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X-RAY MICROANALYTICAL STUDIES OF INITIAL MINERALIZATION IN INDUCED HETEROTOPIC BONE FORMATION IN GUINEA PIGS

BY

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RUNNING TITLE:

X-RAY MICROANALYSIS OF INITIAL MINERALIZATION.

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Abstract.

Allogenic demineralized dentin implanted in the abdominal wall of guinea pigs induced heterotopic osteoid and bone formation. Samples of this tissue were frozen at -140° C, freeze-dried at -80° C, infiltrated with Spurr® epoxy resin and polymerized. The sections were studied in the scanning transmission electron microscope, and analytical studies were performed by means of energy dispersive micro-analysis.

Osteoblasts and young osteoid osteocytes contained much more calcium than undifferentiated cells and mature osteocytes. The Ca/P ratio in the cytoplasm of these young matrix-producing cells was high. Phosphorus was found in the nuclei of all cells and to a lesser degree also in the cytoplasm of cells rich in calcium. The initial extra-cellular mineral accumulation also showed a high Ca/P ratio. Small electron-dense areas in these regions exhibited both an amount of calcium and a Ca/P ratio resembling those of mature mineralized bone. The initial event of the mineralization process seemed to be an accumulation of calcium first in the young matrix-producing cells and then in the surrounding osteoid matrix.

The results suggested that the matrix-producing cells are actively involved in the accumulation of calcium and phosphorus both intra-and extra-cellularly.

Key words: electron microscopy
energy dispersive microanalysis
calcium
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Introduction

Mineralization of bone has previously been regarded as an extra-cellular process (16, 34), while a cellular control mechanism has been suggested lately (3, 9). Several authors have found evidence of intra-cellular binding of calcium in mineralizing hard tissue (5, 12, 18, 19), and it has been suggested that matrix-producing cells are involved in passing minerals to the matrix (4, 11, 40). This cellular activity in the mineralizing process has been closely connected with the function of matrix vesicles which are found in mineralizing tissues (2, 3, 10). These membrane bound vesicles seem to be derived from cells involved in the hard tissue formation (30, 35, 36). Crystal-like particles have been found inside such vesicles and may be the first evidence of crystal formation in the mineralization process (3, 10). Alterations in matrix components could, however, also explain the early extra-cellular and extra-vesicular mineralization and crystal formation in the initial phase of mineralization of hard tissues (8, 9, 33). X-ray microanalysis (XMA) combining electron microscopy and energy dispersive x-ray analysis has become a new useful method for elemental analysis of diffusible as well as bound elements in biological material (13, 16). Scanning transmission electron microscopy (STEM) together with XMA of elements in thin sections of bilogical material after careful freezing and freeze-drying should exclude artefacts due to fixation. XMA is applicated to a wide field of biological tissues and should be suitable for microanalysis of the mineralization process (29).

Implantation of demineralized dentin in the abdominal muscles of guinea pigs and rats has been shown to induce heterotopic osteoid, bone and cartilage formation consistently as well as resulting in remineralization of implanted dentin (6, 7, 30). In this experimental model the bone is formed de-novo (7, 37), and areas with various levels of mineralization are found within the same section. Recently, it has been shown by transmission elctron microscopy that the first mineral crystals in heterotopic bone formation are found mostly extra-cellularly although a few crystals appeared inside matrix vesicles (31). The present study was undertaken to use the advantages of this model to investigate, by means of XMA, the accumulation of calcium and phosphorus in osteoblasts and osteocytes as well as in the surrounding matrix during the initial phase of mineralization of heterotopic bone formation.

Material and Methods

Thirty randomly bred male albino and pigmented guinea pigs were used and divided in three equal groups. They were fed a standard mixed diet (Norwegian standard for guinea pigs and rabbits, Statens Institutt for Folkehelse, Oslo, Norway) supplemented with sweedes and hay, and were given water ad libitum. The animals were weighed before the operation (about 500g) and at sacrifice. Dentin was obtained from adult male guinea pigs, demineralized in HCl, freeze-dried and sterilized before the operation (7).

Four specimens of demineralized dentin were implanted in the abdominal muscles of each guinea pig. The operation was carried out under general anesthesia (Nembutal ®). After 16, 21 and 28 days respectively the animals were killed by an overdose of Nembutal ®, and the implants were dissected free together with some surrounding tissue. They were cut in small pieces of 1 mm^3 and immediately frozen in isopentane prechilled with liquid nitrogen to $-140^{\circ}C$. The specimens were stored in liquid nitrogen until further treatment. Later the tissue was freeze-dried in Tis-U-Freeze Dryer ® at -80° C for three days. The freeze-dried material was warmed to room temperature in a nitrogen atmosphere before contact with air, and then embedded in Spurr ® low viscosity resin by vibration in a vacuum desiccator for three days. Sections of 1 µm were cut with glass knives and stained with toluidine blue.

