwards. In addition, some endpoints may only occur once, for example death. This generates the problem of competing risks, where one endpoint changes the possibility of experiencing another. It has been stated that not properly accounting for competing risk may overestimate the risk association.¹⁵⁰ Thus, in article II, we constructed subdistribution hazard regression models, using the R package *crr*,¹⁵¹ treating cardiovascular and non-cardiovascular death as competing events.

Cox proportional hazards modeling is quite robust. However, the assumption of proportional hazards is crucial, and was assessed by inspecting log-log plots according

to categorical variables, and calculating and testing Schoenfeld residuals, the latter by using the *cox.phz* function in the R package *survival*.

Endpoint data without information on time to event. Assessing the relationship between baseline and dichotomous endpoint data may also be carried out using binary logistic regression, preferably in analyses of endpoints with a pre-defined follow-up period (for example 30 day in-hospital mortality), or when exploring the relationship in studies without (reliable) information on time to events. We had prospective data on all end-points in our studies, but since T2D is not generally characterized by an abrupt onset of symptoms,³³ we chose to assess risk associations by using binary logistic regression in article III.

Subgroup analyses. The association between an independent and a dependent variable can differ between certain population groups, and may be assessed by performing analyses in each subgroup of interest. Random variation in associations between the independent and the dependent variable may occur across subgroups; therefore the potential effect modification should be tested formally. This is usually performed by including an interaction product term between the subgroup variable (the potential effect modifier) and the independent variable of interest in the model, and should also include the interaction term's lower order components.¹⁵²

All subgroup analyses were carried out and reported according to current recommendations.¹⁵³ Subgroups were obtained from predefined categories (e.g. gender, smoking status or study treatment), or created according to the median value of continuous variables, the latter to make the groups as equally sized as possible. We performed interaction analyses using the same methodology as described above, paying particularly attention to subgroups of established CHD risk factors and baseline status of biological parameters related to the choline oxidation pathway. The former may help identify subgroups in which a novel biomarker may have a more pronounced effect on risk prediction. The latter suggests metabolic mechanisms involved in associations between outcome and metabolite status, as differences in the latter may arise from alterations in the production, catabolism or tissue clearance of the

metabolite, or of their combination.

Model discrimination and reclassification. Model discrimination was assessed by calculating the Harrells *C*-statistic, which equals the area under the receiver-operating curve (ROC-AUC; Figure 5). Broadly speaking, the *C*-statistic will provide a probability score between 0.5 and 1 on how well the model actually predicts patients with and without the outcome, with the former and latter values corresponding to a useless coin-toss and a perfect prediction, respectively.

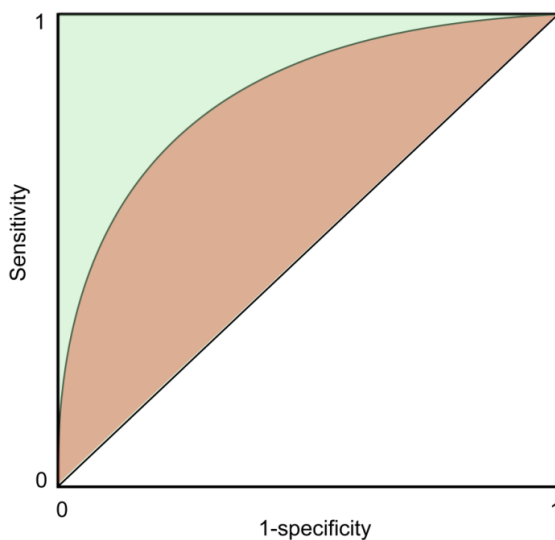


Figure 5. The area under the receiver-operating curve.

The diagonal line illustrates a model with no discriminative value; the brown area illustrates a model with intermediate discriminative predictive value; the green area (also covering the brown area) illustrates an optimal discriminative model.

However, once a few strong and robust predictors are included in the model, the adding of new variables seldom provides any substantial improvement in the *C*-statistic.¹⁵⁴ Thus, other, more sensitive, measures of model discrimination and reclassification have been proposed, with the most notable and utilized being the integrated discrimination index (IDI) and the net reclassification improvement (NRI).¹⁵⁴ The IDI can be defined as the difference in discrimination slopes between models with and without the biomarker(s) of interest; hence it compares the ability of models to discriminate cases from non-cases. The NRI will provide the percentage of patients that are being correctly reclassified to new risk categories, penalized by the percentage of patients that are wrongly reclassified. This estimation is performed both

for patients who do and do not experience an event.

$$NRI_{\text{total}} = NRI_{\text{event}} + NRI_{\text{non-event}}$$

When considering CVD naïve patients, such risk categories for cardiovascular mortality are typically divided according to 0-5%, 5-10%, 10-20% and above 20% of risk of the event within a certain period of time (usually 10 years), as illustrated by the Systematic Coronary Risk Evaluation Project (SCORE) promoted by the European Society of Cardiology.¹⁵⁵ Allocation of patients into a particular risk category will potentially guide the clinician in the decision making of initiating certain preventive interventions. Therefore, the NRI may provide a more clinically useful metric than both the *C*-statistic and the IDI. However, among patients with known (atherosclerotic) CVD, no established risk categories exist, mostly because such patients generally receive quite similar medical treatment (i.e. a statin and aspirin) and life-style advice. In fact, it is advocated against creating new risk categories under conditions where none already exist.¹⁵⁶ Hence, the category free, or continuous, NRI ($NRI > 0$) may serve as a better measure of improvement in reclassification. The $NRI > 0$ equals the net improvement in patients who are either being correctly up- or down-classified on a continuous (category-free) scale from 0 to 100%.¹⁵⁶ Based upon the comparison with Cohen's *d*, it has been suggested that small, medium and large effect sizes correspond to $NRI > 0$ of $< 20\%$, $\sim 40\%$ and $> 60\%$, respectively.¹⁵⁷

3.2.4 REPEATED MEASUREMENTS

Temporal changes in biomarker concentrations. As described earlier, not all WENBIT and NORVIT study patients attended each study visit beyond baseline, and there were also inequalities according to the time spans between sampling points. We could therefore not simply use the well-known and widely used repeated measures analysis of variance (rANOVA), or its derivatives, when analyzing longitudinal biomarker data. Instead, we used the more flexible method of mixed (-effects) linear regression modeling, provided by the R package *nlme*. As opposed to rANOVA, such modeling may include subjects with missing data points, and allows for uneven time-

spans between study visits during follow-up. Moreover, rANOVA modelling requires homogeneous variances in the variables used, whereas in linear mixed modelling, such heterogeneity may be easily adjusted for. Finally, as opposed to the rANOVA method, there is no need for correcting for multiple testing when using the linear mixed modelling approach in the setting of more than one time interval.¹⁵⁸ Data from all study visits in the WENBIT and NORVIT were used for the repeated measures analyses in paper I and II. In paper III we used the baseline and 1-year measurements in WENBIT, since urine sampling was only performed among a few on other study visits. Study treatment effects were explored by including an interaction product term between time and study treatment group in the model.

Within-subject reproducibility. The coefficient of reliability (CoR) is also known as the one-way random effects intraclass coefficient (ICC) and according to Rosner¹⁵⁹ it is defined as

$$\text{CoR} = \text{ICC} = \sigma_{\alpha}^2 / (\sigma_{\alpha}^2 + \sigma_{\epsilon}^2),$$

where σ_{α}^2 denotes the between-subject variance and σ_{ϵ}^2 the within-subject variance. Hence, a CoR (ICC) approximating 1 will imply very low within-subject variance. CoRs (ICCs) of 0.4-0.75 and ≥ 0.75 are considered fair to good, and excellent, respectively.¹⁵⁹ In paper I and II we used linear mixed modelling when calculating crude CoRs; however in paper III we used the R package #ICC, which also provided a 95% confidence interval together with the ICC estimate.

3.3 THE STROBE GUIDELINES – ENHANCING TRANSPARENCY

The vast majority of epidemiological studies stem from observational data. However, it is imperative that studies originating from observational data, as for any other sources for that matter, have transparent methodology in order to highlight strengths and weaknesses. The Strengthening the Reporting of Observational Studies in Epidemiology (STROBE) statement¹⁶⁰ was launched in 2007 as a joint attempt from several authors and medical journals to improve such transparency. The statement

contains a checklist including information to be reported in an observational study, and has been widely endorsed by a number of scientific journals.¹⁶¹ The STROBE initiative has also been extended, and currently comprises specific guidelines on the reporting on several types of observational studies, including the STROBE: Molecular Epidemiology (STROBE-ME),¹⁶² which was applied in articles I-III.

4 SUMMARY OF THE RESULTS

4.1 PLASMA DIMETHYLGLYCINE AND RISK OF INCIDENT ACUTE MYOCARDIAL INFARCTION IN PATIENTS WITH STABLE ANGINA PECTORIS (PAPER I)

Arterioscler Thromb Vasc Biol. 2013;33:2041-8

Among the 4156 eligible patients with SAP, 72.0% were men and the median (5-95 percentiles) age was 62 (44-78) years. About 1/3 were active smokers, approximately half had hypertension, 40.3% had experienced a previous AMI and 11.8% had self-reported diabetes. Median (5-95 percentiles) plasma DMG was 4.1 (2.6-7.3) $\mu\text{mol/L}$. Plasma DMG levels were higher among non-fasting patients, and positively related to male gender, current smoking, higher age, hypertension, previous CVD, CRP, the use of beta-blockers and ACEI and/or ARBs, and circulating levels of most one-carbon and choline metabolites, as well as to plasma methylmalonic acid (MMA). Plasma DMG was inversely related to eGFR, serum total cholesterol and apo A1, in addition to the BHMT 742 G>A minor A allele.

During a median (5-95 percentiles) follow-up time of 4.6 (1.6-6.8) years, 343 (8.3%) of the patients experienced an AMI, and the incidence was positively related to increasing plasma DMG quartiles (P for log-rank < 0.001). In crude Cox regression models, the HR (95% CI) for AMI was 2.43 (1.78-3.31; P<0.001) when comparing the 4th vs. the 1st DMG quartiles. Adjusting for age, gender and fasting status somewhat attenuated the risk estimate (HR (95% CI) 1.95 (1.42-2.68); P<0.001), but further adjustment for serum apo A1 and apo B100, diabetes, smoking, and hypertension had only minor influence on the risk associations. Particular strong relationships between plasma DMG and risk of AMI were observed among non-smokers and patients with serum triglycerides or apoB100 < median (P for interaction \leq 0.03).

Adding plasma DMG to the multivariate model improved model discrimination (ΔC -statistic (95%CI) 0.012 (0.001-0.022); P=0.04), and improved reclassification.

In WENBIT patients we observed increasing plasma DMG throughout the study for the group as a whole, but not among patients receiving folic acid + vitamin B12 (with or without vitamin B6). Among patients randomized to placebo treatment, the

CoR for plasma DMG was 0.93 and slightly lower when the variable was log-transformed.

We did not report on the association between plasma DMG and incident AMI in NORVIT; however, when carrying out such analyses in retrospect, we observed a positive association between plasma DMG and AMI also in this cohort, although the relationship was somewhat weaker (Table 3). The association were similar when adjusting for plasma tHcy (HR [95% CI] for the 4th vs. 1st quartile 1.25 [1.00-1.56]; P=0.05), but attenuated when accounting for eGFR, serum total cholesterol, smoking, diabetes and hypertension (HR [95% CI] 1.14 [0.90-1.43]; P=0.28).

Table 3. The relationship between plasma dimethylglycine and incident acute myocardial infarction among patients in the Norwegian Vitamin Trial, n=3733

Plasma DMG, quartiles	Unadjusted		Adjusted for age and gender	
	HR (95% CI)	P	HR (95% CI)	P
1	Ref		Ref	
2	1.07 (0.85-1.35)	0.55	1.02 (0.81-1.28)	0.87
3	1.20 (0.96-1.49)	0.11	1.06 (0.85-1.33)	0.61
4	1.61 (1.30-1.98)	<0.001	1.30 (1.04-1.61)	0.02

CI indicates confidence interval; DMG, dimethylglycine; HR, hazard ratio.

4.2 ELEVATED PLASMA DIMETHYLGLYCINE IS A RISK MARKER OF MORTALITY IN PATIENTS WITH CORONARY HEART DISEASE (PAPER II)

Eur J Prev Cardiol. 2015;22(6):743-52

The 4150 patients with SAP eligible from the WECAC were generally similar to those included in paper I; hence baseline characteristics and the relationships between baseline plasma DMG and various clinical and biochemical parameters did also not differ to any particular degree from those reported in the previous chapter. In addition, we observed a strong positive relationship between plasma DMG and serum hs-cTnT, and inverse associations between plasma DMG and indices of insulin sensitivity.

The 3733 eligible patients with AMI from the NORVIT cohort had median (5-95 percentiles) age of 63 (44-81) years, and 73.9% were males. As compared to SAP patients, the NORVIT participants were older, more often smokers and had lower eGFR, but had less often preexisting CVD, hypertension or diabetes.

Among patients with AMI baseline plasma DMG was positively related to male gender, age, current smoking, hypertension, serum creatinine, previous CVD, plasma choline and one-carbon metabolites, and the use of beta-blockers and ACEIs and/or ARBs. Plasma DMG was negatively related to eGFR, plasma riboflavin, serum folate and the BHMT 742 G>A minor A allele.

Median (5-95 percentiles) follow-up times were 4.7 (2.8-6.8) and 7.0 (1.1-8.8) years among SAP and AMI patients respectively. Among SAP patients, 308 (7.4%) died, whereas 772 (20.7%) of the patients with AMI died. In both cohorts, the risk of all-cause mortality was positively related to increasing plasma DMG quartiles (P for log-rank<0.001). When comparing the 4th to the 1st DMG quartiles, the crude HRs (95% CI) for all-cause death were 2.52 (1.78-3.57); P<0.001 and 2.80 (2.27-3.44); P<0.001 among patients with SAP and AMI, respectively. Adjusting for age and gender attenuated the risk estimates (1.72 (1.21-2.46) and 1.76 (1.42-2.18)); however, the associations were essentially unaltered in the multivariate models. Plasma DMG was more pronouncedly associated with cardiovascular than non-cardiovascular death, and similar results were obtained when performing competing risk analyses.

Subgroup analyses showed trends towards stronger risk associations between

plasma DMG and all-cause and cardiovascular mortality among SAP patients with serum TG or apoB100 < median.

Adding plasma DMG to the multivariate model improved or trended to improve model discrimination and reclassification for all-cause and cardiovascular mortality, particularly among patients with AMI.

Similar to what was observed in article I, AMI patients who were randomized to either placebo or treatment with vitamin B6 alone experienced a rise in plasma DMG during follow-up; however no changes in plasma DMG were observed for patients receiving folic acid + vitamin B12 (with or without vitamin B6). The CoR for plasma DMG among AMI patients allocated to placebo was 0.72.

4.3 THE PROSPECTIVE ASSOCIATIONS OF SYSTEMIC AND URINARY CHOLINE METABOLITES WITH INCIDENT TYPE 2 DIABETES (PAPER III) *Submitted, January 2016.*

For baseline analyses, 4070 SAP patients were eligible, of whom 71.9% were men, and the median (25-75 percentiles) age was 62 (55-70) years. Patients with T2D (n=449 (11.0%)) had higher plasma choline and TMAO, lower plasma betaine and serum sarcosine, and higher urine choline, betaine, DMG and sarcosine than nondiabetic patients. Higher plasma choline and serum sarcosine, and lower plasma betaine concentrations were correlated with a generally more adverse T2D risk profile. Plasma DMG was positively related to indices of insulin resistance and CRP. Plasma TMAO was positively related to indices of insulin resistance and plasma choline, plasma DMG and serum sarcosine. Most choline metabolites in urine were positively related to an adverse risk profile of T2D.

The levels of all metabolites, except for urine betaine, increased during 1 year of follow-up among WENBIT patients allocated to placebo treatment. Treatment with folic acid + vitamin B12 (with or without vitamin B6) augmented the increase in plasma choline and betaine, whereas plasma and urine DMG and urine sarcosine were lowered, when compared to placebo. Vitamin B6 alone, on the other hand, increased plasma DMG, but dampened the increase in serum sarcosine and urine choline, as compared to placebo treatment. We did not observe any temporal alterations in overall plasma TMAO, also according to WENBIT study treatment.

A total of 3261 patients without registered T2D at baseline were followed for median (25-75 percentiles) 7.5 (6.4, 8.7) years, during which 233 (6.4%) patients were registered with incident T2D. As compared to those who did not develop T2D, patients who did develop T2D had lower baseline plasma betaine, whereas urine betaine and sarcosine were higher. In univariate logistic regression models, plasma betaine was inversely, while urine betaine, DMG and sarcosine levels were positively associated with incident T2D (ORs per 1 SD (95%I) 0.78 (0.69-0.89), 1.27 (1.11-1.45), 1.23 (1.07-1.42), and 1.31 (1.14-1.50), respectively). Adjusting for potential confounders and other T2D risk factors only slightly altered the risk estimates. We also performed a stepwise backwards elimination logistic regression, including all choline metabolites

being associated with incident T2D in univariate analyses. The selection was determined by the Akaike's information criterion, and left only plasma betaine and urine sarcosine of the choline metabolites in the final model. When including plasma betaine or urine sarcosine, the multivariate model discrimination improved, as did the reclassification of patients according to the $NRI > 0$.

The ICCs (95% CIs) for plasma betaine and urine sarcosine among patients randomized to placebo were 0.62 (0.56-0.66) and 0.69 (0.64-0.74), respectively. Similar ICCs were obtained among patients receiving vitamin B6 only treatment, whereas treatment with folic acid + vitamin B12 (with or without vitamin B6) weakened the test-retest reliability for both metabolites.

5 DISCUSSION

5.1 SUMMARY

The current thesis is based on data from either or both of two independent cohorts of patients with suspected or verified CHD, and the aims were to investigate the associations between plasma DMG and risk of incident AMI and mortality, the relationships between choline metabolites and incident T2D, any improvement in model risk prediction and reclassification for relevant biomarkers, and temporal trends in choline metabolites according to B-vitamin intervention.

The studies show that plasma DMG is a predictor of incident AMI among patients with suspected SAP, and predicts all-cause and cardiovascular mortality among patients with both stable and unstable CHD. We also report that lower plasma betaine and higher urine betaine, DMG and sarcosine are associated with new-onset T2D among patients with SAP. Once adjusted for age, gender and fasting status, all associations were essentially independent of traditional risk factors and potential confounders. Moreover, plasma DMG, as well as plasma betaine and urine sarcosine, may improve risk prediction of the respective endpoints. We also extend previous preclinical studies, by showing that treatment with high-dose B-vitamins influences the concentration of several choline metabolites in blood and urine.

5.2 METHODOLOGICAL CONSIDERATIONS

5.2.1 BIAS

The term “bias” refers to a systematic flaw in study design, and comprises a number of potential errors in obtaining the data on which a study is based.¹⁶³

Selection bias. Selection bias concerns the selection of participants included in the study, thereby influencing the generalizability of the results. Since SAP is often diagnosed and treated without diagnostic angiography, the WECAC population may be regarded as a selected group and could possibly be at different risk of adverse events than SAP patients not admitted for angiography. However, once admitted to tertiary centres for diagnostic evaluation, the vast majority of SAP patients (both with and without angiographically verified CAD) were included in the WECAC during the

recruiting period, thereby strengthening the generalizability of the study results, as compared to only investigating WENBIT participants.

We did not have information on hospitalized AMI patients who were not included in the NORVIT; however, the applicability of the NORVIT data for patients hospitalized with AMI in general is strengthened by the nation-wide multi-centre study design, including primary, secondary and tertiary hospitals.

Most patients in both cohorts had information on systemic levels of choline, betaine and DMG; however, data on serum sarcosine and urine choline metabolites were only available in subpopulations of the WECAC. Thus, a selection bias by excluding patients with missing data was not likely in papers I-II, but may have been present in paper III, although the large study size probably attenuated any such effect. This problem could be assessed by further investigating potential differences between included and excluded patients, and in hindsight I admit that it would have been prudent to do so. On the other hand, when performing multivariate analyses in paper II, we used the statistical technique of multiple imputations to also include patients with one or more missing covariates. This did not lead to any alterations in the results, suggesting that excluding a relatively small number of patients was less likely to confer any serious selection bias.

Repeated measurements were not undertaken in WECAC patients not included in the WENBIT. It is, however, unlikely that WENBIT study treatment would have affected biomarker status otherwise among these patients, as only minor differences in baseline choline metabolites levels were observed among participants and non-participants in WENBIT among the entire cohort for SAP patients in WECAC (Table 4). Yet, we cannot exclude that WENBIT study treatment may have influenced the risk estimates, although such treatment did not statistically significantly alter the results in survival analyses in relatively small groups. In addition, the WENBIT did not randomize patients in strata of clinical presentation; hence the subgroup of SAP patients were most likely not allocated to study treatment in a truly randomized fashion. It would therefore have been more appropriate to investigate not only the temporal trends in choline metabolites according to study treatment, but also to

Table 4. Baseline characteristics among patients in the Western Norwegian Coronary Angiography Cohort according to enrolment into the Western Norwegian B-vitamin Intervention Trial

	WECAC SAP patients		P*
	Not WENBIT	WENBIT	
Male gender, n (%)	946 (59.5)	2050 (79.7)	<0.001
Age, years	62 (55-69)	62 (54-70)	0.59
Current smoking, n (%)	497 (31.2)	823 (32.0)	0.64
Diabetes, n (%)	190 (11.9)	306 (11.9)	1.00
Hypertension, n (%)	731 (45.9)	1215 (47.2)	0.44
Previous CVD, n (%)	755 (47.5)	1648 (64.0)	<0.001
BMI, kg/m ²	26.0 (23.7-28.7)	26.5 (24.5-29.0)	<0.001
HbA1c, %	6.4 (5.8-7.1)	5.8 (5.1-6.6)	<0.001
Plasma glucose, mmol/L	5.6 (5.0-6.6)	5.6 (5.1-6.6)	0.45
eGFR,	88 (74-98)	92 (82-100)	<0.001
CRP, mg/L	1.83 (0.92-4.00)	1.74 (0.85-3.42)	0.002
Lipid parameters			
total cholesterol, mmol/L	5.0 (4.3-5.8)	4.9 (4.2-5.7)	0.04
LDL cholesterol, mmol/L	3.0 (2.4-3.8)	2.9 (2.4-3.6)	0.001
HDL cholesterol, mmol/L	1.30 (1.03-1.60)	1.20 (1.00-1.42)	<0.001
triglycerides, mmol/L	1.42 (1.04-2.04)	1.55 (1.10-2.20)	<0.001
apo A1, mg/dL	1.36 (1.19-1.55)	1.26 (1.10-1.43)	<0.001
apo B100, mg/dL	0.90 (0.75-1.08)	0.85 (0.71-1.02)	<0.001
Extent of CAD at angiography			
0-vessel disease	764 (48.0)	282 (11.0)	<0.001
1-vessel disease	236 (14.8)	730 (28.4)	
2-vessel disease	241 (15.1)	687 (26.7)	
3-vessel disease	350 (22.0)	874 (34.0)	
Systemic one-carbon and choline metabolites, µmol/L			
tHcy	10.8 (8.9-13.2)	10.3 (8.6-12.3)	<0.001
tMet	27.7 (23.5-33.1)	25.9 (21.9-31.2)	<0.001
choline	10.0 (8.5-11.8)	9.5 (8.1-11.2)	<0.001
betaine	39.4 (32.1-48.1)	38.9 (32.0-47.5)	0.32
dimethylglycine	4.4 (3.6-5.4)	4.0 (3.3-4.8)	<0.001
sarcosine	1.5 (1.2-1.8)	1.5 (1.2-1.8)	0.10
Urine choline metabolites, mmol/mol creatinine			
choline	2.30 (1.71-3.17)	1.88 (1.38-2.55)	<0.001
betaine	7.84 (4.99-13.32)	7.28 (4.75-12.73)	0.02
dimethylglycine	3.14 (1.93-5.05)	3.25 (2.13-5.07)	0.04
sarcosine	0.13 (0.08-0.21)	0.14 (0.09-0.22)	0.008
B-vitamins			
riboflavin, nmol/L	11.7 (7.3-19.7)	11.0 (7.6-17.4)	0.07
folate, nmol/L	10.5 (7.8-15.6)	9.9 (7.3-14.5)	<0.001
cobalamin, pmol/L	433 (336-552)	337 (258-426)	<0.001

PLP, nmol/L	43.8 (30.8-66.2)	40.0 (29.1-56.0)	<0.001
Medications prior to the baseline visit, n (%)			
beta-blocker	1102 (69.3)	1955 (76.0)	<0.001
statin	995 (62.6)	2029 (78.9)	<0.001
aspirin	1189 (74.7)	2157 (83.8)	<0.001
ACEI and/or ARB	1102 (69.3)	1955 (76.0)	<0.001
Medications at discharge, n (%)			
beta-blocker	1010 (63.5)	2005 (77.9)	<0.001
statin	1054 (66.2)	2281 (88.7)	<0.001
aspirin	1083 (68.1)	2317 (90.1)	<0.001
ACEI and/or ARB	501 (31.5)	829 (32.2)	0.65
Clinical endpoints, n (%)			
AMI	132 (8.3)	212 (8.2)	0.99
mortality	153 (9.6)	156 (6.1)	<0.001
T2D†	67 (4.8)	168 (7.4)	0.002

ACEI indicates angiotensin converting enzyme inhibitor; AMI, acute myocardial infarction; ARB, angiotensin receptor blocker; BMI, body mass index; CAD, coronary artery disease; CRP, C-reactive protein; CVD, cardiovascular disease; eGFR, estimated glomerular filtration rate; HDL, high-density lipoprotein; LDL, low-density lipoprotein; apo, apolipoprotein; PLP, pyridoxal 5'-phosphate; tHcy, total homocysteine; tMet, total methionine which is the sum of methionine and methionine sulfoxide; T2D, type 2 diabetes.

*Kruskal-Wallis test for continuous and Chi square test for categorical variables

baseline biomarker status. On the other hand, such interaction analyses would certainly decrease statistical power.

Information bias. Overall, only a handful of patients in the WECAC and NORVIT withdrew their consent of long-term follow-up, making the proportion of patients lost to follow-up very low. Since we assessed the risk relationships between baseline choline metabolite status and the clinical endpoints, the primary aims of our studies were not vulnerable to further non-response from the participants. Moreover, the use of mixed linear modelling strengthened the validity of repeated-measures analyses, since data from patients not attending all study visits could be included.

We cannot rule out the possibility that some WECAC patients experienced an incident AMI outside the Western Norway Health region, thereby leading to misclassification of non-cases in paper I. Also, some cases of AMI may have been undiagnosed or not reported properly. On the other hand, it is not likely that risk associations should differ in such subjects as compared to those with endpoints

registered in the WENOCARD, thereby reducing the risk of bias. Incident AMI in paper I was defined according to the revised European Society of Cardiology criteria from 2000,¹³⁰ but also included sudden cardiac death (ICD-10 code I461) or other sudden unexplained death outside hospital (ICD-10 code R96)), being in line with definitions used in epidemiological surveys in Sweden and Denmark.¹³² Widening the definition of AMI as compared to a more stringent one including only ICD-10 codes I21-22 may thus have increased sensitivity but decreased the specificity of the endpoints in paper I.

Using the nation-wide Cause of Death Registry, the risk association observed between plasma DMG and the hard endpoint all-cause mortality in paper II was not prone to either misclassification or detection bias. We were not able to validate the classification of mortality due to cardiovascular or non-cardiovascular causes, and the reliability of such classifications has indeed been questioned,¹⁶⁴ although the stronger risk association between DMG and cardiovascular mortality strongly agrees with the results in paper I.

The risk relationships between choline metabolites and incident T2D in paper III certainly pose the greatest concerns regarding detection bias in the current thesis. First, many patients with T2D are not diagnosed at all.²³ Second, the diagnosis of incident T2D was dependent on discharge records from most, but not all, Norwegian public hospitals. It is plausible that patients with new-onset T2D, and being hospitalized, may have differed in several aspects from non-hospitalized or non-diagnosed cases. Third, we were not able to validate the T2D diagnoses for each patient. However, when only looking at WENBIT participants during the in-trial follow-up, similar risk estimates between choline metabolites and T2D were obtained as those for the whole study population during long-term follow-up. Since the WENBIT participants were closely monitored by interview, as well as clinical and biochemical assessment, it is not likely that the results in paper III are flawed due to detection bias.

Regression-dilution bias. Risk associations between once-only assessed biomarker status and the clinical endpoint are inclined to be influenced by errors concerning the risk marker assessment per se, including measurement flaws, as well as the short- and long-term variation in biomarker status within each study participant – the latter collectively termed “within-person reproducibility”. This error is coined the “regression-dilution bias”, because all measurements will tend to regress to less extreme values when performed repeatedly. Consequently one-time assessment biomarker assessment will underestimate (i.e. dilute) the true effect.¹⁶⁵ There have been several suggestions on how to overcome this problem, with perhaps the simplest solution being to directly correct the risk estimates for the ICC in a non-parametric manner (i.e. the corrected association equals the risk estimate/ICC).¹⁶⁶ It follows that when the ICC is relatively low, the real risk estimate will be proportionally higher, whereas a high ICC implies a low degree of within-person variability and a proportionately low chance of regression-dilution bias. From a clinical point of view, however, a biomarker not requiring validation (i.e. having a high ICC) will surely be the more useful.

In the current studies plasma DMG (paper I and II) and betaine (paper III), as well as urine sarcosine (paper III), had ICCs ranging from “good” to “excellent”, and comparing to ICCs reported for various other CHD biomarkers, such as serum total cholesterol (0.75) and serum CRP (0.54).¹⁶⁷ Similar ICCs to our findings have been reported for plasma betaine and DMG in other populations, and seem consistent across genders and different ethnical groups.¹⁶⁸ However, a somewhat lower ICC was reported for plasma DMG in the Nurses’ Health Study¹⁶⁹ and Lever et al observed ICCs over 8 weeks for plasma betaine and DMG of only 0.43 and 0.15, respectively, when investigating a small population of healthy young men.¹⁷⁰ No studies have hitherto reported on the ICC of urine sarcosine, but high ICCs for the highly correlated urine betaine and DMG have been described.¹⁷⁰

In all, our findings imply a relatively low risk of regression-dilution bias, favouring the use of the studies metabolites as clinical biomarkers.

5.2.2 CONFOUNDING

For a given risk association *confounding factors* may be defined, quoting Miettinen and Cook, as “extraneous factors that might be explanatory, partially or totally, of the magnitude of the estimate of the effect”.¹⁷¹ Opposite to randomized clinical trials (RCTs), associations from observational studies are prone to confounding; hence, it is not possible to conclude on causal inferences. We can try to minimize this problem by adjusting the statistical models for potential confounders; however residual confounding will essentially always be present, as we are not able to identify or obtain data on all possible confounding variables.

There exist no firm guidelines regarding selection of the most appropriate covariates in statistical models. In general, statistical textbooks advice against solely relying on stepwise inclusion or exclusion procedures based on statistical measures alone, but advocate the inclusion of biological sound covariates, as well. Baseline choline metabolites had statistically significant relationships with several other clinical and laboratory parameters, which could indeed serve as potential confounders. However, we chose not to adjust for all of them due to several reasons: First, including too many variables will inflate the regression model, attenuating power and making the estimate(s) less reliable (i.e. increasing the confidence interval(s)). Second, several parameters may be highly correlated, (for example urinary choline metabolites in paper III), or they may be redundant (for example the inclusion of serum LDL cholesterol, when serum total cholesterol is already in the model), introducing the problems of collinearity and singularity, respectively. A similar issue may arise when including variables that might reflect common biological mechanisms, as discussed with serum creatinine and plasma DMG in paper II, potentially leading to over adjustment.¹⁷² On the other hand, the weakening of a risk estimate when adjusting for a covariate might also suggest pathophysiological mechanisms relevant for the risk associations. We did not, however, assess these issues formally.¹⁷³ Third, it is difficult, if not impossible, to compare the performance of models not containing the same covariates. In CHD studies, it is usual to adjust for established Framingham risk factors, although, strictly speaking, these are extensively validated for CHD naïve patients only. Therefore, we chose to primarily adjust for such factors in paper I and II;

accordingly we adjusted for several traditional T2D risk factors and biological plausible confounders in paper III. In papers I and II, we created extended multivariate models, further adjusting for variables with biological plausible (as well as statistical) relationships with DMG, in order to address potential pathophysiological pathways. This was also done regarding the associations between choline metabolites and incident T2D in paper III, although they were not reported due to space limitations.

5.2.3 REVERSE CAUSALITY

This term refers to the situation where the disease itself has influenced biomarker status, potentially impacting the real risk relationship. For example, the estimated daily energy intake among patients with T2D in the WECAC was not different from that of patients without diabetes, although the former had significantly higher BMI. This could be due to under-reporting among T2D patients, but also because these patients had reduced their energy intake as a consequence of their disease. If we were to explore the risk relationship between BMI and T2D, this would surely attenuate the true association.

Reverse causality especially poses a problem in retrospective studies; hence our prospective study design is less prone to this error. Yet, a large proportion of the WECAC and NORVIT patients had established CVD, and we cannot exclude that the disease itself, its complications and/or medical therapy and lifestyle interventions somehow may have influenced plasma DMG levels. This is, however, not immediately supported by studies performed in other populations, reporting similar circulating DMG concentrations as those presented for the WECAC and NORVIT cohorts (Table 5). Also, the risk estimates in paper I were similar when excluding patients without significant CAD at angiography, indicating that reverse causality might not pose a serious limitation. In paper III the risk estimates were similar when excluding patients with possible, yet not diagnosed T2D. Nonetheless, patients identified with higher baseline risk of T2D by conventional measures may also have had life-style or other interventions performed that could influence the status of choline metabolites in blood and urine.

Table 5. Systemic and urine dimethylglycine levels in other populations

Author (year)	Population	N	Specimen	DMG, $\mu\text{mol/L}$	Method
Holm et al (2003) ¹³⁵	Healthy blood donors	60	Plasma		LC-MS/MS
	Fasting			median (25-75 pct)	1.7 (1.3-2.0)
	Non-fasting			median (25-75 pct)	2.1 (1.6-2.5)
Fredriksen et al (2007) ⁹³	General population	10 601	Plasma		LC-MS/MS
	Total			median (5-95 pct)	3.7 (2.4, 5.9)
	Men			median (5-95 pct)	4.0 (2.7- 6.3)
	Women			median (5-95 pct)	3.5 (2.3- 5.6)
Holm et al (2005) ¹⁷⁴	Healthy subjects (fasting)		Plasma		LC-MS/MS
	Total	500		median (10-90 pct)	3.1 (2.0-4.9)
	Men	208		median (10-90 pct)	3.4 (2.4-5.2)
	Women	292		median (10-90 pct)	2.8 (1.9-4.6)
Kirsch et al (2010) ¹⁷⁵	Healthy subjects (fasting)		Plasma		UPLC-MS/MS
	Men	24		median (10-90 pct)	3.1 (2.0-4.3)
	Women	20		median (10-90 pct)	2.1 (1.5-3.7)
	Men	24	Urine*	median (10-90 pct)	2.1 (0.6-7.0)
	Women	20		median (10-90 pct)	2.3 (1.3-4.7)
Melse-Boonstra et al (2005) ¹⁷⁶	Healthy subjects		Plasma		LC-MS/MS
	Men	182		median (10-90 pct)	3.4 (2.4-4.7)
	Women	126		median (10-90 pct)	3.0 (2.3-4.5)
Lever et al (2012) ¹⁷⁷	Patients 3 months		Plasma		HPLC

post ACS (fasting)			
Men	383	median (25-75 pct)	3.8 (2.7-5.2)
Women	148	median (25-75 pct)	3.4 (2.2-4.8)
Urine*			
Men	383	median (25-75 pct)	2.9 (1.5-5.9)
Women	148	median (25-75 pct)	2.4 (1.4-4.3)

ACS, acute coronary syndrome; GC-MS, gas chromatography mass spectrometry; HPLC, high-pressure liquid chromatography; LC-MS/MS, liquid chromatography-tandem mass spectrometry; SD, standard deviation; HPLC, high performance liquid chromatography; UPLC-MS/MS, ultra performance liquid chromatography-tandem mass spectrometry;

*Corrected for urine creatinine

5.2.4 MENDELIAN RANDOMIZATION – IMPROVING THE SHORTCOMINGS OF OBSERVATIONAL STUDIES?

One way of omitting the potential pitfalls of residual confounding, reverse causality and certain types of bias is to carry out so-called Mendelian randomization studies.¹⁷⁸ Such studies investigate whether inborn genetic traits affecting biomarker status also are associated with clinical outcomes, thereby bypassing the influence of potential environmental factors. In other words, the subjects investigated are randomized genetically already before they are born. In the era of ‘big data’ Mendelian randomization studies have been widely embraced, and questioned previously proposed causal relationships for several CHD biomarkers, such as CRP,¹⁷⁹ HDL-cholesterol¹⁸⁰ and tHcy.¹⁰⁸

As previously mentioned, genetic traits of both BHMT and DDH predict plasma DMG levels, and a properly conducted Mendelian randomization study using the BHMT 742 G>A data from the largely homogenous WECAC and the NORVIT populations could potentially shed more light on any causal relationship between DMG and adverse cardiovascular prognosis. We did, however, carry out logistic regression analyses, which did not indicate any relationships between the BHMT 742 G>A SNP and CVD or mortality risk in both articles I and II (Appendix I, Supplemental Tables IX and X; Appendix II, Supplemental Tables 10-15). This observation strengthens the assumption that elevated plasma DMG is not a causal risk factor for CVD per se.

Mendelian randomization studies are, however, not free of assumptions, and pose several important shortcomings. These include, but are not limited to, genetic pleiotropy, linkage disequilibrium and issues of statistical power, as discussed in the review by Hansen et al.¹⁸¹ Probably the most important limitation is pleiotropy, meaning that genetic traits may have effects other than those only captured by measuring immediate biomarker levels. Such effects have not, to my knowledge, been investigated for the BHMT 742 G>A SNP; however it has been reported that the SNP does not statistically significantly predict plasma tHcy,¹⁸² nor did we find any differences in either plasma tHcy, betaine or total methionine levels according to this SNP in neither the WECAC or the NORVIT populations (Table 6).

Allele frequencies were assessed according to the Hardy-Weinberg equation. The BHMT 742 G>A SNP was in Hardy-Weinberg equilibrium in the WECAC, but not in the NORVIT cohort. In the WECAC, we observed an approximately 5% decrease of plasma DMG per A allele (Table 4), whereas a decrease in plasma DMG only was observed among homozygotes for the variant allele in the NORVIT.

There was a low minor A frequency, and as we showed in paper I, the impact of the A allele on the endpoint therefore had to be very high if we should have been able to obtain a statistically significant relationship between the genotype and disease risk. This illustrates the problem of insufficient statistical power. In the case of genetic risk associations, this usually necessitates the use of large population samples, as the effects brought about by single genetic traits usually are quite small, being particularly relevant for multifactorial diseases, such as CVD.

5.2.5 GENERALIZABILITY

As discussed in section 5.2.1.1, the current study populations are considered fairly representative of hospital-admitted patients with suspected or verified stable and unstable CHD in present-day Norway. On the other hand, our study subjects were mainly middle-aged to elderly white men, making it not straightforward to extrapolate the results to populations including healthy individuals, as well as those with other age, gender and ethnic characteristics.

5.2.6 MULTIPLE COMPARISONS

The current work includes a large number of statistical analyses. The P-value tells us the probability of falsely rejecting the null-hypothesis on the basis of a proper statistical test (i.e. Type I error). By setting the significance level (α) at 0.05, an average of 1 in 20 comparisons will thus lead to a statistically significant finding just by chance alone. Consequently, the probability of making such a Type I error is high, even when performing a relatively low number of individual tests,¹⁸³ and statistical textbooks advice against drawing conclusions without adjusting for multiple comparisons. The most commonly used correction is the one proposed by the Italian mathematician Carl Emilio Bonferroni, where $\alpha = 0.05/m$, and m denotes the number

of hypotheses being tested. On the other hand, the Bonferroni correction is a very conservative adjustment, especially when a large number of correlating parameters are being assessed.¹⁴⁹ Also, some authors emphasize less on formal correction for multiple testing in post-hoc analyses in general, because observational research cannot be confirmative and must be considered hypothesis generating.¹⁸³ Although it could have been more prudent to have applied other, less conservative, correction methods,¹⁴⁹ we chose not to correct for multiple testing, which may pose a limitation on the interpretation of our results. When using mixed linear regression models, the problem of multiple comparisons was, at least to some extent, attenuated.

5.2.7 SUBGROUP ANALYSES

Post-hoc subgroup analyses always carry the potential of spurious findings. One of the most well-known examples is probably the one reported for the International Study of Infarct Survival (ISIS)-2 trial, where effects of study treatment were shown to significantly differ according to the astrological assignment of the patients.¹⁸⁴ All post-hoc (subgroup) analyses must therefore be considered hypothesis-generating only. This is particularly stressed in observational studies, where some authors actually have advocated against using formal statistical testing between and within subgroups at all.¹⁸⁵ Nonetheless, although being considered at the lower level of scientific methodological hierarchy, unexpected findings from observational studies have made huge impact on medical sciences, reminding us not to dismiss them on a default basis. For example, hardly anyone would reject the observations of the effect from *Penicillin* fungi on bacterial growth, just because they resulted from untidiness in Sir Alexander Flemming's research laboratory.¹⁸⁶ The crux is that observational findings must be further validated.¹⁸⁷

Investigating treatment effects according to subgroups may help focusing on patient populations gaining the most benefit from treatment, and could also identify patients who even may be harmed from a particular intervention. Consequently, this has the potential to improve the clinical applicability of results from RCTs, which tend to generalize treatment effects. These issues are also highly relevant for diagnosis and risk assessment in CHD, as has been shown for the use of serum cardiac troponins and

electrocardiograms among patients with suspected ACS,¹⁸⁸ to mention but a few. Notably, in the current growing era of personalized medicine it has been called for an even more advanced profiling of patients in terms of CVD risk, in order to further improve patient care and to cut costs.¹⁸⁹

Apart from their potentially immediate clinical utility, subgroup analyses in observational biomarker studies may also help elucidate novel pathophysiological mechanisms in an otherwise extremely complex system of biological pathways. This issue will be dealt with in more detail in later sections of the thesis.

In paper I, we found trends towards stronger associations between DMG and adverse prognosis among non-smokers as compared to smokers in WECAC patients; however, there were no signs of any effect modification by smoking in the NORVIT in paper II. Although having relatively similar baseline characteristics, including plasma DMG status, the NORVIT population had a substantially larger proportion of current smokers (paper II). One reason for the lack of any interaction by smoking in the NORVIT could therefore be that more patients may have quit smoking after having experienced an AMI, thereby diluting any interaction effect. On the other hand, median plasma levels of cotinine, a nicotine metabolite, actually increased throughout follow-up in the NORVIT (data not shown). Although patients might have been less likely to be smoking while being hospitalized, this indicates a low level of smoking cessation, although persistent cotinine levels could also be due to the substitution of combustible tobacco for nicotine replacement products or smokeless tobacco.

The subgroup findings according to lipid parameters will be dealt with in more detail in section 5.4.2.

5.2.8 MODEL DISCRIMINATION AND RECLASSIFICATION

We showed that plasma DMG may improve the risk assessment of AMI and mortality, and that plasma betaine and urine sarcosine may aid in the reclassification of patients at risk of T2D. These findings are, however, not sufficient to bring these biomarkers into everyday clinical use. First, the findings should be extensively validated in other populations, preferably different from the ones studied in papers I-III. Second, the effect sizes of the $NRI > 0$ and IDI were, at best, modest,¹⁵⁷ and not immediately easy

Table 6. The BHMT 742 G>A polymorphism in relation to baseline betaine, total homocysteine, dimethylglycine, and total methionine, incident acute myocardial infarction, mortality, and incident type 2 diabetes

	BHMT 742 G>A				P _{trend} *
	N	GG	GA	AA	
Plasma, μmol/L					
Betaine					
WENBIT	2427	39.3 (32.4-47.9)	38.3 (31.5-46.9)	40.8 (31.7-49.5)	0.53
NORVIT	3433	32.8 (27.5-39.8)	33.3 (27.2-40.4)	33.0 (26.5-38.5)	0.32
total homocysteine					
WENBIT	2427	10.2 (8.6-12.2)	10.3 (8.7-12.2)	10.3 (8.7-12.8)	0.36
NORVIT	3433	12.1 (10.1-14.7)	11.9 (10.0-14.8)	12.3 (10.3-15.3)	0.93
Dimethylglycine					
WENBIT	2427	4.1 (3.4-4.9)	3.9 (3.2-4.8)	3.7 (3.1-4.5)	<0.00001
NORVIT	3433	4.1 (3.4-5.1)	4.1 (3.3-4.9)	3.6 (3.1-4.5)	<0.00001
total methionine					
WENBIT	2427	26.2 (22.0-31.3)	25.6 (21.9-31.1)	27.0 (21.8-32.4)	0.47
NORVIT	3433	29.5 (25.1-34.1)	29.5 (24.8-34.1)	29.7 (25.3-34.1)	1.00
Urine, mmol/mol creatinine**	2049				
Betaine					
WENBIT	2427	7.4 (4.9-12.7)	7.1 (4.7-13.3)	6.4 (4.6-10.3)	0.10
NORVIT	3433	3.4 (2.2-5.2)	3.2 (2.1-5.0)	2.9 (2.0-4.4)	0.02
Dimethylglycine					
WENBIT	2427	111/1273 (8.7%)	81/993 (8.2%)	15/161 (9.3%)	0.89
NORVIT	3433	372/1899 (19.6%)	218/1258 (17.3%)	47/276 (17.0%)	0.10
All-cause mortality					
WENBIT	2427	86/1273 (6.8%)	58/993 (5.8%)	7/161 (4.3%)	0.18
NORVIT	3433	385/1899 (20.3%)	270/1258 (21.5%)	52/276 (18.8)	0.94
CVD mortality	2427				
WENBIT	2427	47/1273 (3.7%)	32/993 (3.2%)	6/161 (3.7%)	0.71

NORVIT	3433	237/1899 (12.5%)	155/1258 (12.3%)	32/276 (11.6%)	0.71
Non-CVD mortality					
WENBIT	2427	39/1273 (3.1%)	26/993 (2.6%)	1/161 (0.6%)	0.12
NORVIT	3433	148/1899 (7.8%)	115/1258 (9.1%)	20/276 (7.2%)	0.58
Incident T2D**†	2137	89/1124 (7.9%)	61/868 (7.0%)	11/145 (7.6%)	0.58

AMI indicates acute myocardial infarction; CVD, cardiovascular disease; NORVIT, Norwegian Vitamin Trial; WENBIT, Western-Norway B-vitamin Intervention Trial; T2D, type 2 diabetes.