Cutting for further investigation was performed on a Reichert OM U3 ultramicrotome with a diamond knife, and a section thickness of approximately 100 nm. The sections were floated out on the

inert liquid ethylene glycol and quickly picked up on carbon-coated nylon grids. The scanning transmission mode (STEM) on a Philips 500 was used in the investigation. An energy-dispersive spectrometer (Kevex ®) was attached to the microscope. Sections were examined, and areas for analysis were selected using the full line scan of the STEM. During the analysis the beam was stationed on the structure of interest, and x-ray spectra were obtained showing the elemental composition in the selected spot. The microscope was operated at 50 kV, spot-size 125 nm, magnification 5000 X, counting time 100 sec. live time. The dead time was approximately zero. The beam current was controlled by calibration on a standard polished copper specimen.

Results

All the animals gained weight during the experiment. Histologically the implants showed areas of resorption and fibroblastic ingrowth as well as newly formed bone in the surrounding tissues (plate I,A). Remineralization of the dentin could be seen in some areas.

A detailed description of the cellular reactions has been published previously (7, 30). The light microscope findings correlated well with the STEM picture although the freeze-dried material exhibited some morphological freeze artefacts. It was possible to study the various zones in the same sections, and the unstained sections had enough contrast to permit microanalysis both in the nucleus, the cytoplasm and the extra-cellular matrix. Except for the nucleus the different organelles could mot be recognized in the STEM, and in the undifferentiated soft tissue it was rather difficult to distinguish the cytoplasm of the cells and the extra-cellular matrix.

The different types of tissue which were found in each section permitted intra-sectional comparison between different cell types and tissue reactions. The results of microanalysis from selected cells and the surrounding matrix are shown in plates I, II & III.

The undifferentiated tissue showed very little calcium and phosphorus (plate III: B & C) except in the nuclei which had a small peak of phosphorus (plate III: B). In the osteoblast region it was easier to separate extra-cellular matrix from the cytoplasm. The cytoplasm of the osteoblasts showed distinct signals from calcium (plate I: B,2). The nuclei showed distinct phosphorus peaks which were higher than in the cytoplasm, but only low counts for calcium (plate I: B,1).

The cytoplasm of the osteocytes in the osteoid (osteoid osteocytes) exhibited a high count for calcium (plate II: A,2). The nuclei showed a distinct Kα peak for phosphorus (plate II: A,1). The newly formed matrix had a clear Kα peak for Ca, but very little phosphorus resulting in a Ca/P ratio of about 6:1 (plate I: B,4 and plate II: A,3). Small electron-dense nodules were also seen in the osteoid matrix (plate I: B). It seemed that they increased in number and size in areas showing more mineralization. In these nodules there was about 3 times more Ca (plate I: B,3) than in the surrounding matrix (plate I: B,4). An increased level of phosphorus was found resulting in Ca/P ratio of approximately 2:1 which was almost equal to that of fully mineralized bone (plate: II,A,4).

The cytoplasm of the osteocytes in the fully mineralized bone had only very small Kα peaks for calcium (plate III: A,2) and negligible counts for phosphorus except in the nucleus (plate III: A,1).

Microanalysis of the remineralized dentin gave data resembling that from fully mineralized bone. Sections containing only the embedding medium showed only a distinct peak of Cl indicating that the background was low.

Discussion

Calcium has been demonstrated intra-cellularly in hard tissue forming cells by several methods. Both autoradiography (15, 25) and the methods utilizing calcium binding dyes such as glyoxal-bis (2-hydroxyanil) at the light microscopical level (18, 19) or K-pyroantimonate at the ultrastructural level (4,5,12) have been used. The K-pyroantimonate method gives good ultrastructural morphology (5, 20) in contrast to the application of unfixed frozen sections used in our studies. Since the K-pyroantimonate involves fixation by OsO_4, dehydration in alcohols and routine infiltration in epoxy, the signals from osmium will mask the Kα-peaks of phosphorus, and significant redistribution and loss of the actual elements may occur (14, 21). To avoid these disadvantages we used unfixed freeze-dried material and vacuum-infiltration of the epoxy resin. This method provided a satisfactory morphology to perform microanalysis in the cytoplasm and nucleus of the actual cells.