*Linear trends across groups tested by median linear and logistic regression for continuous and categorical dependent variables, respectively.

**WENBIT participants only

†Patients free of diabetes at baseline.

to communicate, because they do not tell us whether the improvements translate into any significant clinical meaning. This especially concerns the lack of information on patients being reclassified away from the group(s) with intermediate risk, as can be obtained by the category-based NRI. Third, and perhaps most importantly,¹⁹⁰ the current observational studies are insufficient to suggest that patients should be treated differently only on the basis of biomarker status alone. This is especially important among patients with established CHD, who presently receive generally similar treatment, at least in terms of medications and life-style advice.

We used an NRI method based on logistic regression models in all three papers. For papers I and II we did also have information on the time to events, but had to (somewhat arbitrarily) use a cut-off of approximately three years of follow-up, because this was the approximate minimum follow-up for most patients. Therefore, patients who experienced an event after the cut-off period were actually classified as not experiencing an event, which, in addition to not taking into account the time-to-events per se, might have influenced the estimates. Notably, there has recently been launched a method for using Cox regression, instead of logistic regression, models when calculating the NRI, using the R package *SurvIDINRI*. However, this method also makes use of a selected cut-off in follow-up time, similar to our original approach, and consequently it did not alter the results to any particular extent when applied to the current data (data not shown).

There has also recently been raised more profound criticism against the NRI. The NRI per se does not provide us of qualitative measures of the reclassification, i.e. whether unnecessary treatment (or testing) in some patients outweighs the incorrect reclassification of patients who later develop disease. This problem could be addressed by using analyses that take such weighting into account,¹⁹¹ which obviously necessitates the application of a category-based NRI. Moreover, the NRI may have several other limitations, as outlined by Leening et al,¹⁹² prompting carefulness when applying, reporting and interpreting results based on this methodology.

Moreover, rather than being competitive, the *C*-statistic, the NRI, and the IDI seem to complement each other, and should therefore be reported together when assessing the incremental value of novel biomarkers in risk prediction models.¹⁵⁷ We

did not calculate the IDI in paper I, but the IDI was reported in papers II and III. However, as advocated by others,¹⁹² paper I did report separate NRIs for events and non-events, which, due to space limitations, was not performed in paper II and III.

5.2.9 BIOSAMPLE PROCESSING

Routine laboratory measurements were performed on-site in both the WECAC and the NORVIT. We did not have detailed information on how these analyses were carried out, but it is not likely that established methods deviated considerably according to study sites, or introducing any serious bias according to the risk associations studied.

Among SAP patients in the WECAC, study specific blood and urine samples were usually collected few days prior to or at the day of angiography, and immediately frozen. In the NORVIT, blood samples were collected on-site, and then shipped to the core laboratory at Bevital AS in Bergen before freezing. Thus, in the NORVIT, samples may have been exposed to room temperature for several days, which could have led to overestimating concentrations of plasma free choline, due to formation from PC at room temperature. Long-term storage at -80°C , however, most likely did not affect any of the choline metabolites.¹⁹³

In the NORVIT, we were able to adjust the DMG-mortality association for time-span from the index AMI until baseline biosampling, but contrary to the WECAC we did not have information on fasting status. However, it is likely that patients presenting with an acute AMI constitute of a mix of fasting and non-fasting patients at the time of biosampling, similar to the SAP patients in the WECAC, and this might be reflected by equal plasma DMG concentrations in both cohorts.

5.2.10 STATISTICAL ANALYSES

We have applied both traditional and more advanced statistical methods in the current thesis, and some have already been more thoroughly discussed in previous sections.

Cox proportional and logistic regression modelling are robust and rely only on a few assumptions, in addition to having being the preferred statistical method in clinical endpoint studies for decades. However, it must be emphasized that relative risk estimates must be considered in light of absolute risk to more fully support their

clinical interpretability. In hindsight, this latter issue should perhaps have received more attention in all three papers.

By using quantile (median) linear regression for baseline analyses, we obtained effect estimates that are easy to communicate with the readers, because we did not have to transform the variables. Non-parametric statistical tests tend to reduce the information conferred by the variables, since their distributions (hence also variances) are not accounted for when using parametric tests. The semiparametric quantile linear regression, on the other hand, not only provides us with a regression coefficient, but also allows us to examine the conditional distribution of a continuous variable according to all percentiles from 0-100. This makes it possible to explore potential differences across the distribution of the independent variable. We did, however, only focus on trend analyses according to the median (50-percentile) in the current work.

Relationships in biological systems are seldom linear, which motivates the use of non-linear statistical approaches. We used GAM when exploring the associations between potential biomarkers and endpoint data, and presented the results as smoothed splines graphics. Whereas non-linear associations were not a major issue in paper I and II, several choline metabolites trended to deviate from linear relationships with incident T2D in paper III. This way of exploring and presenting data may provide more information in terms of potential underlying mechanisms, as well as avoiding the pitfall of concluding with no overall linear association in the case of a U-shaped relationship. We did not use non-linear models when assessing baseline or repeated measures data, which may pose limitations on their interpretability.

5.3 ETHICAL CONSIDERATIONS

5.3.1 LEGAL ASPECTS

All patients included in the current thesis took part on a voluntary, non-commercial basis, and provided signed informed consent. The patient recruitment, diagnostic procedures, sample handling and import of endpoint data were performed according to the Declaration of Helsinki, approval from the Regional Medical Ethics Committee, and Norwegian law on health research.¹⁹⁴ Moreover, all patients in the WECAC

underwent baseline angiography due to clinical indications, and not primarily for study purposes, thereby not exposing the patients to unnecessary hazard.

5.3.2 FINANCIAL ISSUES

The analyses of choline metabolites are presently not implemented in most routine laboratories. Therefore, the hypothetical implementation of choline metabolism biomarkers would likely add considerable cost if applied to large patient populations. Although such issues must be addressed in properly conducted cost-benefit-analyses, a potentially increment in expenditures is another ethical aspect to consider.

5.4 DISCUSSION OF THE MAIN FINDINGS

5.4.1 PUTTING THE FINDINGS INTO AN EPIDEMIOLOGICAL CONTEXT

Papers I and II. The association between plasma DMG and an overall adverse CVD risk profile found in both papers I and II have not previously been reported. However, one study proposed an indirect relationship between plasma DMG and the extent of CAD at angiography according to the BHMT 742 G>A SNP,⁹² although a later meta-analysis did not confirm any such relationship,¹⁹⁵ thereby agreeing with our baseline findings in paper I.

The prospective associations between DMG and adverse CVD prognosis are in line with results presented by Lever et al among approximately 500 New Zealand patients with a recent ACS.¹¹³ Although being secondary findings and not reaching statistical significance in models adjusted only for a limited number of potential confounders, the risk estimates in that study were similar to those reported by us in papers I and II. Our validation of the associations between plasma DMG and an adverse CVD prognosis in cohorts that were larger and had a more detailed baseline characterization and longer follow-up time, should be considered a strength. Further, by reporting the potential benefit of adding plasma DMG in risk prediction, and exploring temporal trends according to B-vitamin supplementation, the papers I and II extended current knowledge within the field.

In secondary analyses, we also explored the relationships between other choline metabolites and risk of incident AMI in paper I. Plasma choline was weakly related to

the outcome, but was attenuated once DMG was introduced into the model, suggesting that DMG might have confounded previous reports on positive risk relationships between systemic choline concentrations and CVD events.¹¹¹ We did not find any relationship between plasma betaine and incident AMI in paper I, nor was any such overall association evident among WENBIT patients with presumed SAP,¹⁹⁶ thus opposing the findings by Lever et al;¹¹³ however differences in study outcomes may also relate to inequalities between the studied populations.

Paper III. As stated above, previous studies suggested lower systemic betaine¹¹⁸⁻¹²⁰ and higher urine betaine⁵⁷ among patients with established T2D as compared to their non-diabetic counterparts. Our prospective findings affirm these observations by also showing that low plasma betaine status, in addition to higher urine DMG and sarcosine predicted incident T2D. The trends were consistent in sensitivity analyses and when adjusting for traditional T2D risk factors, including age, BMI and HbA1c. These results indicate that alterations in choline- and one-carbon metabolism may be present before the onset of clinical disease, and might even predate features traditionally associated with increased T2D risk.

A recent large Mendelian randomization study from Sweden found evidence of higher diabetes risk among patients with lower circulatory DMG concentrations as determined by polymorphisms in the gene coding for SDH.⁹⁴ Although the association was weak, the authors concluded that DMG shortage might have a causal role in diabetes development, but did not report on the concomitant levels of other choline metabolites. Notably, the lack of systemic sarcosine data was actually highlighted as a limitation of the study.⁹⁴ Indeed, if increased DMG catabolism was responsible for higher relative diabetes risk, one might suspect sarcosine levels to be positively associated with risk, as well. Also, increased downstream DMG metabolism should lead to a higher flux through the BHMT pathway, because of less product inhibition, hence possibly affecting betaine status. Our work reports on the concomitant risk relationships for all choline metabolites. We observed an overall null-effect between plasma DMG and TD2 risk in the multivariate model, and actually a trend towards a higher risk among those with particularly low serum sarcosine, thereby not

immediately supporting the conclusion made by the Swedish authors. It could be, however, that a favourable prognosis related to decreased DMG catabolism among subjects in the Swedish study rather reflected a betaine sparing effect; however a causal relationship between plasma betaine and diabetes was not supported in another Mendelian randomization investigating a specific BHMT polymorphism.¹⁹⁷ Likewise, we did not find a significant difference in the incidence rate of T2D according to the BHMT 742 G>A in the WECAC (Table 6). We have not, however, assessed this relationship in more appropriate statistical models, nor have we investigated it in the NORVIT.

The propensity to having high urine betaine excretion among patients with diabetes has been described in several smaller studies,⁵⁷ and a recent report also showed that urine betaine and DMG was highly correlated among young healthy males,¹⁷⁰ hence agreeing with our results in paper III.

5.4.2 POSSIBLE MECHANISMS

A general disclaimer. It must be emphasized that the present thesis primarily represents an epidemiological approach to the relationships between choline metabolism and non-communicable diseases. We have measured metabolites in body fluids, serving as proxies for very complex metabolic processes within subcellular compartments in various tissues. We are therefore not able to draw firm conclusions regarding mechanisms underlying metabolite status or risk relationships. However, previous, as well as the present studies, do suggest interesting, pathophysiological mechanisms in play. Also, these hypotheses may further serve as inspiration for new research projects, both within our own group, but hopefully also for others.

Mitochondrial function, energy metabolism and lipid homeostasis. The transport of choline from the cytosol into the mitochondrion is assumed to be carrier-mediated, and the rate-limiting step in the conversion of choline into betaine.⁷⁹ The inverse relationship between plasma choline and betaine with several CVD and T2D risk factors, as observed in our as well as in another¹¹² study, could therefore relate to impaired choline transport across the mitochondrial membrane. Moreover, *Chdh*

knockout mice had accumulated choline and severely decreased mitochondrial betaine levels in certain tissues, which also exhibited disturbed mitochondrial function and morphology.¹⁹⁸ Also, BADH is dependent on oxidized nicotinamide adenine dinucleotide (NAD⁺) as a cofactor,¹⁹⁹ and preliminary studies from our group have shown that alterations in the downstream metabolism of tryptophan, essential in the production of NAD⁺, may relate to incident T2D. Taken together, this suggests that the choline-to-betaine conversion may be impaired among patients more susceptible to CVD and T2D. Plasma choline levels did not relate to prevalent or incident T2D in paper III; however any build-up of choline from decreased choline oxidation might be masked by increased use of choline for other purposes, including augmented hepatic lipid export when energy intake is high, as well as increased choline uptake by the kidneys.²⁰⁰ This issue could potentially be further explored by molecular tracer studies.

In agreement with the epidemiological data, preclinical studies have also suggested inverse relationships between betaine levels and several indices of insulin sensitivity. For example, rat models of insulin resistance and T2D showed lower hepatic betaine levels, likely due to increased BHMT flux.²⁰¹ This might also reflect the positive relationship between the HOMA2-IR and plasma DMG in paper II, although no such association was seen in other, and larger, populations.⁹⁴ Moreover, betaine supplementation in insulin resistant rats improved insulin sensitivity in both the liver and in adipose tissue, possibly due to increased adipokine production and lowered endoplasmic reticulum stress.²⁰²

Most data support an increased BHMT flux in obesity and insulin resistance; however whether this represents a compensatory mechanism, for example due to altered demands for methyl groups, or whether increased BHMT flux is detrimental, remains an open question. Nevertheless, it is interesting that no association between plasma DMG and T2D was observed in paper III, and that betaine status itself does not appear to be causally related to T2D risk, as discussed earlier. This might suggest that for betaine to exert favourable effects, there is a need of both sufficient betaine *and* an adequate BHMT function. On the other hand, BHMT knockout mice actually exhibited increased energy expenditure, reduced body fat and improved glucose metabolism and insulin sensitivity in adipose tissue,²⁰³ similar to choline deprived

mice, further complicating the interpretability in terms of BHMT flux. Professor Steven Zeisel addressed this discrepancy in a recent review.⁵³ He proposed that different effects might arise from the influence by choline and betaine on PC composition, which again could affect insulin sensitivity via altered regulation of sterol regulatory element-binding protein 1.⁵³ However, to make matters even more complex, high-dose betaine supplementation in healthy persons did not alter indices of the metabolic syndrome⁸⁹ or resting energy consumption,²⁰⁴ although such therapy increased plasma betaine levels,^{89, 204} and thereby likely enhanced BHMT flux, as well.

The link between BHMT and T2D may also relate to the association between statin therapy and increased risk of incident T2D.²⁰⁵ The BHMT function might actually be attenuated by statins, which were widely used in the WECAC population, because statin therapy has been related to higher plasma betaine in other,¹⁷⁷ as well as in the WECAC cohort (data presented at the EuroPrevent Congress, Amsterdam, The Netherlands, 2014). Notably, simvastatin has been associated with impaired mitochondrial oxidative phosphorylation in skeletal muscle,²⁰⁶ thereby suggesting a potential relationship with deteriorations in the choline oxidation pathway.

In line with our results, a metabolomics study among almost 900 young and healthy Italian men and women showed an inverse relationship between circulating DMG and LDL-cholesterol, total cholesterol, and triglycerides.²⁰⁷ We found particularly strong relationships between plasma DMG and CVD risk among those with low levels of (fasting) serum triglycerides and apoB 100. It is thus alluring to speculate whether the increased CVD risk for higher plasma DMG may be related to impaired downstream DMG catabolism. This could potentially decrease BHMT activity by product inhibition, and contribute to decreased VLDL output, as well as impaired oxidation of fatty acids.²⁰³ As treatment with folic acid likely attenuates BHMT flux, it is therefore interesting that we did observe a trend towards a stronger risk relationship between plasma DMG and incident AMI among those who received folic acid intervention in the WENBIT (Appendix 1; Supplemental Tables VI and VIII). A theory of BHMT inhibition is further supported by rat studies showing that peroxisome proliferation-activator alpha (PPAR α) agonists, well-known for their

systemic triglyceride reducing abilities, likely impair DMG catabolism via down-regulated expression of DDH and SDH.²⁰⁸

The LDL hypothesis regarding ACVD holds firm ground, not least due to the recent results from the IMPROVED Reduction of Outcomes: Vytarin Efficacy International Trial (IMPROVE-IT)²⁰⁹ and results from clinical trials investigating treatment with proprotein convertase subtilisin/kexin type 9 antibodies.^{210, 211} However, the provoking proposal of increased CVD risk by decreased hepatic VLDL cholesterol output²¹² lends support from the fact that non-alcoholic fatty liver disease (NAFLD) is indeed considered an independent CVD risk factor, and is also strongly related to T2D.²¹³ In addition, studies in apoE knockout mice showed that betaine supplementation led to higher systemic levels of atherogenic lipids, but reduced body weight and atherosclerotic lesions,²¹⁴ as well as relieving NAFLD. In line with these data, analyses in the WECAC show that glycine, the immediate sarcosine catabolite, is inversely related to incident AMI, and that the association is stronger when serum apoB levels are low.²¹⁵ Notably, diminished GNMT flux has been shown to attenuate hepatic lipid export and induce hyperlipidemia,²¹⁶ in addition to impairing reverse cholesterol transport and increasing atherosclerosis.²¹⁷ Moreover, low systemic glycine levels have been related to higher risk of T2D.²¹⁸ Glycine may be produced from other metabolic pathways than the SDH and GNMT reactions, as well;⁸⁶ however our results may indirectly point to a high CVD and T2D risk when the lipid flux between the liver and peripheral tissues is reduced due to curtailed downstream choline oxidation. Hard clinical endpoint data to back up a theory of detrimental effects due to reduced hepatic VLDL export might be obtained from RCTs concerning treatment with mipomersen. This is an antisense oligonucleotide, which impairs hepatic VLDL efflux and lowers systemic LDL levels, but increases the risk of hepatic steatosis.²¹⁹

Further, it is interesting that choline- and one-carbon metabolism may also be related to apo A1 and HDL-cholesterol metabolism. Apo A1 is the major component of the HDL particle, low levels of which have been related to increased risk of both CHD and T2D, albeit Mendelian randomization studies have questioned causal relationships.^{220, 221} In line with previous epidemiological data,¹¹² plasma betaine was negatively associated with both apoA1 and HDL-cholesterol in the WECAC. Also,

studies in mice have suggested that choline deficiency lowers systemic apo A1 synthesis, probably due to reduced *PPARα* transcription,²²² and that supplementation with betaine may improve apo A1 status.²²³ These effects could also indirectly relate to BHMT, since *PPARα* decreases flux through the CBS pathway.²²⁴ In addition, a betaine deficient state may lead to reduced BHMT flux and accumulating Hcy, the latter which may lower apo A1 as well as HDL-cholesterol by several proposed mechanisms, as discussed by Obeid et al.²²⁵

Ramifications to methyl metabolism, nucleotide production, cell turnover and epigenetic regulation. The interconversion of free folates between the mitochondrial and cytosolic compartments is limited, and one-carbon units therefore move from the mitochondrion to the cytosol mainly as glycine, serine or formate.⁷⁹ Formate is used for purine synthesis, as it condenses with THF to form 10-formyl-THF.⁶⁴ Furthermore, THF can be methylated by serine via the vitamin B6 dependent enzyme serine-methyltransferase to generate 5, 10-methylene-THF, used for the production of thymidylate, and glycine.²²⁶ Consequently, altered metabolism down the choline oxidation pathway may influence the production of nucleotides necessary for DNA and RNA synthesis.²²⁷ In addition, reduced choline availability might affect genomic stability.²²⁷ These issues may have adverse effects on the cells' regenerative and reparative abilities, and could be linked to cell apoptosis associated with vulnerable atherosclerotic plaques,²²⁸ as well as apoptosis among cardiomyocytes being observed in CVD.²²⁹ The latter could, at least partly, be reflected by the strong positive relationship between plasma DMG and serum hs-cTnT among patients with suspected SAP (paper II), although adjusting for hs-cTnT did not alter the risk associations between DMG and mortality in the WECAC. The mechanisms involved in apoptosis are very complex; however, a propensity towards a catabolic state could potentially be relevant for other organs than the cardiovascular system, as well. Notably, patients with T2D are characterized by reduced mass of pancreatic beta cells, which may be ascribed to a mismatch in the dynamic process of cell replicating and apoptotic cell death.²³⁰

Choline deficiency²³¹ and impaired BHMT flux^{67, 232} are associated with lower hepatic SAM and increased SAH. The SAM/SAH ratio may reflect the methylation potential,²³³ and although partially compensated for by up-regulation of methyltransferases,⁶⁴ a lower methylation potential due to impairments along the choline oxidation pathway could influence epigenetic regulation in terms of reduced methylation of cytosine-guanine repeat (CpG) islands in the DNA.²³⁴ However, the extreme complexity of interactions between genes makes it difficult to predict how an overall reduction in methylation capacity may influence phenotype or prognosis. Reflecting this complexity, DNA hypo- and hypermethylation may coexist at specific regions within the genome, yet several studies have associated global DNA hypomethylation with atherosclerosis.^{235, 236} Moreover, DNA methylation is inversely associated with age,²³⁷ the latter being far the most important predictor of adverse CVD prognosis in both the WECAC and the NORVIT cohorts (data not shown).

Epigenetic regulation is also important in glucose and energy metabolism,²³⁵ as well as in T2D. The role of methylation was further highlighted in a study among nicotinamide *N*-methyltransferase knocked-out mice, which had a propensity to leanness because of increased energy expenditure. This was explained by increased SAM available for methylation reactions including polyamine synthesis in peripheral adipose tissue, as well as higher flux of oxidized nicotinamide adenine dinucleotide (NAD⁺).²³⁸ Betaine supplementation was also associated with increased hepatic SAM levels in fat-fed, insulin resistant mice,²³⁹ and a recent study indicated that improved methylation status from betaine might favorably influence several enzymes involved in lipogenesis and fatty acid synthesis, including the fat mass and obesity-associated (FTO) gene.²⁴⁰

Choline metabolites in the urine. It is plausible that systemic choline metabolite status primarily reflects altered metabolism in the liver, whereas urinary concentrations of choline metabolites mirror their handling and/or metabolism in the kidney. This is supported by the relative modest correlations between systemic and urinary levels of choline metabolites in the WECAC (paper III), and the low correlation reported between plasma and urinary betaine in previous studies.⁵⁷

However, we observed a negative correlation between baseline plasma and urine betaine among patients with established T2D (paper III), indicating that patients with overt disease actually may have urinary betaine loss contributing to low systemic betaine levels. This agrees with the findings by Lever et al, who reported that some patients with T2D might actively excrete betaine from the kidneys, thereby exceeding 100% fractional clearance for betaine.²⁴¹

Urine betaine excretion has been positively associated with parameters of impaired glucose control in patients with diabetes;^{125, 241, 242} however the increase may not be dependent on high systemic glucose levels per se,²⁴³ but a link to hyperosmolarity has been suggested.⁵⁷ Proximal tubular dysfunction has been proposed as a likely contributor to increased urinary betaine excretion among patients with diabetes, because betaine is freely filtered in the glomeruli and urine betaine is correlated with urinary concentrations of retinol binding protein, a marker of tubular dysfunction.²⁴² This corresponds to our findings in paper III, since the predictive ability of urine betaine, as well as DMG and sarcosine, was independent of microalbuminuria which primarily reflects glomerular leakage.²⁴⁴ Our findings extend previous data, as we also observed that patients who later developed T2D had a 18% and 23% increase in baseline urine betaine and sarcosine concentrations, respectively, as compared to those who did not develop T2D (Article III). A 9.9% increase in urine DMG was also observed; however this difference was not statistically significant. These differences did not seem to be explained by reverse causation due to misclassification of T2D at baseline, as urine choline metabolites levels were similar among patients with possible and patients without T2D. This suggests that altered renal handling and/or metabolism of choline metabolites is present before the onset of overt T2D.