Until about 1970 mineralization was considered mainly an extra-cellular process, although Watt as early as 1928 (40) proposed that the minerals were secreted from cells. The present findings regarding the sequence of calcium accumulation in cells and matrix during heterotopic bone formation strongly suggest that osteoblasts and young osteocytes are active in the mineralization process. However, unfixed and unstained tissue, with its low contrast and resolution, made detection of other organelles than the nucleus difficult.

Other studies using ordinary electron microscopy or the K-pyroantimonate method have suggested that mitochondriae are involved in calcium accumulation (11,22,23). Electron-dense granules which have been found in mitochondria are interpreted as the site of calcium accumulation, but they could possibly also be the result of cell injury (12). How the calcium is transported out of the cells is poorly understood, although it has been proposed that calcium may be bound to organic components such as collagen precursors (4), phosphoproteins (39), glycoproteins (13) or lipids (41) and subsequently secreted into the intercellular space. Matrix vesicles which are also found in the mineralizing area of this model (32) may be involved in transport of minerals prior to crystal formation (26). The present results that show high calcium levels in the cytoplasm of the young matrix producing cells, favour such a transport function of the matrix vesicles rather than the theory that matrix vesicles are the site of calcium accumulation as proposed in cartilage (1).

The high Ca/P ratio observed in the osteoid and lower ratio, but higher total amounts of minerals in the electron-dense nodules, suggest that phosphatases could be involved in increasing the phosphorus level (24, 29). Formation of hydroxyapatite crystals which have a Ca/P ratio of approximately 2 may than be due to an alteration of the matrix into a calcifiable matrix (33, 38).

The present study showed that the initial events in mineralization are found intra-cellularly with an increased level of calcium. Later calcium could also be demonstrated in

the matrix of the newly formed osteoid. Then the total amount of calcium and especially phosphorus increased and electron-dense nodules appeared in the tissue, possibly reflecting extensive crystal formation. Although the cells showed distinct mineral accumulation it can not be excluded that calcium and phosphorus also may pass outside the cells to the sites of initial crystal formation. Such a possibility has been indicated in studies of dentin formation (27, 28). Based on the present results we conclude that young matrix-producing cells are greatly involved in mineral accumulation in the initial mineralization during heterotopic bone formation.

Legends to figures

Plate I

A. Semi-thin section of frozen, freeze-dried and Spurr® embedded material showing an osteoblast region (obl) and undifferentiated connective tissue lying outside the osteoblasts. Toluidine blue.

B. STEM image of an osteoblast in the region (obl) shown in A. Point of analysis in the nucleus (1) and the cytoplasm (2). Analysis of the mineralizing osteoid with electron-dense area (3) and the surrounding less electron-dense zone (4). Uncoated, 100 nm section-thickness. 15000 X

Energy spectra from spots corresponding to the numbers and arrows on B. Spot-size 125 nm.

B,1. X-ray spectrum showing well defined signals from the nucleus representing phosphorus (P), and only small amounts of calcium (Ca). Peaks for sodium (Na), silicium (Si) and chlorine (Cl) are due to radiation from external factors as for instance Si from the instrument and Na and Cl from the embedding medium.

B,2. X-ray spectrum from an analysis of the cytoplasm showing a distinct Kα peak for Ca. The counts for P is lower than in the nucleus.

B,3. X-ray spectrum from the electron-dense nodule. Note the reduced vertical scale (V.S) compared to B 1,2 and 3, and the lower Ca/P ratio (approx. 2) than in B,4.

B,4. X-ray spectrum form the less electron-dense area of the osteoid with lower $K\alpha$ peak for Ca and P, but higher Ca/P ratio (approx. 5) than in the electron-dense nodules (B,3). Note the reduced vertical scale compared to B,1, 2 and 3.

Plate II

A. STEM picture from an uncontrasted section of 100 nm showing two osteocytes which are nearly completely surrounded by fully mineralized bone. Nucleus (1) and cytoplasm (2). One part was not fully mineralized (3) resembling the zone around the electron-dense nodules in plate I:B, 4. Electron-dense matrix (4). 5300 X

Energy spectra from spots corresponding to the number and arrows in A. Spot-size 125 nm.

A,1. Analysis in the nucleus showing strong signals from P and weaker from Ca.

A,2. Analysis of the cytoplasm exhibiting more Ca and less P than in the nucleus.

A,3. Extra -cellular measurements in osteoid showing a high Ca content and high Ca/P ratio (approx.6).

A,4. Fully mineralized bone having a high Ca level and lower Ca/P ration (approx.2) than the osteoid (A,3). Note the reduced vertical scales in A,3 and A,4.