As compared to betaine, relatively little attention has previously been paid to urinary DMG and sarcosine excretion. Betaine ingestion is associated with a transient increase in urine DMG concentration, and a slightly higher fractional excretion has been reported for DMG than for betaine in pigs.⁵⁷ Treatment with fibrates, which are PPAR α agonists, has also been related to a massive increase in both urine betaine and DMG, and especially so among patients with T2D.^{241, 245} Moreover, urine betaine and

DMG correlated strongly with urine choline in another study, which also reported that fenofibrate led to larger increments in urine betaine, than DMG and choline.²⁴⁵ In paper III, urine choline was not associated with incident T2D; however it is plausible that betaine and DMG, and perhaps also other choline metabolites, may have some common renal handling, also in terms of active excretion from renal epithelial cells.^{241,}
245

Almost no sarcosine is excreted in the urine among normal subjects.²⁴⁶ Higher urinary sarcosine levels have been observed among patients with impaired SDH,²⁴⁷ probably reflecting the role of sarcosine in scavenging excess methyl.⁸⁵ Most sarcosine filtered in the glomeruli is assumed to be absorbed in the tubular system, and sarcosine may compete with glycine for reuptake in tubuli.²⁴⁷ One might therefore speculate whether increased sarcosine excretion partly results from enhanced renal glycine uptake in a glycine deficient state being associated with T2D. Our data does not support this, however, as urine glycine concentrations did not relate to T2D in the WECAC (data not shown).

Both GNMT²⁴⁸ and BHMT⁴⁸ are abundant in the kidneys, and increased flux over these enzymes, at least locally, could perhaps lead to a spill-over of products from renal cells and into the tubulus system. On the other hand, insulin resistance and T2D may affect GNMT differently in the kidney as compared to the liver. For example, renal GNMT flux might not be up-regulated in prediabetic state, as is the case in the diabetic state. This somewhat contrasts hepatic GNMT flux, which is most likely up-regulated in both conditions.²⁴⁹ We also observed that treating the patients with B-vitamins altered systemic choline metabolites during follow-up, most probably related to reduced BHMT flux; however less consistent findings were made in the urine. Potential tissue specific alterations in BHMT and GNMT flux brought about by insulin resistance and T2D therefore makes it problematic to draw firm conclusions when only metabolite concentrations in the blood and urine are measured.

Can dietary factors explain our findings? In paper III, we also had data available on estimated daily intake of both choline and betaine in a subpopulation of the WECAC participants, suggesting the daily intake of choline to be far below that recommended

by USDA.⁵² We did not, however, find any differences in these measures according to T2D, although plasma TMAO, a proposed marker of choline intake,¹⁰² was about two-fold among those with existing T2D at baseline, the latter agreeing with previous observations.^{120, 123, 250} Moreover, recent studies actually found lower plasma TMAO and betaine levels to predict gestational diabetes in women,^{251, 252} thereby complicating the relationship between TMAO and diabetes risk further. Higher intakes of PC were recently related to increased T2D risk,¹²² however, we did not explore dietary determinants of choline metabolites, nor did we assess the relationships between total choline and betaine intake with the risk of new-onset T2D per se. Nevertheless, including data on choline and betaine intake in the multivariate models in paper III did not affect the risk estimates, nor was plasma TMAO predictive of incident T2D in general.

A recent study actually questioned the alleged causal association between plasma TMAO and ACVD, redirecting the focus to hepatic FMO3 instead.²⁵³ Accordingly, preliminary results from the WECAC also challenge the relationship between plasma TMAO and CVD risk. This agrees with previous epidemiological data reporting no statistically significant associations between intakes of choline and betaine with incident CVD^{116, 117, 254}

Relatively weak associations between dietary factors and plasma choline and betaine levels have previously been reported.⁵⁵ Although we did not estimate methionine intake, it therefore seems reasonable to conclude that dietary intake of labile methyl groups did not severely impact the overall risk relationships observed in our studies.

6 CONCLUSIONS

The aims of our patient population studies were to investigate prospective risk relationships between plasma DMG with incident AMI and mortality, as well as systemic and urinary choline metabolites in relation to incident T2D. Additionally, we explored temporal changes in systemic and urinary choline metabolites, also according to treatment with B-vitamins crucial for enzymes in the choline oxidation pathway. In conclusion, the current thesis answered the study aims as follows:

6.1 STUDY I

Plasma DMG was positively associated with the risk of incident AMI during about four and a half years of follow-up. The risk associations were independent of traditional CHD risk factors and remained also when adjusting for potential confounders. Adding plasma DMG to a model of traditional CHD risk factors improved model discrimination and suggested enhanced classification of patients at risk. Treatment with folic acid and vitamin B12 lowered plasma DMG levels during repeated measures. Among patients allocated to placebo treatment, plasma DMG showed excellent test-retest reliability.

6.2 STUDY II

Plasma DMG was positively related to long-term risk of all-cause and CVD mortality in two independent cohorts of patients with suspected SAP and AMI, respectively. The risk associations did not seem to be explained by traditional CVD risk factors, nor by potential confounders, and there were indications of improved risk prediction for plasma DMG beyond traditional CVD risk factors. The attenuation of temporal plasma DMG increments by combined folic acid and B12 treatment was consistent in both cohorts, as was the within-person reproducibility of plasma DMG.

6.3 STUDY III

Low plasma betaine and high urine betaine, DMG and sarcosine were related to increased risk of incident T2D throughout 7 years of follow-up. The risk relationships were not dependent on several T2D risk factors or potential confounders, and adding

either plasma betaine or urine sarcosine suggested improved model discrimination and reclassification of patients at risk. Both metabolites also showed good within-person reproducibility. As compared to placebo, treatment with folic acid with or without vitamin B6 led to relative increases in plasma choline and betaine, but decreases in plasma DMG and serum sarcosine. Similar, although less pronounced alterations were seen for urine choline metabolites.

Our results partly provide important validation on previous results in smaller epidemiological studies, but also add new knowledge into the field of choline metabolism. As described above, the present thesis therefore generates several novel hypotheses relating to diseases like CVD and diabetes, which determinate morbidity and mortality worldwide, thereby warranting further studies within this research field.

7 FUTURE PERSPECTIVES

Our research group will continue to explore the choline oxidation pathway in relation to CVD and T2D, with a particular focus on lipid and energy homeostasis.

We are currently in the process of linking both the WECAC and the NORVIT cohorts (collectively named the Norwegian Study on Coronary Heart Disease; NORCAD) with data from extended follow-up on incident CVD events and mortality throughout 2009, thereby providing us with even more reliable risk estimates; however we must also keep in mind that any potential effect modification by WENBIT and NORVIT intervention might be diluted when analyzing these extended follow-up data. Moreover, the research group is also applying for linkage between the NORCAD and the Norwegian prescription database, which will hopefully allow for more accurate identification of incident cases of diabetes, as well as the ability for exploring risk associations according to pharmacological treatment and patients' metabolic profiles.

In the dawn of the "big data" era it seems imperative that researchers across groups and borders take advantage of each other's data material, in order to optimize publicly funded projects. We therefore also seek to extend our collaboration with other research environments, not least to validate epidemiological findings, but also because experimental research among both animals and humans will provide valuable information on novel pathophysiological pathways. For example, a human dietary intervention trial is about to be launched in our department, partly based upon findings from studies of choline- and one-carbon metabolites in relation to dietary intake patterns in the WENBIT, as well as experiments on mice carried out by the collaborating Lipid Research Group at the University of Bergen.

8 REFERENCES

1. GBD 2013 Mortality and Causes of Death Collaborators. Global, regional, and national age-sex specific all-cause and cause-specific mortality for 240 causes of death, 1990-2013: a systematic analysis for the Global Burden of Disease Study 2013. *Lancet*. 2015;385:117-71..
2. Ford ES and Capewell S. Proportion of the decline in cardiovascular mortality disease due to prevention versus treatment: public health versus clinical care. *Annu Rev Public Health*. 2011;32:5-22..
3. Murray CJ, Vos T, Lozano R et al. Disability-adjusted life years (DALYs) for 291 diseases and injuries in 21 regions, 1990-2010: a systematic analysis for the Global Burden of Disease Study 2010. *Lancet*. 2012;380:2197-223.
4. Perk J, De Backer G, Gohlke H, et al. European guidelines on cardiovascular disease prevention in clinical practice (version 2012). The fifth joint task force of the European Society of Cardiology and other societies on cardiovascular disease prevention in clinical practice (constituted by representatives of nine societies and by invited experts). *Eur Heart J*. 2012;33:1635-701.
5. Statistics Norway. Causes of death 2012. Accessed Apr 24 2015. URL: <http://www.ssb.no>
6. The Norwegian Institute of Public Health. Norwegian Cardiovascular Disease Registry. Accessed Sep 29 2014. URL: <http://www.fhi.no/helseregistre/hjerte-og-karregisteret>
7. Tomas J. SWEDEHEART Annual report 2013. Accessed Apr 15 2015. URL: http://www.ucr.uu.se/swedeheart/index.php/component/docman/doc_download/345-swedeheart-arsrapport-2013-english-engelsk
8. Statens Institut for Folkesundhed. Dansk hjerteregister. Årsberetning 2013. URL: http://www.si-folkesundhed.dk/upload/%C3%A5rsberetning_2013.pdf
9. Øyen N, Nygård O, Iglund J et al. Sykehusinnleggelser for hjerte- og karsykdom i Helse Vest i perioden 1992 – 2001. *Tidsskr Nor Legerforen*. 2007;128:17-23.

10. Graff-Iversen S, Jenum AK, Grotvedt L et al. [Risk factors for myocardial infarction, stroke and diabetes in Norway]. *Tidsskr Nor Laegeforen*. 2007;127:2537-41.
11. Nichols M, Townsend N, Luengo-Fernandez R, et al. European Cardiovascular Disease Statistics 2012. European Heart Network, Brussels, European Society of Cardiology, Sophia Antipolis. Access date 21 Jan 2016. URL: http://www.escardio.org/static_file/Escardio/Press-media/press-releases/2013/EU-cardiovascular-disease-statistics-2012.pdf
12. Abrams J. Clinical practice. Chronic stable angina. *N Engl J Med*. 2005;352:2524-33.
13. Thygesen K, Alpert JS, Jaffe AS, et al. Third universal definition of myocardial infarction. *Eur Heart J*. 2012;33:2551-67.
14. Maddox TM, Stanislawski MA, Grunwald GK, et al. Nonobstructive coronary artery disease and risk of myocardial infarction. *JAMA*. 2014;312:1754-63.
15. D'Agostino RB, Sr., Vasan RS, Pencina MJ, et al. General cardiovascular risk profile for use in primary care: the Framingham Heart Study. *Circulation*. 2008;117:743-53.
16. Cholesterol Treatment Trialists Collaboration, Baigent C, Blackwell L, Emberson J, et al. Efficacy and safety of more intensive lowering of LDL cholesterol: a meta-analysis of data from 170,000 participants in 26 randomised trials. *Lancet*. 2010;376:1670-81.
17. Visser ME, Jakulj L, Kastelein JJ and Stroes ES. LDL-C-lowering therapy: current and future therapeutic targets. *Curr Cardiol Rep*. 2008;10:512-20.
18. Koskinas KC, Windecker S and Raber L. Regression of coronary atherosclerosis: Current evidence and future perspectives. *Trends Cardiovasc Med*. 2015 19. pii: S1050-1738(15)00141-3.
19. Pedersen TR. Pleiotropic effects of statins: evidence against benefits beyond LDL-cholesterol lowering. *Am J Cardiovasc Drugs*. 2010;10 Suppl 1:10-7.
20. Mihos CG, Pineda AM and Santana O. Cardiovascular effects of statins, beyond lipid-lowering properties. *Pharmacol Res*. 2014;88:12-9.

21. Mora S, Wenger NK, Demicco DA, et al. Determinants of residual risk in secondary prevention patients treated with high- versus low-dose statin therapy: the Treating to New Targets (TNT) study. *Circulation*. 2012;125:1979-87.
22. American Diabetes Association. Standards of medical care in diabetes--2014. *Diabetes Care*. 2014;37 Suppl 1:S14-80.
23. International Diabetes Federation. IDF Diabetes Atlas, 6th edn. 2013. Access date 21 Jan 2016.
URL: https://www.idf.org/sites/default/files/EN_6E_Atlas_Full_0.pdf
24. Cooper JG, Claudi T, Thordarson HB, et al. Treatment of type 1 diabetes in the specialist health service--data from the Norwegian Diabetes Register for Adults. *Tidsskr Nor Laegeforen*. 2013;133:2257-62.
25. Guðbjörnsdóttir S; Eliasson B; Cederholm et al. Swedish National Diabetes Register Annual report 2013. Access date 21 Jan 2016.
URL: https://www.ndr.nu/pdfs/Annual_Report_NDR_2013.pdf
26. Statens Serum Institut. Tal på diabetes 1996-2012. Access date Jul 09 2015.
URL: <http://www.ssi.dk/Sundhedsdataogit/Sundhedsvaesenet%20i%20tal/Specifikke%20omraader/Diabetes.aspx>
27. The Norwegian Prescription Database. Diabetesmidler 2014, alle aldre, begge kjønn, hele landet. Access date Apr 24 2015.
URL: <http://www.reseptregisteret.no>
28. Statistics Norway. Allmennlegetjenesten 2014. Access date Apr 24 2015.
URL: <http://www.ssb.no>
29. Statistics Norway. Pasienter på somatiske sykehus 2014. Access date Apr 13 2015. URL: <http://www.ssb.no>
30. Reaven G. Insulin resistance and coronary heart disease in nondiabetic individuals. *Arterioscler Thromb Vasc Biol*. 2012;32:1754-9.
31. DeFronzo RA. Banting Lecture. From the triumvirate to the ominous octet: a new paradigm for the treatment of type 2 diabetes mellitus. *Diabetes*. 2009;58:773-95.

32. Kahn SE, Cooper ME and Del Prato S. Pathophysiology and treatment of type 2 diabetes: perspectives on the past, present, and future. *Lancet*. 2014;383:1068-83.
33. Campbell IW. Epidemiology and clinical presentation of type 2 diabetes. *Value Health*. 2000;3 Suppl 1:3-6.
34. Tancredi M, Rosengren A, Svensson AM, et al. Excess mortality among persons with type 2 diabetes. *N Engl J Med*. 2015;373:1720-32.
35. Huxley R, Barzi F and Woodward M. Excess risk of fatal coronary heart disease associated with diabetes in men and women: meta-analysis of 37 prospective cohort studies. *BMJ*. 2006;332:73-8.
36. Emerging Risk Factors Collaboration, Di Angelantonio E, Kaptoge S, et al. Association of cardiometabolic multimorbidity with mortality. *JAMA*. 2015;314:52-60.
37. Norhammar A, Tenerz A, Nilsson G, et al. Glucose metabolism in patients with acute myocardial infarction and no previous diagnosis of diabetes mellitus: a prospective study. *Lancet*. 2002;359:2140-4.
38. Stern MP. Do non-insulin-dependent diabetes mellitus and cardiovascular disease share common antecedents? *Ann Intern Med*. 1996;124:110-6.
39. Magnusson M, Lewis GD, Ericson U, et al. A diabetes-predictive amino acid score and future cardiovascular disease. *Eur Heart J*. 2013;34:1982-9.
40. Wang TJ, Larson MG, Vasan RS, et al. Metabolite profiles and the risk of developing diabetes. *Nat Med*. 2011;17:448-53.
41. Ong KL, Barter PJ and Waters DD. Cardiovascular drugs that increase the risk of new-onset diabetes. *Am Heart J*. 2014;167:421-8.
42. Phung OJ, Schwartzman E, Allen RW, et al. Sulphonylureas and risk of cardiovascular disease: systematic review and meta-analysis. *Diabet Med*. 2013;30:1160-71.
43. Mannucci E, Monami M, Di Bari M, et al. Cardiac safety profile of rosiglitazone: a comprehensive meta-analysis of randomized clinical trials. *Int J Cardiol*. 2010;143:135-40.

44. Stern MP. Diabetes and cardiovascular disease. The "common soil" hypothesis. *Diabetes*. 1995;44:369-74.
45. Capewell S and Buchan I. Why have sustained increases in obesity and type 2 diabetes not offset declines in cardiovascular mortality over recent decades in Western countries? *Nutr Metab Cardiovasc Dis*. 2012;22:307-11.
46. Porter RK, Scott JM and Brand MD. Choline transport into rat liver mitochondria. Characterization and kinetics of a specific transporter. *J Biol Chem*. 1992;267:14637-46.
47. Ueland PM. Choline and betaine in health and disease. *J Inherit Metab Dis*. 2011;34:3-15.
48. Pajares MA and Perez-Sala D. Betaine homocysteine S-methyltransferase: just a regulator of homocysteine metabolism? *Cell Mol Life Sci*. 2006;63:2792-803.
49. Cernei N, Heger Z, Gumulec J, et al. Sarcosine as a potential prostate cancer biomarker--a review. *Int J Mol Sci*. 2013;14:13893-908.
50. Li Z and Vance DE. Phosphatidylcholine and choline homeostasis. *J Lipid Res*. 2008;49:1187-94.
51. DeLong CJ, Shen YJ, Thomas MJ and Cui Z. Molecular distinction of phosphatidylcholine synthesis between the CDP-choline pathway and phosphatidylethanolamine methylation pathway. *J Biol Chem*. 1999;274:29683-8.
52. Institute of Medicine (US) Standing Committee on the Scientific Evaluation of Dietary Reference Intakes and its Panel on Folate OBV, and Choline. Dietary Reference Intakes for Thiamin, Riboflavin, Niacin, Vitamin B6, Folate, Vitamin B12, Pantothenic Acid, Biotin, and Choline. Dietary Reference Intakes for Thiamin, Riboflavin, Niacin, Vitamin B6, Folate, Vitamin B12, Pantothenic Acid, Biotin, and Choline. Washington (DC); 1998.
53. Zeisel SH. Metabolic crosstalk between choline/1-carbon metabolism and energy homeostasis. *Clin Chem Lab Med*. 2013;51:467-75.
54. Nordic Counsel of Ministers 2014. Nordic nutrition recommendations 2012

- Integrating nutrition and physical activity, 5th edition. Access date Jul 09 2015.
URL: <https://www.norden.org/en/theme/tidligere-temaer/themes-2014/nordic-nutrition-recommendation/nordic-nutrition-recommendations-2012>
55. Konstantinova SV, Tell GS, Vollset SE, et al. Dietary patterns, food groups, and nutrients as predictors of plasma choline and betaine in middle-aged and elderly men and women. *Am J Clin Nutr.* 2008;88:1663-9.
 56. Samuelson G. Dietary habits and nutritional status in adolescents over Europe. An overview of current studies in the Nordic countries. *Eur J Clin Nutr.* 2000;54 Suppl 1:S21-8.
 57. Lever M and Slow S. The clinical significance of betaine, an osmolyte with a key role in methyl group metabolism. *Clin Biochem.* 2010;43:732-44.
 58. Salvi F and Gadda G. Human choline dehydrogenase: medical promises and biochemical challenges. *Arch Biochem Biophys.* 2013;537:243-52.
 59. Pietruszko R and Chern M. Betaine aldehyde dehydrogenase from rat liver mitochondrial matrix. *Chem Biol Interact.* 2001;130-132:193-9.
 60. Figueroa-Soto CG, Lopez-Cervantes G and Valenzuela-Soto EM. Immunolocalization of betaine aldehyde dehydrogenase in porcine kidney. *Biochem Biophys Res Commun.* 1999;258:732-6.
 61. Munoz-Clares RA, Diaz-Sanchez AG, Gonzalez-Segura L and Montiel C. Kinetic and structural features of betaine aldehyde dehydrogenases: mechanistic and regulatory implications. *Arch Biochem Biophys.* 2010;493:71-81.
 62. Zhang Y, Zhu T, Wang L, et al. Homocysteine homeostasis and betaine-homocysteine *S*-methyltransferase expression in the brain of hibernating bats. *PLoS One.* 2013;8:e85632.
 63. Brosnan JT and Brosnan ME. The sulfur-containing amino acids: an overview. *J Nutr.* 2006;136:1636S-1640S.
 64. Stover PJ. Polymorphisms in 1-carbon metabolism, epigenetics and folate-related pathologies. *J Nutrigenet Nutrigenomics.* 2011;4:293-305.
 65. Teng YW, Cerdena I and Zeisel SH. Homocysteinemia in mice with genetic betaine homocysteine *S*-methyltransferase deficiency is independent of dietary folate intake. *J Nutr.* 2012;142:1964-7.

66. Selicharova I, Korinek M, Demianova Z, et al. Effects of hyperhomocysteinemia and betaine-homocysteine *S*-methyltransferase inhibition on hepatocyte metabolites and the proteome. *Biochim Biophys Acta*. 2013;1834:1596-606.
67. Strakova J, Gupta S, Kruger WD, et al. Inhibition of betaine-homocysteine *S*-methyltransferase in rats causes hyperhomocysteinemia and reduces liver cystathionine beta-synthase activity and methylation capacity. *Nutr Res*. 2011;31:563-71.
68. Allen RH, Stabler SP and Lindenbaum J. Serum betaine, N,N-dimethylglycine and N-methylglycine levels in patients with cobalamin and folate deficiency and related inborn errors of metabolism. *Metabolism*. 1993;42:1448-60.
69. Finkelstein JD. Metabolic regulatory properties of *S*-adenosylmethionine and *S*-adenosylhomocysteine. *Clin Chem Lab Med*. 2007;45:1694-9.
70. Park EI and Garrow TA. Interaction between dietary methionine and methyl donor intake on rat liver betaine-homocysteine methyltransferase gene expression and organization of the human gene. *J Biol Chem*. 1999;274:7816-24.
71. Lysne V, Bjorndal B, Vik R, et al. A protein extract from chicken reduces plasma homocysteine in rats. *Nutrients*. 2015;7:4498-511.
72. Finkelstein JD and Martin JJ. Methionine metabolism in mammals. Adaptation to methionine excess. *J Biol Chem*. 1986;261:1582-7.
73. Finkelstein JD, Harris BJ, Martin JJ and Kyle WE. Regulation of hepatic betaine-homocysteine methyltransferase by dietary methionine. *Biochem Biophys Res Commun*. 1982;108:344-8.
74. Ratnam S, Wijekoon EP, Hall B, et al. Effects of diabetes and insulin on betaine-homocysteine *S*-methyltransferase expression in rat liver. *Am J Physiol Endocrinol Metab*. 2006;290:E933-9.
75. Wijekoon EP, Hall B, Ratnam S, et al. Homocysteine metabolism in ZDF (type 2) diabetic rats. *Diabetes*. 2005;54:3245-51.
76. Lawson-Yuen A and Levy HL. The use of betaine in the treatment of elevated homocysteine. *Mol Genet Metab*. 2006;88:201-7.

77. Strakova J, Williams KT, Gupta S, et al. Dietary intake of S-(alpha-carboxybutyl)-DL-homocysteine induces hyperhomocysteinemia in rats. *Nutr Res.* 2010;30:492-500.
78. Binzak BA, Vockley JG, Jenkins RB and Vockley J. Structure and analysis of the human dimethylglycine dehydrogenase gene. *Mol Genet Metab.* 2000;69:181-7.
79. Tibbetts AS and Appling DR. Compartmentalization of mammalian folate-mediated one-carbon metabolism. *Annu Rev Nutr.* 2010;30:57-81.
80. Steenkamp DJ and Husain M. The effect of tetrahydrofolate on the reduction of electron transfer flavoprotein by sarcosine and dimethylglycine dehydrogenases. *Biochem J.* 1982;203:707-15.
81. Reap EA and Lawson JW. Stimulation of the immune response by dimethylglycine, a nontoxic metabolite. *J Lab Clin Med.* 1990;115:481-6.
82. Graber CD, Goust JM, Glassman AD, et al. Immunomodulating properties of dimethylglycine in humans. *J Infect Dis.* 1981;143:101-5.
83. Lawson BR, Belkowski SM, Whitesides JF, et al. Immunomodulation of murine collagen-induced arthritis by N, N-dimethylglycine and a preparation of Perna canaliculus. *BMC Complement Altern Med.* 2007;7:20.
84. Kalmar ID, Cools A, Verstegen MW, et al. Dietary supplementation with dimethylglycine affects broiler performance and plasma metabolites depending on dose and dietary fatty acid profile. *J Anim Physiol Anim Nutr (Berl).* 2011;95:146-53.
85. Mudd SH, Brosnan JT, Brosnan ME, et al. Methyl balance and transmethylation fluxes in humans. *Am J Clin Nutr.* 2007;85:19-25.
86. Wang W, Wu Z, Dai Z, et al. Glycine metabolism in animals and humans: implications for nutrition and health. *Amino Acids.* 2013;45:463-77.
87. Wang H, Wang L, Zhang H, et al. (1)H NMR-based metabolic profiling of human rectal cancer tissue. *Mol Cancer.* 2013;12:121.
88. Yang CP, Wang HA, Tsai TH, et al. Characterization of the neuropsychological phenotype of glycine N-methyltransferase-/- mice and evaluation of its

- responses to clozapine and sarcosine treatments. *Eur Neuropsychopharmacol.* 2012;22:596-606.
89. Schwab U, Alifhan G, Aro A and Uusitupa M. Long-term effect of betaine on risk factors associated with the metabolic syndrome in healthy subjects. *Eur J Clin Nutr.* 2011;65:70-6.
90. Moazzami AA, Bondia-Pons I, Hanhineva K, et al. Metabolomics reveals the metabolic shifts following an intervention with rye bread in postmenopausal women--a randomized control trial. *Nutr J.* 2012;11:88.
91. McGregor DO, Dellow WJ, Lever M, et al. Dimethylglycine accumulates in uremia and predicts elevated plasma homocysteine concentrations. *Kidney Int.* 2001;59:2267-72.
92. Weisberg IS, Park E, Ballman KV, et al. Investigations of a common genetic variant in betaine-homocysteine methyltransferase (BHMT) in coronary artery disease. *Atherosclerosis.* 2003;167:205-14.
93. Fredriksen A, Meyer K, Ueland PM, et al. Large-scale population-based metabolic phenotyping of thirteen genetic polymorphisms related to one-carbon metabolism. *Hum Mutat.* 2007;28:856-65.
94. Magnusson M, Wang TJ, Clish C, et al. Dimethylglycine deficiency and the development of diabetes mellitus. *Diabetes.* 2015 Aug;64(8):3010-6..
95. Chern MK and Pietruszko R. Evidence for mitochondrial localization of betaine aldehyde dehydrogenase in rat liver: purification, characterization, and comparison with human cytoplasmic E3 isozyme. *Biochem Cell Biol.* 1999;77:179-87.
96. Guzman-Partida AM and Valenzuela-Soto EM. Porcine kidney betaine aldehyde dehydrogenase: purification and properties. *Comp Biochem Physiol B Biochem Mol Biol.* 1998;119:485-91.
97. Lang H, Minaian K, Freudenberg N, et al. Tissue specificity of rat mitochondrial dimethylglycine dehydrogenase expression. *Biochem J.* 1994;299 (Pt 2):393-8.
98. Bergeron F, Otto A, Blache P, et al. Molecular cloning and tissue distribution of rat sarcosine dehydrogenase. *Eur J Biochem.* 1998;257:556-61.

99. Yoon JK, Kim do H and Koo JS. Implications of differences in expression of sarcosine metabolism-related proteins according to the molecular subtype of breast cancer. *J Transl Med.* 2014;12:149.
100. Ogawa H, Gomi T, Takusagawa F and Fujioka M. Structure, function and physiological role of glycine *N*-methyltransferase. *Int J Biochem Cell Biol.* 1998;30:13-26.
101. Field MS, Shields KS, Abarinov EV, et al. Reduced MTHFD1 activity in male mice perturbs folate- and choline-dependent one-carbon metabolism as well as transsulfuration. *J Nutr.* 2013;143:41-5.
102. Wang Z, Klipfell E, Bennett BJ, et al. Gut flora metabolism of phosphatidylcholine promotes cardiovascular disease. *Nature.* 2011;472:57-63.
103. Sparks JD, Collins HL, Chirieac DV, et al. Hepatic very-low-density lipoprotein and apolipoprotein B production are increased following in vivo induction of betaine-homocysteine *S*-methyltransferase. *Biochem J.* 2006;395:363-71.
104. Olthof MR, van Vliet T, Verhoef P, et al. Effect of homocysteine-lowering nutrients on blood lipids: results from four randomised, placebo-controlled studies in healthy humans. *PLoS Med.* 2005;2:e135.
105. Byrne CD and Targher G. NAFLD: A multisystem disease. *J Hepatol.* 2015;62:S47-S64.
106. Nygard O, Vollset SE, Refsum H, et al. Total homocysteine and cardiovascular disease. *J Intern Med.* 1999;246:425-54.
107. Clarke R, Halsey J, Lewington S, et al. Effects of lowering homocysteine levels with B vitamins on cardiovascular disease, cancer, and cause-specific mortality: Meta-analysis of 8 randomized trials involving 37 485 individuals. *Arch Intern Med.* 2010;170:1622-31.
108. Clarke R, Bennett DA, Parish S, et al. Homocysteine and coronary heart disease: meta-analysis of MTHFR case-control studies, avoiding publication bias. *PLoS Med.* 2012;9:e1001177.
109. van Meurs JB, Pare G, Schwartz SM, et al. Common genetic loci influencing plasma homocysteine concentrations and their effect on risk of coronary artery disease. *Am J Clin Nutr.* 2013;98:668-76.

110. Loland KH, Bleie O, Blix AJ, et al. Effect of homocysteine-lowering B vitamin treatment on angiographic progression of coronary artery disease: a Western Norway B Vitamin Intervention Trial (WENBIT) substudy. *Am J Cardiol.* 2010;105:1577-84.
111. Danne O and Mockel M. Choline in acute coronary syndrome: an emerging biomarker with implications for the integrated assessment of plaque vulnerability. *Expert Rev Mol Diagn.* 2010;10:159-71.
112. Konstantinova SV, Tell GS, Vollset SE, et al. Divergent associations of plasma choline and betaine with components of metabolic syndrome in middle age and elderly men and women. *J Nutr.* 2008;138:914-20.
113. Lever M, George PM, Elmslie JL, et al. Betaine and secondary events in an acute coronary syndrome cohort. *PLoS One.* 2012;7:e37883.
114. Lundberg P, Dudman NP, Kuchel PW and Wilcken DE. ¹H NMR determination of urinary betaine in patients with premature vascular disease and mild homocysteinemia. *Clin Chem.* 1995;41:275-83.
115. Tang WH, Wang Z, Levison BS, et al. Intestinal microbial metabolism of phosphatidylcholine and cardiovascular risk. *N Engl J Med.* 2013;368:1575-84.
116. Dalmeijer GW, Olthof MR, Verhoef P, et al. Prospective study on dietary intakes of folate, betaine, and choline and cardiovascular disease risk in women. *Eur J Clin Nutr.* 2008;62:386-94.
117. Bidulescu A, Chambless LE, Siega-Riz AM, et al. Usual choline and betaine dietary intake and incident coronary heart disease: the Atherosclerosis Risk in Communities (ARIC) study. *BMC Cardiovasc Disord.* 2007;7:20.
118. Chen L, Chen YM, Wang LJ, et al. Higher homocysteine and lower betaine increase the risk of microangiopathy in patients with diabetes mellitus carrying the GG genotype of PEMT G774C. *Diabetes Metab Res Rev.* 2013;29:607-17.
119. Lever M, Slow S, McGregor DO, et al. Variability of plasma and urine betaine in diabetes mellitus and its relationship to methionine load test responses: an observational study. *Cardiovasc Diabetol.* 2012;11:34.

120. Lever M, George PM, Slow S, et al. Betaine and Trimethylamine-*N*-Oxide as Predictors of Cardiovascular Outcomes Show Different Patterns in Diabetes Mellitus: An Observational Study. *PLoS One*. 2014;9:e114969.
121. Koutros S, Meyer TE, Fox SD, et al. Prospective evaluation of serum sarcosine and risk of prostate cancer in the Prostate, Lung, Colorectal and Ovarian Cancer Screening Trial. *Carcinogenesis*. 2013;34:2281-5.
122. Li Y, Wang DD, Chiuve SE, et al. Dietary phosphatidylcholine intake and type 2 diabetes in men and women. *Diabetes Care*. 2015;38:e13-4.
123. Tang WH, Wang Z, Shrestha K, et al.. Intestinal microbiota-dependent phosphatidylcholine metabolites, diastolic dysfunction, and adverse clinical outcomes in chronic systolic heart failure. *J Card Fail*. 2015;21:91-6.
124. Miao J, Ling AV, Manthena PV, et al. Flavin-containing monooxygenase 3 as a potential player in diabetes-associated atherosclerosis. *Nat Commun*. 2015;6:6498.
125. Schartum-Hansen H, Ueland PM, Pedersen ER, et al Assessment of urinary betaine as a marker of diabetes mellitus in cardiovascular patients. *PLoS One*. 2013;8:e69454.
126. Ebbing M, Bleie O, Ueland PM, et al. Mortality and cardiovascular events in patients treated with homocysteine-lowering B vitamins after coronary angiography: a randomized controlled trial. *JAMA*. 2008;300:795-804.
127. Bonna KH, Njolstad I, Ueland PM, et al. Homocysteine lowering and cardiovascular events after acute myocardial infarction. *N Engl J Med*. 2006;354:1578-88.
128. Windecker S, Kolh P, Alfonso F et al. 2014 ESC/EACTS Guidelines on myocardial revascularization: The Task Force on Myocardial Revascularization of the European Society of Cardiology (ESC) and the European Association for Cardio-Thoracic Surgery (EACTS) Developed with the special contribution of the European Association of Percutaneous Cardiovascular Interventions (EAPCI). *Eur Heart J*. 2014;35(37): 2541–2619.

129. Laerum E JN, Smith P, Larsen S. Myocardial infarction may induce positive changes in life-style and in the quality of life. *Scand J Prim Health Care*. 1988;6:67-71.
130. Myocardial infarction redefined--a consensus document of The Joint European Society of Cardiology/American College of Cardiology Committee for the redefinition of myocardial infarction. *Eur Heart J*. 2000;21:1502-13.
131. Pedersen ER, Ueland T, Seifert R, et al. Serum osteoprotegerin levels and long-term prognosis in patients with stable angina pectoris. *Atherosclerosis*. 2010;212:644-9.
132. Rasmussen S, Abildstrom SZ, Rosen M and Madsen M. Case-fatality rates for myocardial infarction declined in Denmark and Sweden during 1987-1999. *J Clin Epidemiol*. 2004;57:638-46.
133. Levey AS, Stevens LA, Schmid CH, et al. A new equation to estimate glomerular filtration rate. *Ann Intern Med*. 2009;150:604-12.
134. Ueland PM, Midttun O, Windelberg A, et al. Quantitative profiling of folate and one-carbon metabolism in large-scale epidemiological studies by mass spectrometry. *Clin Chem Lab Med*. 2007;45:1737-45.
135. Holm PI, Ueland PM, Kvalheim G and Lien EA. Determination of choline, betaine, and dimethylglycine in plasma by a high-throughput method based on normal-phase chromatography-tandem mass spectrometry. *Clin Chem*. 2003;49:286-94.
136. Midttun O, Kvalheim G and Ueland PM. High-throughput, low-volume, multianalyte quantification of plasma metabolites related to one-carbon metabolism using HPLC-MS/MS. *Anal Bioanal Chem*. 2013;405:2009-17.
137. Windelberg A, Arseth O, Kvalheim G and Ueland PM. Automated assay for the determination of methylmalonic acid, total homocysteine, and related amino acids in human serum or plasma by means of methylchloroformate derivatization and gas chromatography-mass spectrometry. *Clin Chem*. 2005;51:2103-9.

138. Midttun O, Hustad S, Solheim E, et al. Multianalyte quantification of vitamin B6 and B2 species in the nanomolar range in human plasma by liquid chromatography-tandem mass spectrometry. *Clin Chem*. 2005;51:1206-16.
139. O'Broin S and Kelleher B. Microbiological assay on microtitre plates of folate in serum and red cells. *J Clin Pathol*. 1992;45:344-7.
140. Kelleher BP and Broin SD. Microbiological assay for vitamin B12 performed in 96-well microtitre plates. *J Clin Pathol*. 1991;44:592-5.
141. <http://www.bevital.no>. Access date Jul 01 2015.
142. <http://www.bevital.no>. Bevital - Assessment of nutritional status by vitamin markers. Access date May 13 2015.
143. Meyer K and Ueland PM. Targeted quantification of C-reactive protein and cystatin c and its variants by immuno-MALDI-MS. *Anal Chem*. 2014;86:5807-14.
144. Meyer K, Fredriksen A and Ueland PM. MALDI-TOF MS genotyping of polymorphisms related to 1-carbon metabolism using common and mass-modified terminators. *Clin Chem*. 2009;55:139-49.
145. Biroccio A, Urbani A, Massoud R, et al. A quantitative method for the analysis of glycated and glutathionylated hemoglobin by matrix-assisted laser desorption ionization-time of flight mass spectrometry. *Anal Biochem*. 2005;336:279-88.
146. Wallace TM, Levy JC and Matthews DR. Use and abuse of HOMA modeling. *Diabetes Care*. 2004;27:1487-95.
147. Manger MS, Strand E, Ebbing M, et al. Dietary intake of n-3 long-chain polyunsaturated fatty acids and coronary events in Norwegian patients with coronary artery disease. *Am J Clin Nutr*. 2010;92:244-51.
148. USDA Agricultural Research Service. USDA Database for the Choline Content of Common Foods, Release 2 2008. Access date Jun 29 2015. URL: <http://www.ars.usda.gov/SP2UserFiles/Place/80400525/Data/Choline/Choln02.pdf>
149. Altman DG. *Practical statistics for medical research*. 1 ed. London UK: Chapman & Hall/CRC; 1991.

150. Pintilie M. [An introduction to competing risks analysis]. *Rev Esp Cardiol.* 2011;64:599-605.
151. Scrucca L, Santucci A and Aversa F. Regression modeling of competing risk using R: an in depth guide for clinicians. *Bone Marrow Transplant.* 2010;45:1388-95.
152. D.G. Kleinbaum MK. Modeling Strategy for Assessing Interaction and Confounding. In: K. K. M. Gail, J.M. Samet, A. Tsiatis, W. Wong, ed. *Logistic regression, Statistics for Biology and Health* Atlanta US: Springer; 2010: 206.
153. Vandembroucke JP, von Elm E, Altman DG, et al. Strengthening the Reporting of Observational Studies in Epidemiology (STROBE): explanation and elaboration. *PLoS Med.* 2007;4:e297.
154. Pencina MJ, D'Agostino RB, Sr., D'Agostino RB, Jr. and Vasan RS. Evaluating the added predictive ability of a new marker: from area under the ROC curve to reclassification and beyond. *Stat Med.* 2008;27:157-72; discussion 207-12.
155. SCORE Risk Charts. The European cardiovascular disease risk assessment model. Access date May 15 2015. URL: <http://www.escardio.org/Guidelines-&-Education/Practice-tools/CVD-prevention-toolbox/SCORE-Risk-Charts>.
156. Pencina MJ, D'Agostino RB, Sr. and Steyerberg EW. Extensions of net reclassification improvement calculations to measure usefulness of new biomarkers. *Stat Med.* 2011;30:11-21.
157. Pencina MJ, D'Agostino RB, Pencina KM, et al. Interpreting incremental value of markers added to risk prediction models. *Am J Epidemiol.* 2012;176:473-81.
158. Gueorguieva R and Krystal JH. Move over ANOVA: progress in analyzing repeated-measures data and its reflection in papers published in the Archives of General Psychiatry. *Arch Gen Psychiatry.* 2004;61:310-7.
159. Rosner B. *Fundamentals of Biostatistics.* 7 ed. Boston, USA: Brooks/Cole; 2011.
160. von Elm E, Altman DG, Egger M, et al. The Strengthening the Reporting of Observational Studies in Epidemiology (STROBE) statement: guidelines for reporting observational studies. *PLoS Med.* 2007;4:e296.

161. STROBE Statement - Strengthening the Reporting of Observational studies in Epidemiology. Access date Dec 09 2014.URL: <http://www.strobe-statement.org>.
162. Gallo V, Egger M, McCormack V, et al. STrengthening the Reporting of OBservational studies in Epidemiology--Molecular Epidemiology (STROBE-ME): an extension of the STROBE Statement. *PLoS Med.* 2011;8:e1001117.
163. Bhopal R. *Concepts of epidemiology*. 2 ed: Oxford, UK: Oxford University Press; 2008.
164. Pedersen AG and Ellingsen CL. Data quality in the Causes of Death Registry. *Tidsskr Nor Laegeforen.* 2015;135:768-770.
165. Tripepi G, Jager KJ, Dekker FW, et al. Bias in clinical research. *Kidney Int.* 2008;73:148-53.
166. Clarke R, Shipley M, Lewington S, et al. Underestimation of risk associations due to regression dilution in long-term follow-up of prospective studies. *Am J Epidemiol.* 1999;150:341-53.
167. Koenig W, Sund M, Frohlich M, et al. Refinement of the association of serum C-reactive protein concentration and coronary heart disease risk by correction for within-subject variation over time: the MONICA Augsburg studies, 1984 and 1987. *Am J Epidemiol.* 2003;158:357-64.
168. Cope EL, Shrubsole MJ, Cohen SS, et al. Intraindividual variation in one-carbon metabolism plasma biomarkers. *Cancer Epidemiol Biomarkers Prev.* 2013;22:1894-9.
169. Midttun O, Townsend MK, Nygard O, et al. Most blood biomarkers related to vitamin status, one-carbon metabolism, and the kynurenine pathway show adequate preanalytical stability and within-person reproducibility to allow assessment of exposure or nutritional status in healthy women and cardiovascular patients. *J Nutr.* 2014;144:784-90.
170. Lever M, Atkinson W, Slow S, et al. Plasma and urine betaine and dimethylglycine variation in healthy young male subjects. *Clin Biochem.* 2009;42:706-12.

171. Miettinen OS and Cook EF. Confounding: essence and detection. *Am J Epidemiol.* 1981;114:593-603.
172. Schisterman EF, Cole SR and Platt RW. Overadjustment bias and unnecessary adjustment in epidemiologic studies. *Epidemiology.* 2009;20:488-95.
173. Tabachnick BG and Fidell LS. Using Multivariate Statistics. 5th ed. Boston, USA: Pearson; 2007: 88-91.
174. Holm PI, Ueland PM, et al. Betaine and folate status as cooperative determinants of plasma homocysteine in humans. *Arterioscler Thromb Vasc Biol.* 2005;25:379-85.
175. Kirsch SH, Herrmann W, Rabagny Y and Obeid R. Quantification of acetylcholine, choline, betaine, and dimethylglycine in human plasma and urine using stable-isotope dilution ultra performance liquid chromatography-tandem mass spectrometry. *J Chromatogr B Analyt Technol Biomed Life Sci.* 2010;878:3338-44.
176. Melse-Boonstra A, Holm PI, Ueland PM, et al. Betaine concentration as a determinant of fasting total homocysteine concentrations and the effect of folic acid supplementation on betaine concentrations. *Am J Clin Nutr.* 2005;81:1378-82.
177. Lever M, George PM, Atkinson W, et al. Plasma lipids and betaine are related in an acute coronary syndrome cohort. *PLoS One.* 2011;6:e21666.
178. Smith GD and Ebrahim S. Mendelian randomization: prospects, potentials, and limitations. *Int J Epidemiol.* 2004;33:30-42.
179. C Reactive Protein Coronary Heart Disease Genetics Collaboration (CCGC), Wensley F, Gao P, Burgess S, et al. Association between C reactive protein and coronary heart disease: mendelian randomisation analysis based on individual participant data. *BMJ.* 2011;342:d548.
180. Holmes MV, Asselbergs FW, Palmer TM, et al. Mendelian randomization of blood lipids for coronary heart disease. *Eur Heart J.* 2015;36:539-50.
181. Jansen H, Samani NJ and Schunkert H. Mendelian randomization studies in coronary artery disease. *Eur Heart J.* 2014;35:1917-24.

182. Heil SG, Lievers KJ, Boers GH, et al. Betaine-homocysteine methyltransferase (BHMT): genomic sequencing and relevance to hyperhomocysteinemia and vascular disease in humans. *Mol Genet Metab.* 2000;71:511-9.
183. Streiner DL and Norman GR. Correction for multiple testing: is there a resolution? *Chest.* 2011;140:16-8.
184. Sleight P. Debate: Subgroup analyses in clinical trials: fun to look at - but don't believe them! *Curr Control Trials Cardiovasc Med.* 2000;1:25-27.
185. Stallones RA. The use and abuse of subgroup analysis in epidemiological research. *Prev Med.* 1987;16:183-94.
186. Fleming A. On the antibacterial action of cultures of a penicillium, with special reference to their use in the isolation of B. influenzae. 1929. *Bull World Health Organ.* 2001;79:780-90.
187. Ioannidis JP. Why most published research findings are false. *PLoS Med.* 2005;2:e124.
188. Roffi M, Patrono C, Collet JP, et al. 2015 ESC Guidelines for the management of acute coronary syndromes in patients presenting without persistent ST-segment elevation: Task Force for the Management of Acute Coronary Syndromes in Patients Presenting without Persistent ST-Segment Elevation of the European Society of Cardiology (ESC). *Eur Heart J.* 2016 Jan 14;37(3):267-315
189. Kirchhof P, Sipido KR, Cowie MR, et al.. The continuum of personalized cardiovascular medicine: a position paper of the European Society of Cardiology. *Eur Heart J.* 2014;35:3250-7.
190. McGeechan K, Macaskill P, Irwig L, et al. Assessing new biomarkers and predictive models for use in clinical practice: a clinician's guide. *Arch Intern Med.* 2008;168:2304-10.
191. Vickers AJ and Pepe M. Does the net reclassification improvement help us evaluate models and markers? *Ann Intern Med.* 2014;160:136-7.
192. Leening MJ, Vedder MM, Witteman JC, et al. Net reclassification improvement: computation, interpretation, and controversies: a literature review and clinician's guide. *Ann Intern Med.* 2014;160:122-31.

193. Hustad S, Eussen S, Midttun O, et al. Kinetic modeling of storage effects on biomarkers related to B vitamin status and one-carbon metabolism. *Clin Chem.* 2012;58:402-10.
194. Det kongelige helse- og omsorgsdepartement. Lov om medisinsk og helsefaglig forskning (helseforskningsloven). Access date Jun 29 2015.
URL: <https://lovdata.no/dokument/NL/lov/2008-06-20-44>.
195. Singh PR and Lele SS. Folate gene polymorphisms MTR A2756G, MTRR A66G, and BHMT G742A and risk for coronary artery disease: a meta-analysis. *Genet Test Mol Biomarkers.* 2012;16:471-5.
196. Schartum-Hansen H, Pedersen ER, Svingen GF, et al. Plasma choline, smoking, and long-term prognosis in patients with stable angina pectoris. *Eur J Prev Cardiol.* 2015 May;22(5):606-14.
197. Xie W, Wood AR, Lyssenko V, et al. Genetic variants associated with glycine metabolism and their role in insulin sensitivity and type 2 diabetes. *Diabetes.* 2013;62:2141-50.
198. Johnson AR, Craciunescu CN, Guo Z, et al. Deletion of murine choline dehydrogenase results in diminished sperm motility. *FASEB J.* 2010;24:2752-61.
199. Zhang J, Blusztajn JK and Zeisel SH. Measurement of the formation of betaine aldehyde and betaine in rat liver mitochondria by a high pressure liquid chromatography-radioenzymatic assay. *Biochim Biophys Acta.* 1992;1117:333-9.
200. Suzuki Y, Fausto A, Hruska KA and Avioli LV. Stimulation of phosphatidylcholine biosynthesis in diabetic hypertrophic kidneys. *Endocrinology.* 1987;120:595-601.
201. Wijekoon EP, Brosnan ME and Brosnan JT. Homocysteine metabolism in diabetes. *Biochem Soc Trans.* 2007;35:1175-9.
202. Wang Z, Yao T, Pini M, et al. Betaine improved adipose tissue function in mice fed a high-fat diet: a mechanism for hepatoprotective effect of betaine in nonalcoholic fatty liver disease. *Am J Physiol Gastrointest Liver Physiol.* 2010;298:G634-42.

203. Teng YW, Ellis JM, Coleman RA and Zeisel SH. Mouse betaine-homocysteine S-methyltransferase deficiency reduces body fat via increasing energy expenditure and impairing lipid synthesis and enhancing glucose oxidation in white adipose tissue. *J Biol Chem.* 2012;287:16187-98.
204. Schwab U, Torronen A, Toppinen L, et al. Betaine supplementation decreases plasma homocysteine concentrations but does not affect body weight, body composition, or resting energy expenditure in human subjects. *Am J Clin Nutr.* 2002;76:961-7.
205. Mills EJ, Wu P, Chong G, et al. Efficacy and safety of statin treatment for cardiovascular disease: a network meta-analysis of 170,255 patients from 76 randomized trials. *QJM.* 2011;104:109-24.
206. Larsen S, Stride N, Hey-Mogensen M, et al. Simvastatin effects on skeletal muscle: relation to decreased mitochondrial function and glucose intolerance. *J Am Coll Cardiol.* 2013;61:44-53.
207. Bernini P, Bertini I, Luchinat C, et al. The cardiovascular risk of healthy individuals studied by NMR metabonomics of plasma samples. *J Proteome Res.* 2011;10:4983-92.
208. Sheikh K, Camejo G, Lanne B, et al. Beyond lipids, pharmacological PPARalpha activation has important effects on amino acid metabolism as studied in the rat. *Am J Physiol Endocrinol Metab.* 2007;292:E1157-65.
209. Cannon CP, Blazing MA, Giugliano RP, et al. Ezetimibe added to statin therapy after acute coronary syndromes. *N Engl J Med.* 2015.
210. Navarese EP, Kolodziejczak M, Schulze V, et al. Effects of proprotein convertase subtilisin/kexin type 9 antibodies in adults with hypercholesterolemia: a systematic review and meta-analysis. *Ann Intern Med.* 2015;163:40-51.
211. Sabatine MS, Giugliano RP, Wiviott SD, et al. Efficacy and safety of evolocumab in reducing lipids and cardiovascular events. *N Engl J Med.* 2015;372:1500-9.
212. Jarcho JA and Keaney JF, Jr. Proof That Lower Is Better--LDL Cholesterol and IMPROVE-IT. *N Engl J Med.* 2015;372:2448-50.

213. Armstrong MJ, Adams LA, Canbay A and Syn WK. Extrahepatic complications of nonalcoholic fatty liver disease. *Hepatology*. 2014;59:1174-97.
214. Lv S, Fan R, Du Y, et al. Betaine supplementation attenuates atherosclerotic lesion in apolipoprotein E-deficient mice. *Eur J Nutr*. 2009;48:205-12.
215. Ding Y, Svingen GF, Pedersen ER, et al. Plasma glycine and risk of acute myocardial infarction in patients with suspected stable angina pectoris. *J Am Heart Assoc*. 2015;5.
216. Liao YJ, Chen TL, Lee TS, et al. Glycine *N*-methyltransferase deficiency affects Niemann-Pick type C2 protein stability and regulates hepatic cholesterol homeostasis. *Mol Med*. 2012;18:412-22.
217. Chen CY, Ching LC, Liao YJ, et al. Deficiency of glycine *N*-methyltransferase aggravates atherosclerosis in apolipoprotein E-null mice. *Mol Med*. 2012;18:744-52.
218. Floegel A, Stefan N, Yu Z, et al. Identification of serum metabolites associated with risk of type 2 diabetes using a targeted metabolomic approach. *Diabetes*. 2013;62:639-48.
219. Thomas GS, Cromwell WC, Ali S, et al. Mipomersen, an apolipoprotein B synthesis inhibitor, reduces atherogenic lipoproteins in patients with severe hypercholesterolemia at high cardiovascular risk: a randomized, double-blind, placebo-controlled trial. *J Am Coll Cardiol*. 2013;62:2178-84.
220. Voight BF, Peloso GM, Orho-Melander M, et al. Plasma HDL cholesterol and risk of myocardial infarction: a mendelian randomisation study. *Lancet*. 2012;380:572-80.
221. Haase CL, Tybjaerg-Hansen A, Nordestgaard BG and Frikke-Schmidt R. HDL cholesterol and risk of type 2 diabetes: a Mendelian randomization study. *Diabetes*. 2015;64:3328-33.
222. Mikael LG, Pancer J, Wu Q and Rozen R. Disturbed one-carbon metabolism causing adverse reproductive outcomes in mice is associated with altered expression of apolipoprotein AI and inflammatory mediators PPARalpha, interferon-gamma, and interleukin-10. *J Nutr*. 2012;142:411-8.

223. Schwahn BC, Wang XL, Mikael LG, et al. Betaine supplementation improves the atherogenic risk factor profile in a transgenic mouse model of hyperhomocysteinemia. *Atherosclerosis*. 2007;195:e100-7.
224. Dahlhoff C, Desmarchelier C, Sailer M, et al. Hepatic methionine homeostasis is conserved in C57BL/6N mice on high-fat diet despite major changes in hepatic one-carbon metabolism. *PLoS One*. 2013;8:e57387.
225. Obeid R and Herrmann W. Homocysteine and lipids: *S*-adenosyl methionine as a key intermediate. *FEBS Lett*. 2009;583:1215-25.
226. Glier MB, Green TJ and Devlin AM. Methyl nutrients, DNA methylation, and cardiovascular disease. *Mol Nutr Food Res*. 2014;58:172-82.
227. Zeisel SH. Dietary choline deficiency causes DNA strand breaks and alters epigenetic marks on DNA and histones. *Mutat Res*. 2012;733:34-8.
228. Kavurma MM, Bhindi R, Lowe HC, et al. Vessel wall apoptosis and atherosclerotic plaque instability. *J Thromb Haemost*. 2005;3:465-72.
229. Russo GL and Russo M. Ins and outs of apoptosis in cardiovascular diseases. *Nutr Metab Cardiovasc Dis*. 2003;13:291-300.
230. Rhodes CJ. Type 2 diabetes-a matter of beta-cell life and death? *Science*. 2005;307:380-4.
231. Zeisel SH, Zola T, daCosta KA and Pomfret EA. Effect of choline deficiency on *S*-adenosylmethionine and methionine concentrations in rat liver. *Biochem J*. 1989;259:725-9.
232. Teng YW, Mehedint MG, Garrow TA and Zeisel SH. Deletion of betaine-homocysteine *S*-methyltransferase in mice perturbs choline and 1-carbon metabolism, resulting in fatty liver and hepatocellular carcinomas. *J Biol Chem*. 2011;286:36258-67.
233. Cacciapuoti F. Hyper-homocysteinemia: a novel risk factor or a powerful marker for cardiovascular diseases? Pathogenetic and therapeutical uncertainties. *J Thromb Thrombolysis*. 2011;32:82-8.
234. Niculescu MD and Zeisel SH. Diet, methyl donors and DNA methylation: interactions between dietary folate, methionine and choline. *J Nutr*. 2002;132:2333S-2335S.

235. Baccarelli A, Rienstra M and Benjamin EJ. Cardiovascular epigenetics: basic concepts and results from animal and human studies. *Circ Cardiovasc Genet*. 2010;3:567-73.
236. Lund G and Zaina S. Atherosclerosis: an epigenetic balancing act that goes wrong. *Curr Atheroscler Rep*. 2011;13:208-14.
237. Bollati V, Schwartz J, Wright R, et al. Decline in genomic DNA methylation through aging in a cohort of elderly subjects. *Mech Ageing Dev*. 2009;130:234-9.
238. Kraus D, Yang Q, Kong D, et al. Nicotinamide N-methyltransferase knockdown protects against diet-induced obesity. *Nature*. 2014;508:258-62.
239. Kathirvel E, Morgan K, Nandgiri G, et al. Betaine improves nonalcoholic fatty liver and associated hepatic insulin resistance: a potential mechanism for hepatoprotection by betaine. *Am J Physiol Gastrointest Liver Physiol*. 2010;299:G1068-77.
240. Chen J, Zhou X, Wu W, et al. FTO-dependent function of N6-methyladenosine is involved in the hepatoprotective effects of betaine on adolescent mice. *J Physiol Biochem*. 2015.
241. Lever M, McEntyre CJ, George PM, et al. Extreme urinary betaine losses in type 2 diabetes combined with bezafibrate treatment are associated with losses of dimethylglycine and choline but not with increased losses of other osmolytes. *Cardiovasc Drugs Ther*. 2014;28:459-68.
242. Dellow WJ, Chambers ST, Lever M, et al. Elevated glycine betaine excretion in diabetes mellitus patients is associated with proximal tubular dysfunction and hyperglycemia. *Diabetes Res Clin Pract*. 1999;43:91-9.
243. Dellow WJ, Chambers ST, Barrell GK, et al. Glycine betaine excretion is not directly linked to plasma glucose concentrations in hyperglycaemia. *Diabetes Res Clin Pract*. 2001;52:165-9.
244. Matheson A, Willcox MD, Flanagan J and Walsh BJ. Urinary biomarkers involved in type 2 diabetes: a review. *Diabetes Metab Res Rev*. 2010;26:150-71.

245. Lever M, McEntyre CJ, George PM, et al. Fenofibrate causes elevation of betaine excretion but not excretion of other osmolytes by healthy adults. *J Clin Lipidol.* 2014;8:433-40.
246. Mudd SH, Ebert MH and Scriver CR. Labile methyl group balances in the human: the role of sarcosine. *Metabolism.* 1980;29:707-20.
247. Glorieux FH, Scriver CR, Delvin E and Mohyuddin F. Transport and metabolism of sarcosine in hypersarcosinemic and normal phenotypes. *J Clin Invest.* 1971;50:2313-22.
248. Yeo EJ and Wagner C. Tissue distribution of glycine *N*-methyltransferase, a major folate-binding protein of liver. *Proc Natl Acad Sci U S A.* 1994;91:210-4.
249. Williams KT and Schalinske KL. Tissue-specific alterations of methyl group metabolism with DNA hypermethylation in the Zucker (type 2) diabetic fatty rat. *Diabetes Metab Res Rev.* 2012;28:123-31.
250. Mueller DM, Allenspach M, Othman A, et al. Plasma levels of trimethylamine-*N*-oxide are confounded by impaired kidney function and poor metabolic control. *Atherosclerosis.* 2015;243:638-44.
251. Pinto J, Almeida LM, Martins AS, et al. Prediction of gestational diabetes through NMR metabolomics of maternal blood. *J Proteome Res.* 2015;14:2696-706.
252. Diaz SO, Pinto J, Graca G, et al. Metabolic biomarkers of prenatal disorders: an exploratory NMR metabonomics study of second trimester maternal urine and blood plasma. *J Proteome Res.* 2011;10:3732-42.
253. Warriar M, Shih DM, Burrows AC, et al. The TMAO-Generating Enzyme Flavin monooxygenase 3 is a central regulator of cholesterol balance. *Cell Rep.* 2015.
254. Nagata C, Wada K, Tamura T, et al. Choline and betaine intakes are not associated with cardiovascular disease mortality risk in Japanese men and women. *J Nutr.* 2015;145:1787-92.

References

1. Poirier LA, Brown AT, Fink LM, Wise CK, Randolph CJ, Delongchamp RR, Fonseca VA. Blood *S*-adenosylmethionine concentrations and lymphocyte methylenetetrahydrofolate reductase activity in diabetes mellitus and diabetic neuropathy. *Metabolism* 2001;50:1014-8.
2. Ueland PM. Choline and betaine in health and disease. *J Inher Metab Dis* 2011;34:3-15.
3. Mukherjee S. Betaine and nonalcoholic steatohepatitis: Back to the future? *World J Gastroenterol* 2011;17:3663-4.
4. Tibbetts AS, Appling DR. Compartmentalization of mammalian folate-mediated one-carbon metabolism. *Annu Rev Nutr* 2010;30:57-81.
5. Lever M, Slow S, McGregor DO, Dellow WJ, George PM, Chambers ST. Variability of plasma and urine betaine in diabetes mellitus and its relationship to methionine load test responses: An observational study. *Cardiovasc Diabetol* 2012;11:34.
6. Lever M, George PM, Slow S, Bellamy D, Young JM, Ho M, et al. Betaine and trimethylamine-*N*-oxide as predictors of cardiovascular outcomes show different patterns in diabetes mellitus: An observational study. *PloS One* 2014;9:e114969.
7. Magnusson M, Wang TJ, Clish C, Engstrom G, Nilsson P, Gerszten RE, Melander O. Dimethylglycine deficiency and the development of diabetes mellitus. *Diabetes* 2015;64:3010-6.
8. Wang Z, Klipfell E, Bennett BJ, Koeth R, Levison BS, Dugar B, et al. Gut flora metabolism of phosphatidylcholine promotes cardiovascular disease. *Nature* 2011;472:57-63.
9. Tang WH, Wang Z, Shrestha K, Borowski AG, Wu Y, Troughton RW, et al. Intestinal microbiota-dependent phosphatidylcholine metabolites, diastolic dysfunction, and adverse clinical outcomes in chronic systolic heart failure. *J Card Fail* 2015;21:91-6.
10. Lever M, Slow S. The clinical significance of betaine, an osmolyte with a key role in methyl group metabolism. *Clin Biochem* 2010;43:732-44.

11. Schartum-Hansen H, Ueland PM, Pedersen ER, Meyer K, Ebbing M, Bleie O, et al. Assessment of urinary betaine as a marker of diabetes mellitus in cardiovascular patients. *PloS One* 2013;8:e69454.
12. Svingen GF, Ueland PM, Pedersen EK, Schartum-Hansen H, Seifert R, Ebbing M, et al. Plasma dimethylglycine and risk of incident acute myocardial infarction in patients with stable angina pectoris. *Arterioscler Thromb Vasc Biol* 2013;33:2041-8.
13. Ebbing M, Bleie O, Ueland PM, Nordrehaug JE, Nilsen DW, Vollset SE, et al. Mortality and cardiovascular events in patients treated with homocysteine-lowering B vitamins after coronary angiography: A randomized controlled trial. *JAMA* 2008;300:795-804.
14. Midttun O, Kvalheim G, Ueland PM. High-throughput, low-volume, multianalyte quantification of plasma metabolites related to one-carbon metabolism using hplc-ms/ms. *Anal Bioanal Chem* 2013;405:2009-17.
15. Ueland PM, Midttun O, Windelberg A, Svardal A, Skalevik R, Hustad S. Quantitative profiling of folate and one-carbon metabolism in large-scale epidemiological studies by mass spectrometry. *Clin Chem Lab Med* 2007;45:1737-45.
16. BEVITAL. Assessment of nutritional status by vitamin markers. <http://bevital.no>. Last updated Oct 28 2015. Accessed August 24 2015.
17. USDA Agricultural Research Service. USDA Database for the Choline Content of Common Foods, Release 2 2008. Last updated May 17 2013. Accessed June 29 2015. Available from: www.ars.usda.gov/Services/docs.htm?docid=6232.
18. Manger MS, Strand E, Ebbing M, Seifert R, Refsum H, Nordrehaug JE, et al. Dietary intake of n-3 long-chain polyunsaturated fatty acids and coronary events in Norwegian patients with coronary artery disease. *Am J Clin Nutr* 2010;92:244-51.
19. Svingen GF, Schartum-Hansen H, Ueland PM, Pedersen ER, Seifert R, Ebbing M, et al. Elevated plasma dimethylglycine is a risk marker of mortality in patients with coronary heart disease. *Eur J Prev Cardiol* 2015;22(6):743-52.

20. Gerhard Sulo JI, Stein Emil Vollset, Ottar Nygård, Nina Øyen, Grethe S. Tell. Cardiovascular disease and diabetes mellitus in Norway during 1994-2009 CVDNOR – a nationwide research project. *Nor Epidemiol* 2013;23:101-7.
21. American Diabetes Association. Diagnosis and classification of diabetes mellitus. *Diabetes Care* 2010;33 Suppl 1:S62-9.
22. Rosner B. *Fundamentals of Biostatistics*. 7 ed. Boston, USA: Brooks/Cole; 2011: p. 569.
23. Konstantinova SV, Tell GS, Vollset SE, Nygard O, Bleie O, Ueland PM. Divergent associations of plasma choline and betaine with components of metabolic syndrome in middle age and elderly men and women. *J Nutr* 2008;138:914-20.
24. Diaz SO, Pinto J, Graca G, Duarte IF, Barros AS, Galhano E, et al. Metabolic biomarkers of prenatal disorders: An exploratory NMR metabonomics study of second trimester maternal urine and blood plasma. *J Proteome Res* 2011;10:3732-42.
25. Koutros S, Meyer TE, Fox SD, Issaq HJ, Veenstra TD, Huang WY, et al. Prospective evaluation of serum sarcosine and risk of prostate cancer in the prostate, lung, colorectal and ovarian cancer screening trial. *Carcinogenesis* 2013;34:2281-5.
26. Gloyn AL, Faber JH, Malmodin D, Thanabalasingham G, Lam F, Ueland PM, et al. Metabolic profiling in maturity-onset diabetes of the young (MODY) and young onset type 2 diabetes fails to detect robust urinary biomarkers. *PloS One* 2012;7:e40962.
27. Szendroedi J, Phielix E, Roden M. The role of mitochondria in insulin resistance and type 2 diabetes mellitus. *Nat Rev Endocrinol* 2012;8:92-103.
28. Pajares MA, Perez-Sala D. Betaine homocysteine *S*-methyltransferase: Just a regulator of homocysteine metabolism? *Cell Mol Life Sci* 2006;63:2792-803.
29. Teng YW, Mehedint MG, Garrow TA, Zeisel SH. Deletion of betaine-homocysteine *S*-methyltransferase in mice perturbs choline and 1-carbon metabolism, resulting in fatty liver and hepatocellular carcinomas. *J Biol Chem* 2011;286:36258-67.

30. Zeisel SH. Metabolic crosstalk between choline/1-carbon metabolism and energy homeostasis. *Clin Chem Lab Med* 2013;51:467-75.
31. Bergeron R, Yao J, Woods JW, Zycband EI, Liu C, Li Z, et al. Peroxisome proliferator-activated receptor (PPAR)-alpha agonism prevents the onset of type 2 diabetes in Zucker diabetic fatty rats: A comparison with PPAR gamma agonism. *Endocrinology* 2006;147:4252-62.
32. Dahlhoff C, Desmarchelier C, Sailer M, Furst RW, Haag A, Ulbrich SE, et al. Hepatic methionine homeostasis is conserved in c57bl/6n mice on high-fat diet despite major changes in hepatic one-carbon metabolism. *PloS One* 2013;8:e57387.
33. Wang L, Chen L, Tan Y, Wei J, Chang Y, Jin T, Zhu H. Betaine supplement alleviates hepatic triglyceride accumulation of apolipoprotein E deficient mice via reducing methylation of peroxisomal proliferator-activated receptor alpha promoter. *Lipids Health Dis* 2013;12:34.
34. Luka Z, Mudd SH, Wagner C. Glycine *N*-methyltransferase and regulation of *S*-adenosylmethionine levels. *J Biol Chem* 2009;284:22507-11.
35. Xie W, Wood AR, Lyssenko V, Weedon MN, Knowles JW, Alkayyali S, et al. Genetic variants associated with glycine metabolism and their role in insulin sensitivity and type 2 diabetes. *Diabetes* 2013;62:2141-50.
36. Dellow WJ, Chambers ST, Lever M, Lunt H, Robson RA. Elevated glycine betaine excretion in diabetes mellitus patients is associated with proximal tubular dysfunction and hyperglycemia. *Diabetes Res Clin Pract* 1999;43:91-9.
37. Glorieux FH, Scriver CR, Delvin E, Mohyuddin F. Transport and metabolism of sarcosine in hypersarcosinemic and normal phenotypes. *J Clin Invest* 1971;50(11):2313-22.
38. Csernus K, Lanyi E, Erhardt E, Molnar D. Effect of childhood obesity and obesity-related cardiovascular risk factors on glomerular and tubular protein excretion. *Eur J Pediatr* 2005;164:44-9.
39. Midttun O, Townsend MK, Nygard O, Tvoroger SS, Brennan P, Johansson M, Ueland PM. Most blood biomarkers related to vitamin status, one-carbon metabolism, and the kynurenine pathway show adequate preanalytical stability

- and within-person reproducibility to allow assessment of exposure or nutritional status in healthy women and cardiovascular patients. *J Nutr* 2014;144:784-90.
40. Reaven G. Insulin resistance and coronary heart disease in nondiabetic individuals. *Arterioscler Thromb Vasc Biol* 2012;32:1754-9.
 41. Sattar N, Preiss D, Murray HM, Welsh P, Buckley BM, de Craen AJ, et al. Statins and risk of incident diabetes: A collaborative meta-analysis of randomised statin trials. *Lancet* 2010;375:735-42.
 42. Tabak AG, Herder C, Rathmann W, Brunner EJ, Kivimaki M. Prediabetes: A high-risk state for diabetes development. *Lancet* 2012;379:2279-90.
 43. Balion CM, Raina PS, Gerstein HC, Santaguida PL, Morrison KM, Booker L, Hunt DL. Reproducibility of impaired glucose tolerance (IGT) and impaired fasting glucose (IFG) classification: A systematic review. *Clin Chem Lab Med* 2007;45:1180-5.
 44. Leening MJ, Vedder MM, Wittteman JC, Pencina MJ, Steyerberg EW. Net reclassification improvement: Computation, interpretation, and controversies: A literature review and clinician's guide. *Ann Int Med* 2014;160:122-31.
 45. Duprez DA, Otvos J, Tracy RP, Feingold KR, Greenland P, Gross MD, et al. High-density lipoprotein subclasses and noncardiovascular, noncancer chronic inflammatory-related events versus cardiovascular events: The multi-ethnic study of atherosclerosis. *J Am Heart Assoc* 2015;14;4(9): e002295.

Table 1. Baseline characteristics according to type 2 diabetes at baseline

	Type 2 diabetes at baseline		<i>P</i> ^b
	No	Yes	
	<i>n</i> ^a	<i>n</i> ^a	
Age, years	3621 62 (55, 69)	449 65 (58, 72)	<0.000001
Male gender, n (%)	3621 2603 (71.9)	449 325 (72.4)	0.48
Prior CVD, n (%)	3621 2044 (56.4)	449 298 (66.4)	0.001
Current smoking, n (%)	3621 1175 (32.4)	449 113 (25.2)	0.09
Estimated total daily intake	1735	204	
energy, kcal	2052 (1663, 2503)	1912 (1530, 2384)	0.12
choline, mg	242 (193, 302)	240 (182, 299)	0.83
betaine, mg	136 (105, 169)	133 (104, 168)	0.20
BMI, kg/m ²	3618 26.1 (24.1, 28.7)	449 28.1 (25.4, 31.4)	<0.000001
HbA1c, %	3621 6.0 (5.3, 6.6)	449 7.7 (6.7, 8.9)	<0.000001
Plasma glucose, mmol/L	3621 5.5 (5.0, 6.2)	449 10.0 (7.7, 12.5)	<0.000001
HOMA-2	877 ^c		
β-cell function, %	53 (43, 80)	-	-
insulin sensitivity, %	238 (88, 265)	-	-
insulin resistance	0.40 (0.40, 1.10)	-	-

eGFR, mL/min per 1.73m ²	3621	91 (79, 99)	448	90 (74, 99)	<0.001
Plasma/serum					
CRP, mg/L	3620	1.74 (0.85, 3.51)	449	2.15 (1.09, 4.81)	0.009
HDL-C, mmol/L	3620	1.26 (1.01, 1.50)	449	1.10 (0.90, 1.30)	<0.000001
triglycerides, mmol/L	3617	1.47 (1.06, 2.09)	449	1.80 (1.29, 2.63)	<0.000001
ALT, IU/L	3025	28 (20, 38)	378	30 (22, 42)	0.005
total homocysteine, μ mol/L	3621	10.4 (8.7, 12.5)	449	10.7 (8.6, 12.9)	0.30
methionine, μ mol/L	3621	25.6 (22.5, 31.9)	449	26.7 (22.5, 33.1)	0.31
choline metabolites, μ mol/L					
TMAO	3610	5.6 (3.6, 9.3)	446	7.2 (4.3, 12.3)	0.0004
choline	3621	9.6 (8.2, 11.4)	449	10.1 (8.4, 12.2)	0.10
betaine	3621	39.4 (32.5, 48.1)	449	35.6 (28.3, 45.0)	<0.000001
dimethylglycine	3621	4.1 (3.4, 5.1)	449	4.2 (3.2, 5.2)	0.88
sarcosine	3345	1.5 (1.2, 1.8)	422	1.4 (1.1, 1.8)	0.92
B-vitamins					
riboflavin, nmol/L	3605	11.0 (7.4, 18.0)	442	12.9 (8.6, 21.2)	<0.001
folate, nmol/L	3619	10.0 (7.3, 14.6)	449	10.8 (7.9, 15.6)	0.008
cobalamin, pmol/L	3181	362 (275, 466)	400	358 (270, 464)	0.53

Urine							
PLP, nmol/L	3605	41.5 (29.6, 59.8)	442	39.0 (27.4, 59.4)	0.21		
choline metabolites, mmol/mol creatinine							
choline	3242	1.98 (1.47-2.69)	399	2.81 (1.85, 4.28)	<0.000001		
betaine	3242	6.96 (4.67-42.25)	399	22.27 (10.21, 45.25)	<0.000001		
dimethylglycine	3242	3.0 (1.98, 4.64)	399	5.61 (3.50, 8.09)	<0.000001		
sarcosine	3242	0.13 (0.09, 0.20)	398	0.25 (0.15, 0.42)	<0.000001		
albumin, g/mmol	2952	0.52 (0.38, 0.86)	375	1.01 (0.55, 3.06)	<0.000001		
creatinine							
Medications and supplements, n (%)							
prior to baseline							
statin	3621	2601 (71.8)	449	349 (77.9)	0.009		
folic acid	3407	313 (9.2)	408	42 (10.3)	0.39		
multivitamins	3407	520 (15.3)	408	46 (11.3)	0.05		
at discharge from hospital							
statin	3621	2875 (79.4)	-	-	-		
folic acid	3407 ^d	226 (6.6)	-	-	-		
multivitamin	3407 ^d	203 (6.0)	-	-	-		

ALT, alanine aminotransferase; BMI, body mass index; CRP, C-reactive protein; CVD, cardiovascular disease; eGFR, estimated glomerular filtration rate; HbA1c, glycated hemoglobin; HDL-C, high density lipoprotein cholesterol; HOMA2, homeostatic model assessment; PLP, 5'-pyridoxal phosphate; TMAO, trimethylamine *N*-oxide.

^aPatients with valid measurements

^bAdjusted for age, gender and fasting status

^cFasting patients without established diabetes

^dPatients included in the Western Norway B-Vitamin Intervention Trial were instructed not to use any additional vitamin supplements

Table 2. Baseline characteristics according to incident type 2 diabetes during follow-up

	Type 2 diabetes during follow-up		<i>P</i> ^b
	No	Yes	
	<i>n</i> ^a	<i>n</i> ^a	
Age, years	3388	233	0.06
Male gender, n (%)	62 (54, 69)	62 (56, 70)	
Prior CVD, n (%)	3388	233	0.61
Current smoking, n (%)	2434 (71.8)	169 (72.5)	
Estimated total daily intake	3388	233	0.06
energy, kcal	1103 (32.6)	72 (30.9)	
choline, mg	1605	130	0.96
betaine, mg	2053 (1672, 2509)	1997 (1596, 2418)	0.77
BMI, kg/m ²	242 (193, 302)	242 (182, 301)	0.71
HbA1c, %	136 (105, 169)	135 (105, 169)	0.59
Plasma glucose, mmol/L	3385	233	<0.00001
HOMA-2	3388	233	<0.00001
β-cell function, %	3388	233	<0.00001
insulin sensitivity, %	5.9 (5.3, 6.6)	6.2 (5.5, 7.0)	
insulin resistance	3388	233	<0.00001
	835	42	
	53 (43, 80)	67 (46, 106)	0.19
	247 (90, 266)	101 (39, 219)	<0.00001
	0.40 (0.40, 1.10)	1.00 (0.48, 2.55)	<0.00001

eGFR, mL/min per 1.73m ²	3388	91 (79, 99)	233	91 (80, 98)	0.13
Plasma/serum					
CRP, mg/L	3387	1.71 (0.84, 3.43)	233	2.34 (1.17, 4.32)	0.09
HDL-C, mmol/L	3387	1.30 (1.08, 1.50)	233	1.10 (0.90, 1.30)	<0.00001
triglycerides, mmol/L	3384	1.43 (1.05, 2.05)	233	1.96 (1.35, 2.69)	<0.00001
ALT, IU/L	2835	27 (20, 38)	190	32 (24, 48)	0.008
total homocysteine, µmol/L	3388	10.4 (8.7, 12.5)	233	10.5 (8.6, 12.9)	0.79
methionine, µmol/L	3388	26.5 (22.5, 31.8)	233	26.9 (22.4, 33.9)	0.08
choline metabolites, µmol/L					
TMAO	3378	5.5 (3.6, 9.3)	232	5.9 (4.2, 9.4)	0.77
choline	3388	9.6 (8.2, 11.4)	233	10.0 (8.2, 11.4)	0.73
betaine	3388	39.5 (32.8, 48.2)	233	36.9 (28.8, 44.6)	0.0009
dimethylglycine	3388	4.1 (3.4, 5.1)	233	4.2 (3.4, 5.2)	0.56
sarcosine	3127	1.5 (1.2, 1.8)	218	1.5 (1.2, 1.8)	0.84
B-vitamins					
riboflavin, nmol/L	3372	11.0 (7.3, 17.8)	233	12.2 (7.7, 20.2)	0.42
folate, nmol/L	3386	10.0 (7.3, 14.6)	233	10.2 (7.8, 15.6)	0.78
cobalamin, pmol/L	2969	363 (275, 467)	212	350 (275, 445)	0.27
PLP, nmol/L	3372	41.5 (29.7, 59.9)	233	41.8 (28.4, 59.8)	0.21
Urine					

choline metabolites, mmol/mol creatinine						
choline	3037	1.97 (1.47, 2.69)	205	2.03 (1.45, 2.93)	1.00	
betaine	3037	6.91 (4.67, 10.86)	205	8.18 (4.97, 13.60)	<0.00001	
dimethylglycine	3037	3.02 (1.96, 4.56)	205	3.32 (2.15, 5.43)	0.61	
sarcosine	3037	0.13 (0.08, 0.20)	205	0.16 (0.09, 0.24)	<0.00001	
albumin, g/mmol creatinine	2778	0.51 (0.37, 0.84)	192	0.67 (0.46, 1.33)	0.89	
Medications and supplements, n (%)						
prior to baseline						
statin	3388	2432 (71.8)	233	169 (72.5)	0.76	
folic acid	3184	294 (9.2)	223	19 (8.5)	0.74	
multivitamins	3184	488 (15.3)	223	32 (14.3)	0.73	
at discharge from hospital						
statin	3388	2678 (79.0)	233	197 (84.5)	0.05	
folic acid	3184 ^d	215 (6.8)	223	11 (4.9)	0.71	
multivitamin	3184 ^d	191 (6.0)	223	12 (5.4)	0.76	

AL.T, alanine aminotransferase; BMI, body mass index; CRP, C-reactive protein; CVD, cardiovascular disease; eGFR, estimated glomerular filtration rate; HbA1c, glycated hemoglobin; HDL-C, high density lipoprotein cholesterol; HOMA2, homeostatic model assessment; PLP, 5'-pyridoxal phosphate; TMAO, trimethylamine N-oxide.

^aPatients with valid measurements

^bAdjusted for age, gender and fasting status

^cFasting patients without established diabetes

^dPatients included in the Western Norway B-Vitamin Intervention Trial were instructed not to use any additional vitamin supplements

Table 3. The associations between systemic and urine choline metabolites with incident type 2 diabetes

Odds ratio ^a (95% confidence interval)						
	Univariate	<i>P</i>	Adjusted for age, gender and fasting status	<i>P</i>	Multivariate model ^b	<i>P</i>
Plasma/serum						
TMAO	1.05 (0.92, 1.20)	0.46	1.02 (0.88, 1.17)	0.81	1.08 (0.91, 1.27)	0.39
choline	1.06 (0.93, 1.22)	0.37	1.01 (0.87, 1.16)	0.94	0.89 (0.75, 1.06)	0.19
betaine	0.78 (0.69, 0.89)	<0.001	0.72 (0.62, 0.83)	<0.00001	0.74 (0.63, 0.88)	<0.001
dimethylglycine	0.99 (0.86, 1.13)	0.83	0.94 (0.82, 1.09)	0.42	0.92 (0.77, 1.09)	0.33
sarcosine	0.97 (0.84, 1.11)	0.65	0.93 (0.81, 1.08)	0.34	1.07 (0.91, 1.26)	0.43
Urine^c						
choline	1.09 (0.96, 1.25)	0.20	1.07 (0.92, 1.23)	0.39	1.09 (0.93, 1.29)	0.30
betaine	1.27 (1.11, 1.45)	0.001	1.25 (1.09, 1.43)	0.001	1.23 (1.06, 1.43)	0.006
dimethylglycine	1.23 (1.07, 1.42)	0.003	1.22 (1.06, 1.40)	0.007	1.19 (1.01, 1.39)	0.03
sarcosine	1.31 (1.14, 1.50)	<0.001	1.30 (1.13, 1.49)	<0.001	1.25 (1.07, 1.46)	0.004

TMAO, trimethylamine *N*-oxide.

^aPer 1 standard deviation of logarithmically transformed variable

^bIncluding age, gender, fasting status, BMI, HbA1c, estimated GFR, CRP, HDL-cholesterol, urine albumin-to-creatinine ratio, and the use of loop diuretics, thiazides, beta blockers, statins and angiotensin converting enzyme

inhibitors and/or angiotensin receptor blockers.

^cCorrected for urine creatinine.

Table 4. Model discrimination and reclassification

	C-statistic (95% CI)	P	NRI>0 (95% CI)	P	IDI (95% CI)	P
Basic model ^a	0.750 (0.714, 0.786)		-		-	-
+ plasma	0.751 (0.715, 0.787)	0.79	0.33 (0.19, 0.47)	<0.000001	0.0084 (0.0038, 0.0130)	<0.001
+ urine	0.757 (0.721, 0.793)	0.19	0.16 (0.01, 0.31)	0.03	0.0048 (0.0002, 0.0094)	0.04

IDI, integrated discrimination improvement; NRI>0, continuous net reclassification index.

^aIncluding age, gender, fasting status, BMI, HbA1c, estimated GFR, CRP, HDL-cholesterol, urine albumin-to-creatinine ratio, and the use of loop diuretics, thiazides, beta blockers, statins and angiotensin converting enzyme inhibitors and/or angiotensin receptor blockers.

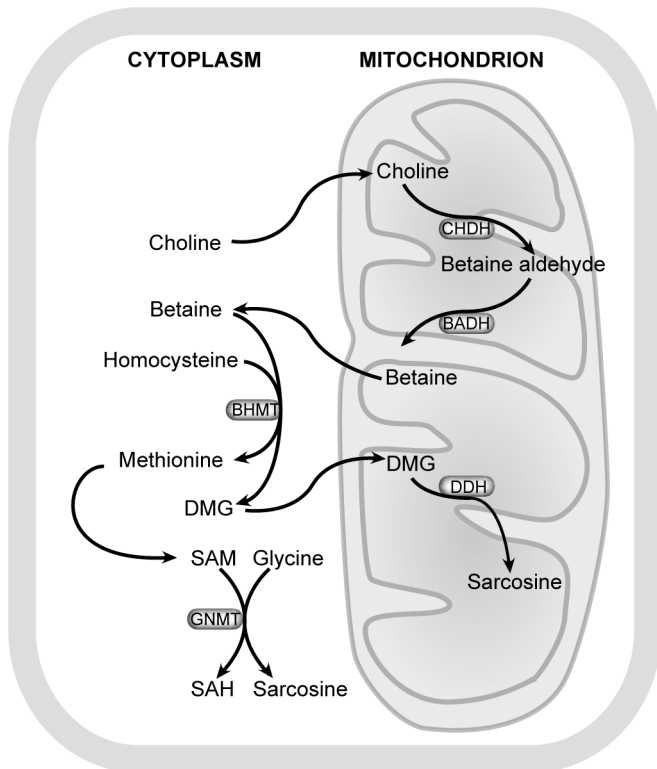


Figure 1. Choline metabolism and its ramifications to homocysteine and methyl group metabolism. BADH, betaine-aldehyde dehydrogenase; BHMT, betaine-homocysteine S-methyl transferase; CHDH, choline dehydrogenase; DDH, dimethylglycine dehydrogenase; DMG, dimethylglycine; GNMT, glycine-*N*-methyltransferase; SAH, *S*-adenosyl-homocysteine; SAM, *S*-adenosylmethionine; SDH, sarcosine dehydrogenase.

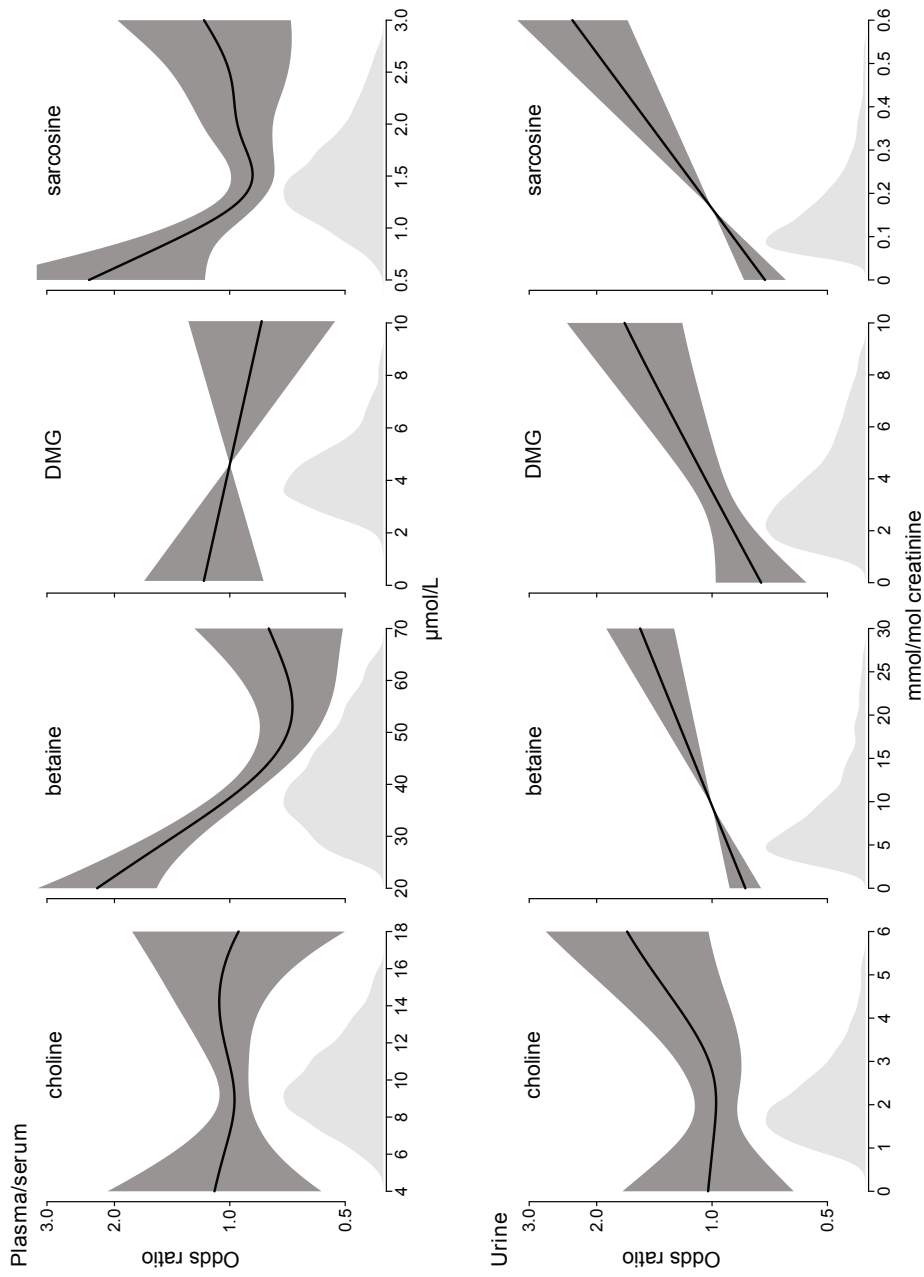
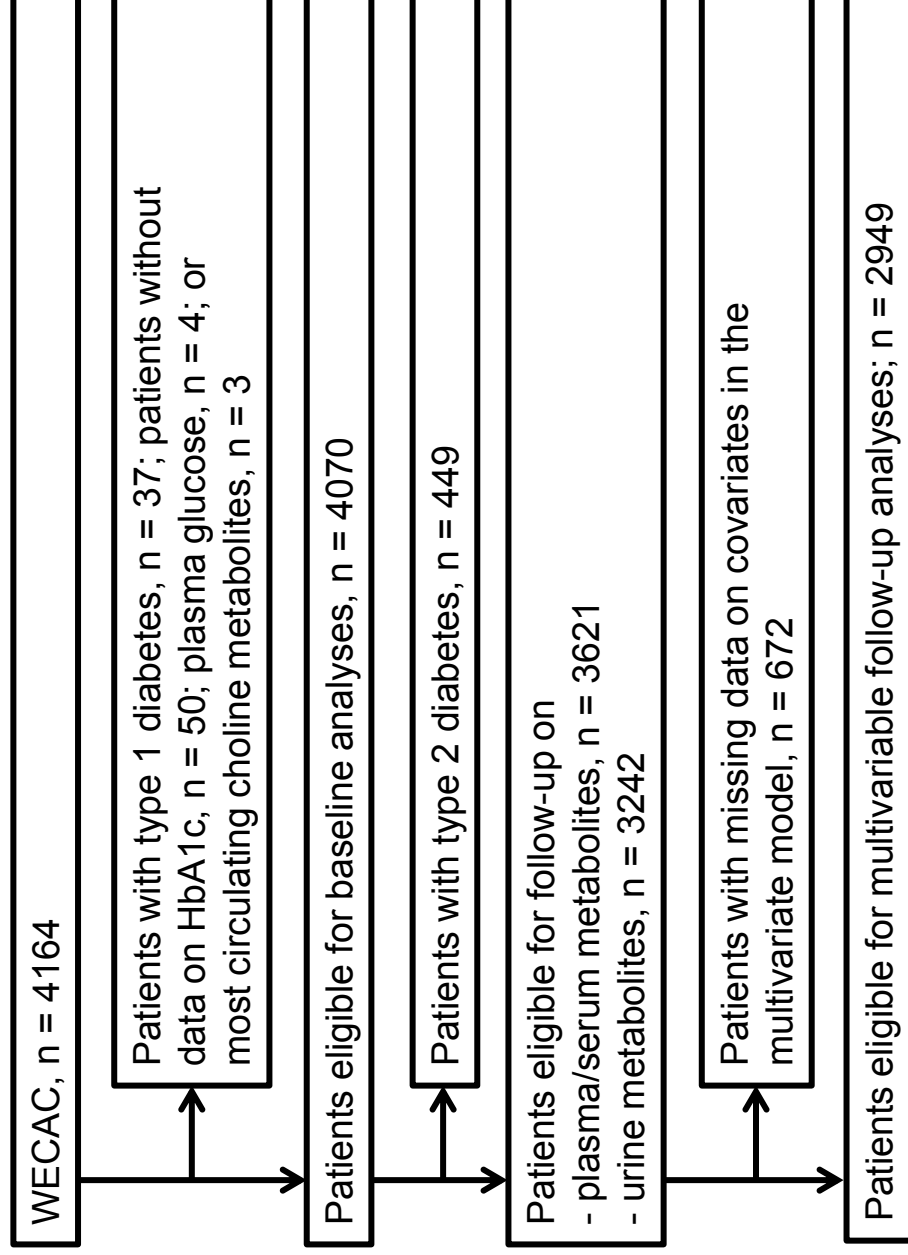
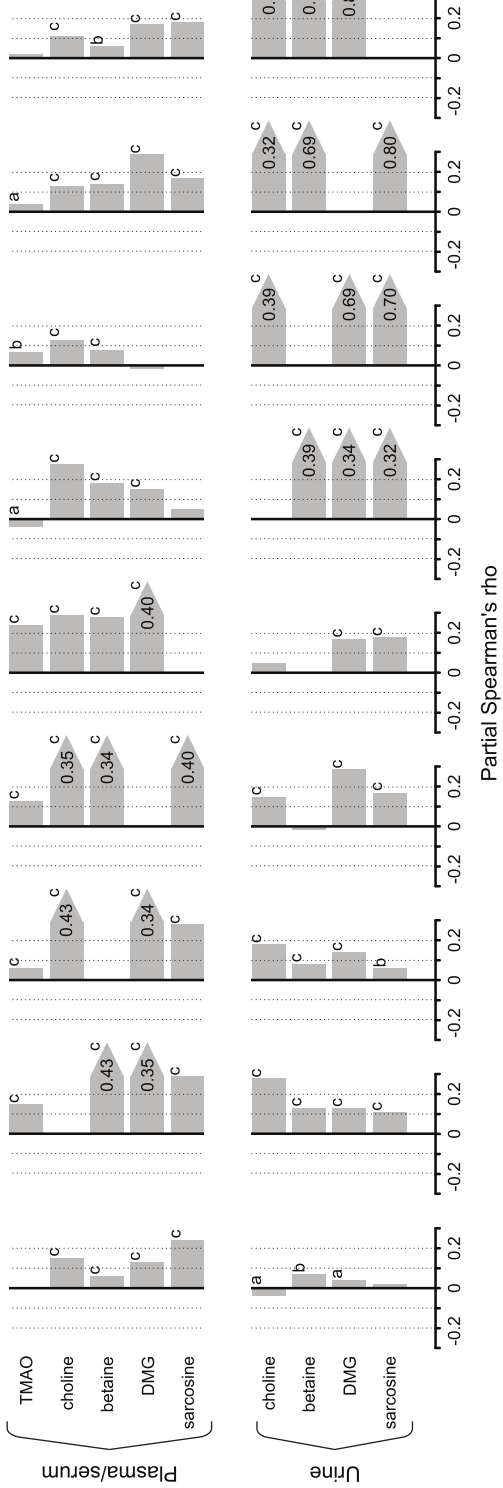
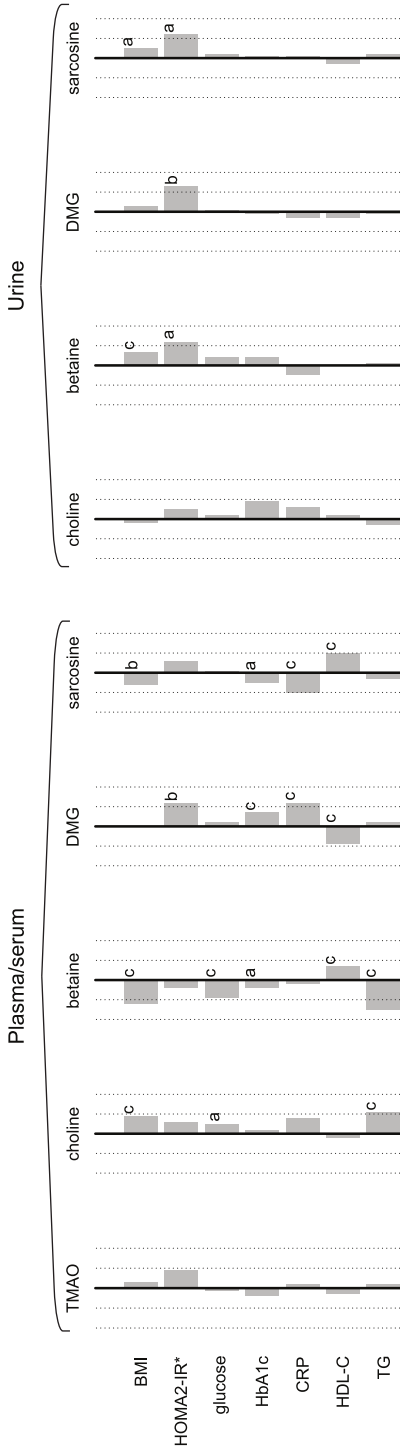


Figure 2. The relationships between systemic and urinary choline related metabolites and incident type 2 diabetes. The solid lines depict the smoothed spline of the generalized additive logistic regressions model, adjusted for age, gender and fasting status. The shaded areas depict 95% confidence intervals. Density plots are aligned along the X-axes.

Supplemental Material



Supplementary Figure 1. Flow-chart over patients eligible for the study. HbA1c, glycated hemoglobin; WECAC, Western Norway Coronary Angiography Cohort.



Supplementary Figure 2. Partial correlation plots between systemic and urinary choline metabolites and indices of insulin resistance among patients without type 2 diabetes at baseline.

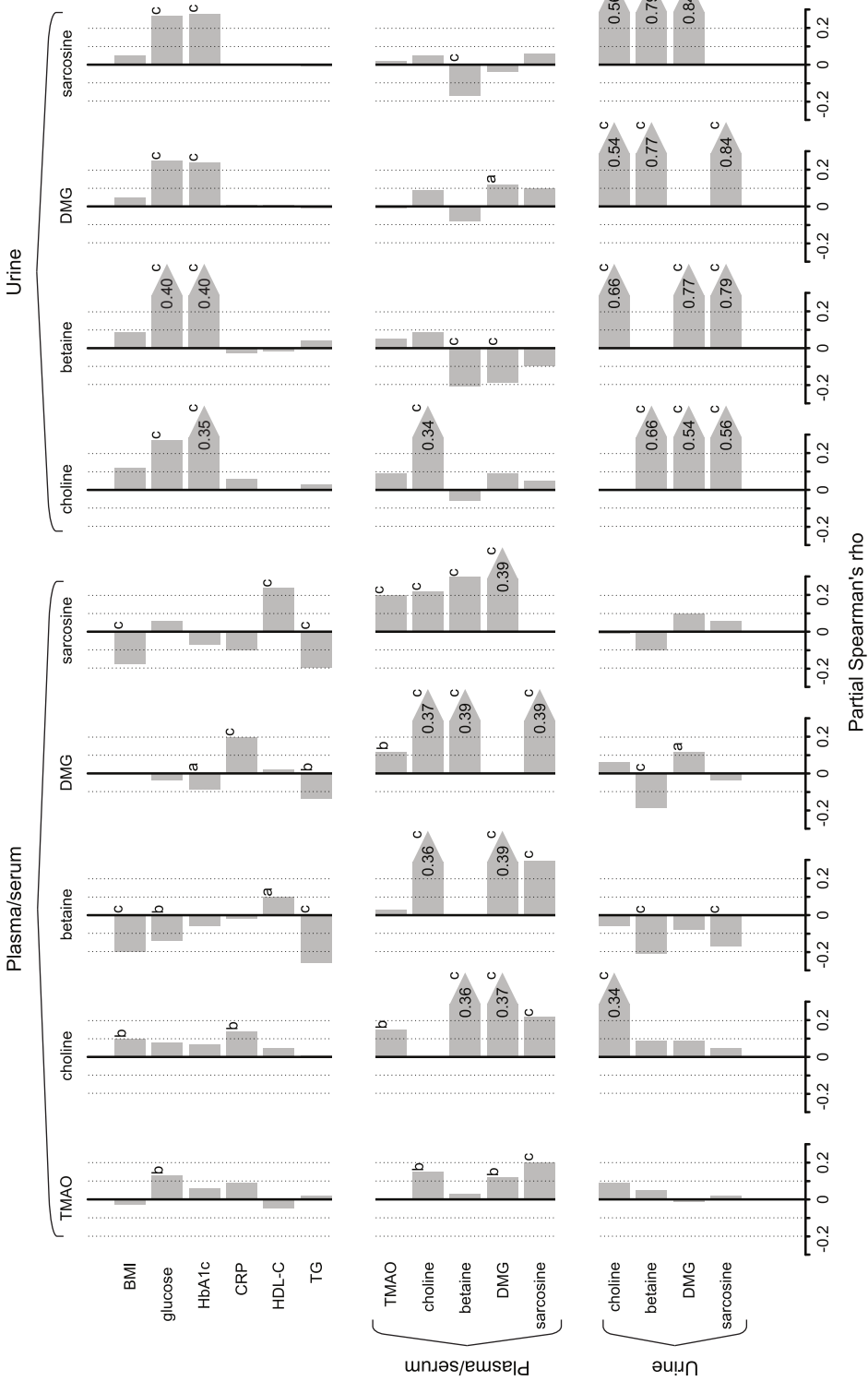
All correlations are adjusted for age, gender and fasting status. Tapered bars indicate partial Spearman's rho > 0.30, with the actual value superimposed.

BMI, body mass index; CRP, C-reactive protein; DMG, dimethylglycine; HDL-C, high-density lipoprotein cholesterol; HOMA2-IR, homeostatic model assessment of insulin resistance; TG, triglycerides; TMAO, trimethylamine N-oxide.

^a $P < 0.05$

^b $P < 0.01$

^c $P < 0.001$



Supplementary Figure 3. Partial correlation plots between systemic and urinary choline metabolites and indices of insulin resistance among patients with type 2 diabetes at baseline. All correlations are adjusted for age, gender and fasting status. Tapered bars indicate partial Spearman's rho > 0.30, with the actual value superimposed. BMI, body mass index; CRP, C-reactive protein; DMG, dimethylglycine; HDL-C, high-density lipoprotein cholesterol; TG, triglycerides; TMAO, trimethylamine N-oxide.

^aP<0.05
^bP<0.01
^cP<0.001

Supplementary table 1. Baseline characteristics according to no, possible and established type 2 diabetes

	Type 2 diabetes at baseline		
	No	Possible	Established
	n ^a	n ^a	n ^a
Age, years	62 (54, 69)	62 (55, 70)	65 (58, 72) ^f
Male gender, n (%)	1838 (73.1)	765 (69.0) ^e	325 (72.4)
Prior CVD, n (%)	1428 (56.8)	616 (55.6)	298 (66.4) ^e
Current smoking, n (%)	806 (32.1)	369 (33.3)	113 (25.2)
Estimated total daily intake	1343	392	204
energy, kcal	2038 (1658, 2498)	2091 (1691, 2528)	1912 (1530, 2384)
choline, mg	240 (191, 301)	250 (199, 304)	240 (182, 299)
betaine, mg	135 (104, 169)	138 (106, 169)	133 (104, 168)
BMI, kg/m ²	2511 26.0 (23.9, 28.4)	1107 26.6 (24.4, 29.2) ^f	449 28.1 (25.4, 31.4) ^f
HbA1c, %	2513 5.6 (5.0, 6.0)	1108 7.0 (6.7, 7.5) ^f	449 7.7 (6.7, 8.9) ^f
Plasma glucose, mmol/L	2513 5.4 (5.0, 6.1)	1108 5.7 (5.1, 6.5) ^f	449 10.0 (7.7, 12.5) ^f
HOMA-2	568 ^b	309 ^b	

β-cell function, %	53 (43, 78)	51 (40-84)	-
insulin sensitivity, %	253 (94, 266)	227 (76-263)	-
insulin resistance	0.40 (0.40, 1.10)	0.40 (0.40-1.30)	-
eGFR, mL/min per 1.73m ²	2513	91 (79, 99)	1108
Plasma/serum		90 (78-99)	448
CRP, mg/L	2513	1.69 (0.83, 3.38)	1107
HDL-C, mmol/L	2512	1.26 (1.02, 1.50)	1108
triglycerides, mmol/L	2510	1.44 (1.06, 2.03)	1107
ALT, U/L	2105	27 (20, 38)	920
total homocysteine, μmol/L	2513	10.4 (8.7, 12.5)	1108
methionine, μmol/L	2513	26.8 (22.6, 32.1)	1108
choline metabolites, μmol/L		26.1 (22.1, 31.2) ^d	449
TMAO	2504	5.6 (3.6, 9.4)	1106
choline	2513	9.6 (8.2, 11.4)	1108
betaine	2513	39.7 (33.0, 48.4)	1108
dimethylglycine	2513	4.1 (3.4, 5.0)	1108
		2.15 (1.09, 4.81) ^f	449
		1.10 (0.90, 1.30) ^f	449
		1.80 (1.29, 2.63) ^f	449
		30 (22, 42) ^f	378
		10.7 (8.6, 12.9)	449
		26.7 (22.5, 33.1)	449
		7.2 (4.3, 12.3) ^d	446
		10.1 (8.4, 12.2) ^d	449
		35.6 (28.3, 45.0) ^f	449
		4.2 (3.2, 5.2)	449

sarcosine	2274	1.5 (1.2, 1.9)	1071	1.4 (1.2, 1.8)	422	1.4 (1.1, 1.8) ^e
B-vitamins						
riboflavin, nmol/L	2502	11.0 (7.3, 17.8)	1103	11.2 (7.6, 18.4)	442	12.9 (8.6, 21.2)
folate, nmol/L	2511	10.0 (7.2, 14.9)	1108	10.0 (7.6, 14.1)	449	10.8 (7.9, 15.6) ^d
cobalamin, pmol/L	2215	358 (272, 453)	966	374 (283, 499)	400	358 (270, 464)
PLP, nmol/L	2502	42.1 (29.8, 60.4)	1103	40.6 (29.2, 58.2)	442	39.0 (27.4, 59.4)
Urine						
choline metabolites, mmol/mol creatinine	2258		984		399	
choline		1.95 (1.44, 2.63)		2.07 (1.55, 2.86) ^d		2.81 (1.85, 4.28) ^f
betaine		6.77 (4.59, 10.76)		7.38 (4.78, 11.66)		22.27 (10.21, 45.25) ^f
dimethylglycine		3.00 (2.01, 4.51)		3.15 (1.86, 4.92)		5.61 (3.50, 8.09) ^f
sarcosine		0.13 (0.08, 0.20)		0.13 (0.09, 0.20)		0.25 (0.15, 0.42) ^f
albumin, g/mmol creatinine	2106	0.51 (0.37, 0.82)	864	0.57 (0.39, 0.97) ^e	379	1.01 (0.55, 3.06) ^f
Medications and supplements, n (%)						
Prior to baseline						
statin	2513	1790 (71.2)	1108	811 (73.2)	448	349 (77.9) ^e

folic acid	2391	183 (7.7)	1016	130 (12.8) ^f	408	42 (10.3)
multivitamins	2391	367 (15.3)	1016	153 (15.1)	408	46 (11.3) ^d
At discharge from hospital ^c						
statin	2513	2014 (80.1)	1108	861 (77.7)	-	-
folic acid	2391	116 (4.9)	1016	110 (10.8) ^f	-	-
multivitamins	2391	122 (5.1)	1016	81 (8.0) ^f	-	-

ALT, alanine aminotransferase; BMI, body mass index; CRP, C-reactive protein; CVD, cardiovascular disease; eGFR, estimated glomerular filtration

rate; HbA1c, glycated hemoglobin; HDL-C, high density lipoprotein cholesterol; HOMA2, homeostatic model assessment; PLP, 5'-pyridoxal

phosphate; TMAO, trimethylamine *N*-oxide.

^aPatients with valid measurements

^bFasting patients with stable angina pectoris

^cPatients were instructed not to use any additional vitamin supplements after inclusion into the Western Norway B-Vitamin Intervention Trial

^d $P < 0.05$; ^e $P < 0.01$; ^f $P < 0.001$ in mixed linear or logistic regression models, adjusted for age, gender and fasting status. The category "No T2D at

baseline" is used as reference.

Supplementary table 2. Choline metabolites at baseline and after 1 year according to WENBIT study treatment

	Baseline	1 year
Plasma/serum, $\mu\text{mol/L}$		
TMAO		
placebo (ref)	9.0 (8.3, 9.7)	8.5 (7.7, 9.3)
FA+B12+B6	8.2 (7.5, 8.9)	7.6 (6.9, 8.4)
FA+B12	8.6 (7.9, 9.3)	7.9 (7.1, 8.6)
B6	7.7 (6.9, 8.4)	7.6 (6.9, 8.4)
choline		
placebo (ref)	9.8 (9.6, 10.0)	10.4 (10.2, 10.7) ^b
FA+B12+B6	9.7 (9.5, 9.9)	11.0 (10.7, 11.2) ^c
FA+B12	9.7 (9.5, 9.9)	11.0 (10.7, 11.2) ^c
B6	9.9 (9.7, 10.2)	10.5 (10.2, 10.7)
betaine		
placebo (ref)	40.7 (39.6, 41.8)	43.1 (42.0, 44.4) ^b
FA+B12+B6	40.7 (39.6, 41.8)	49.6 (48.4, 50.8) ^d
FA+B12	41.0 (39.9, 42.1)	48.1 (47.0, 49.3) ^d
B6	42.0 (40.8, 43.1)	45.1 (44.0, 46.3)
dimethylglycine		
placebo	4.4 (4.3, 4.6)	4.6 (4.4, 4.7) ^a
FA+B12+B6	4.2 (4.1, 4.4)	4.1 (4.0, 4.3) ^c
FA+B12	4.4 (4.2-4.5)	4.3 (4.2-4.5) ^c
B6	4.2 (4.0-4.3)	4.5 (4.3-4.6) ^c
sarcosine		

placebo	1.63 (1.57-1.68)	1.89 (1.76-2.03) ^b
FA+B12+B6	1.61 (1.56-1.66)	1.55 (1.43-1.68) ^d
FA+B12	1.58 (1.53-1.63)	1.55 (1.41-1.69) ^c
B6	1.59 (1.54-1.64)	1.66 (1.52-1.80) ^c

Urine, mmol/mol creatinine

choline

placebo	1.98 (1.87-2.10)	2.20 (1.09-2.32) ^b
FA+B12+B6	2.13 (2.02-2.24)	2.35 (2.24-2.47)
FA+B12	2.12 (2.01-2.23)	2.29 (2.17-2.40)
B6	2.13 (2.02-2.24)	2.14 (2.03-2.26) ^c

betaine

placebo	10.0 (9.0-10.9)	10.5 (9.6-11.5)
FA+B12+B6	9.3 (8.4-10.2)	11.0 (10.0-11.9)
FA+B12	9.3 (8.4-10.3)	10.0 (9.1-11.0)
B6	9.5 (8.6-10.4)	9.8 (8.9-10.8)

dimethylglycine

placebo	4.1 (3.7-4.4)	4.5 (4.1-4.9) ^b
FA+B12+B6	3.8 (3.4-4.1)	3.6 (3.2-3.9) ^d
FA+B12	3.7 (3.4-4.1)	3.6 (3.2-3.9) ^d
B6	3.6 (3.3-4.0)	3.8 (3.5-4.2)

sarcosine

placebo	0.18 (0.16-0.19)	0.19 (0.18-0.20) ^a
FA+B12+B6	0.16 (0.15, 0.17)	0.13 (0.12, 0.14) ^d
FA+B12	0.17 (0.16, 0.18)	0.13 (0.11, 0.14) ^d
B6	0.16 (0.15, 0.17)	0.18 (0.16, 0.19)

Values are given as mean (95% confidence interval).

B6, vitamin B6; B12, vitamin B12; FA, folic acid; TMAO,

trimethylamine *N*-oxide; WENBIT, Western Norway B-Vitamin

Intervention Trial.

^a $P < 0.05$ for difference from baseline; ^b $P < 0.001$ for difference from

baseline; ^c $P < 0.05$ for difference from placebo; ^d $P < 0.001$ for difference

from placebo.

Supplementary table 3. The associations between systemic and urine choline metabolites, and incident type 2 diabetes, excluding patients with indices of possible type 2 diabetes at baseline

		Odds ratio ^a (95% CI)				
	Univariate	<i>P</i>	Adjusted for	<i>P</i>	Multivariate	<i>P</i>
			age, gender and		model ^b	
			fasting status			
<hr/>						
Plasma/serum						
TMAO	1.18 (1.00, 1.39)	0.05	1.13 (0.95, 1.34)	0.17	1.13 (0.92, 1.40)	0.25
choline	1.08 (0.90, 1.29)	0.40	1.01 (0.84, 1.21)	0.93	0.86 (0.69, 1.08)	0.19
betaine	0.84 (0.71, 1.00)	0.05	0.77 (0.64, 0.93)	0.007	0.83 (0.66, 1.03)	0.09
dimethylglycine	1.05 (0.89, 1.24)	0.57	1.00 (0.83, 1.19)	0.97	1.05 (0.85, 1.30)	0.63
sarcosine	0.94 (0.78, 1.13)	0.50	0.90 (0.75, 1.09)	0.30	1.07 (0.87, 1.33)	0.52
<hr/>						
Urine ^c						
choline	1.00 (0.83, 1.21)	0.98	0.97 (0.79, 1.18)	0.73	1.07 (0.86, 1.32)	0.56
betaine	1.19 (0.99, 1.42)	0.06	1.16 (0.96, 1.39)	0.12	1.18 (0.97, 1.44)	0.09
dimethylglycine	1.19 (0.99, 1.43)	0.06	1.16 (0.97, 1.40)	0.11	1.21 (0.99, 1.49)	0.07

sarcosine	1.22 (1.02, 1.46)	0.03	1.19 (0.98, 1.43)	0.07	1.22 (1.00, 1.50)	0.05
-----------	-------------------	------	-------------------	------	-------------------	------

CI, confidence interval; TMAO, trimethylamine *N*-oxide.

^aPer 1 standard deviation of logarithmically transformed variable

^bIncluding age, gender, fasting status, body mass index, glycoated hemoglobin, estimated glomerular filtration rate,

C-reactive protein, high-density lipoprotein cholesterol, urine albumin-to-creatinine ratio, and the use of loop

diuretics, thiazides, beta blockers, statins and angiotensin converting enzyme inhibitors and/or angiotensin receptor blockers.

^cCorrected for urine creatinine.

Supplementary table 4. The associations between systemic and urine choline metabolites, and incident type 2 diabetes within the WENBIT study population during the in-trial follow-up period

		Odds ratio ^a (95% CI)				
	Univariate	<i>P</i>	Adjusted for age, gender and fasting status	<i>P</i>	Multivariate model ^b	<i>P</i>
Plasma/serum						
TMAO	1.08 (0.86, 1.35)	0.52	1.04 (0.82, 1.31)	0.75	1.06 (0.81, 1.39)	0.65
choline	1.12 (0.88, 1.42)	0.37	1.06 (0.83, 1.37)	0.64	0.98 (0.75, 1.29)	0.91
betaine	0.88 (0.70, 1.11)	0.28	0.79 (0.62, 1.01)	0.06	0.88 (0.68, 1.15)	0.36
dimethylglycine	0.95 (0.75, 1.21)	0.69	0.92 (0.72, 1.19)	0.53	0.86 (0.65, 1.14)	0.30
sarcosine	1.12 (0.88, 1.43)	0.35	0.99 (0.83, 1.19)	0.95	1.11 (0.89, 1.37)	0.36
Urine^c						
choline	1.09 (0.87, 1.36)	0.48	1.15 (0.91, 1.45)	0.25	1.14 (0.89, 1.46)	0.32
betaine	1.33 (1.07, 1.64)	0.009	1.36 (1.10, 1.69)	0.005	1.28 (1.03, 1.59)	0.02
dimethylglycine	1.23 (0.98, 1.54)	0.07	1.27 (1.01, 1.60)	0.04	1.26 (0.99, 1.61)	0.06

sarcosine 1.32 (1.06, 1.65) 0.02 1.36 (1.09, 1.70) 0.007 1.31 (1.04, 1.65) 0.02

CI, confidence interval; TMAO, trimethylamine N-oxide; WENBIT, Western Norway B-Vitamin Intervention

Trial

^aPer 1 standard deviation of logarithmically transformed variable

^bIncluding age, gender, fasting status, body mass index, glycosylated hemoglobin, estimated glomerular filtration rate, C-reactive protein, high-density lipoprotein cholesterol, urine albumin-to-creatinine ratio, and the use of loop diuretics, thiazides, beta blockers, statins and angiotensin converting enzyme inhibitors and/or angiotensin receptor blockers.

^cCorrected for urine creatinine

Supplementary table 5. The intraclass correlation coefficients for plasma betaine and urine sarcosine over 1 year

	ICC (95% CI)			
	Placebo	FA + B12 + B6	FA + B12	B6
Plasma betaine	0.62 (0.56, 0.66)	0.43 (0.36, 0.50)	0.44 (0.37, 0.51)	0.65 (0.60, 0.70)
Urine sarcosine	0.69 (0.64, 0.74)	0.49 (0.41, 0.56)	0.37 (0.28, 0.46)	0.73 (0.68, 0.77)

B6, vitamin B6; B12, vitamin B12; CI, confidence interval; FA, folic acid; ICC, intraclass correlation coefficient.