Plate III

A. The STEM image of an osteocyte in fully mineralized induced heterotopic bone. Section thickness 100 nm, uncoated. Frozen, freeze-dried and Spurr® embedded material. Uncontrasted 9500 X.

The energy spectra using 125 nm spot-size showed:

A,1. Analysis in the nucleus with a distinct $K\alpha$-peak for P but a low peak for Ca.

A,2. Analysis in the cytoplasm showed only small $K\alpha$-peak for C and P.

B X-ray spectrum from the nucleus of a cell in undifferentiated connective tissue. 125 nm spot-size.

C X-ray spectrum from the cytoplasm of the same cell as in B having no detectable Ca or P. 125 nm spot-size.

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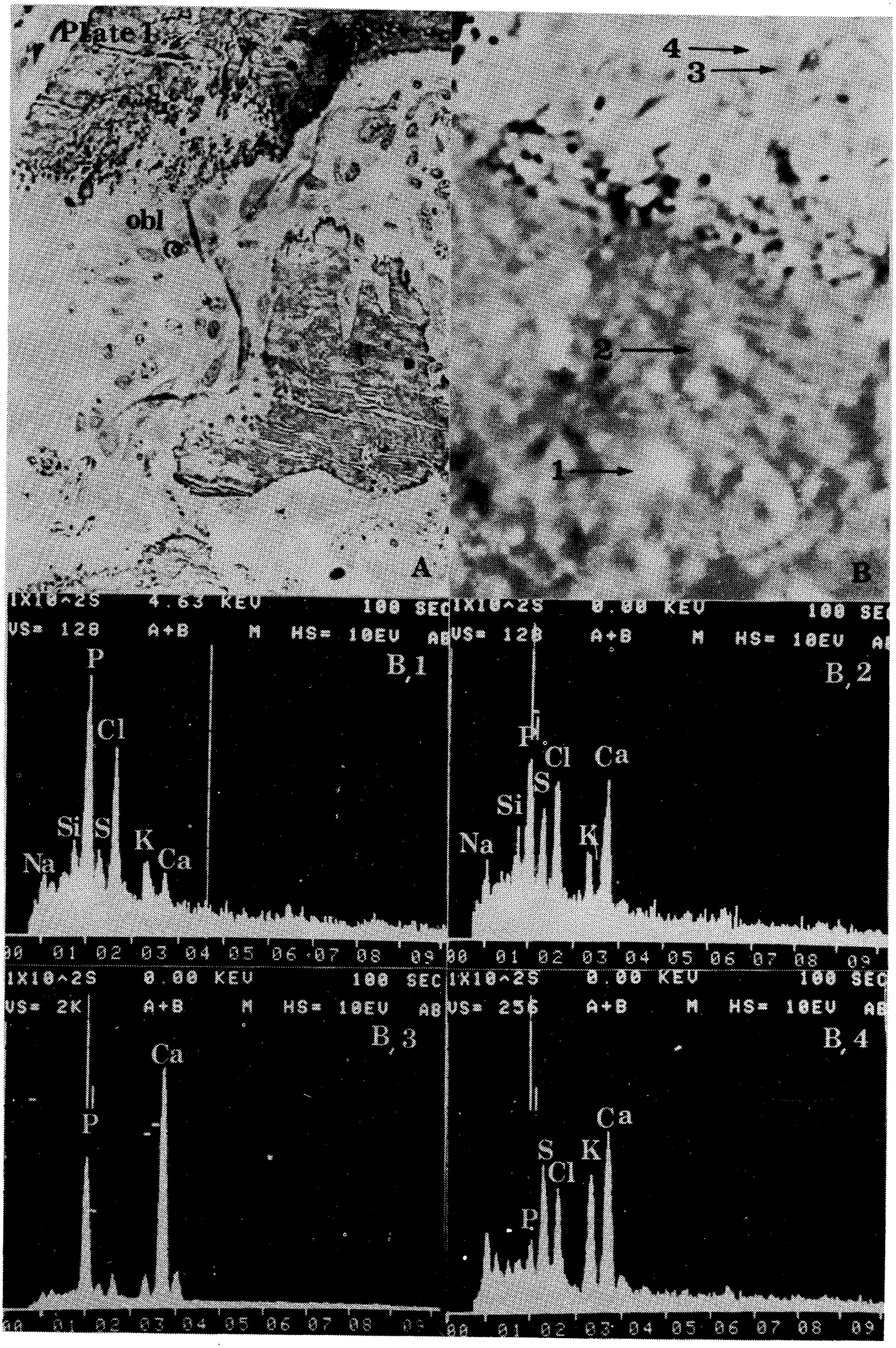
Plate 1
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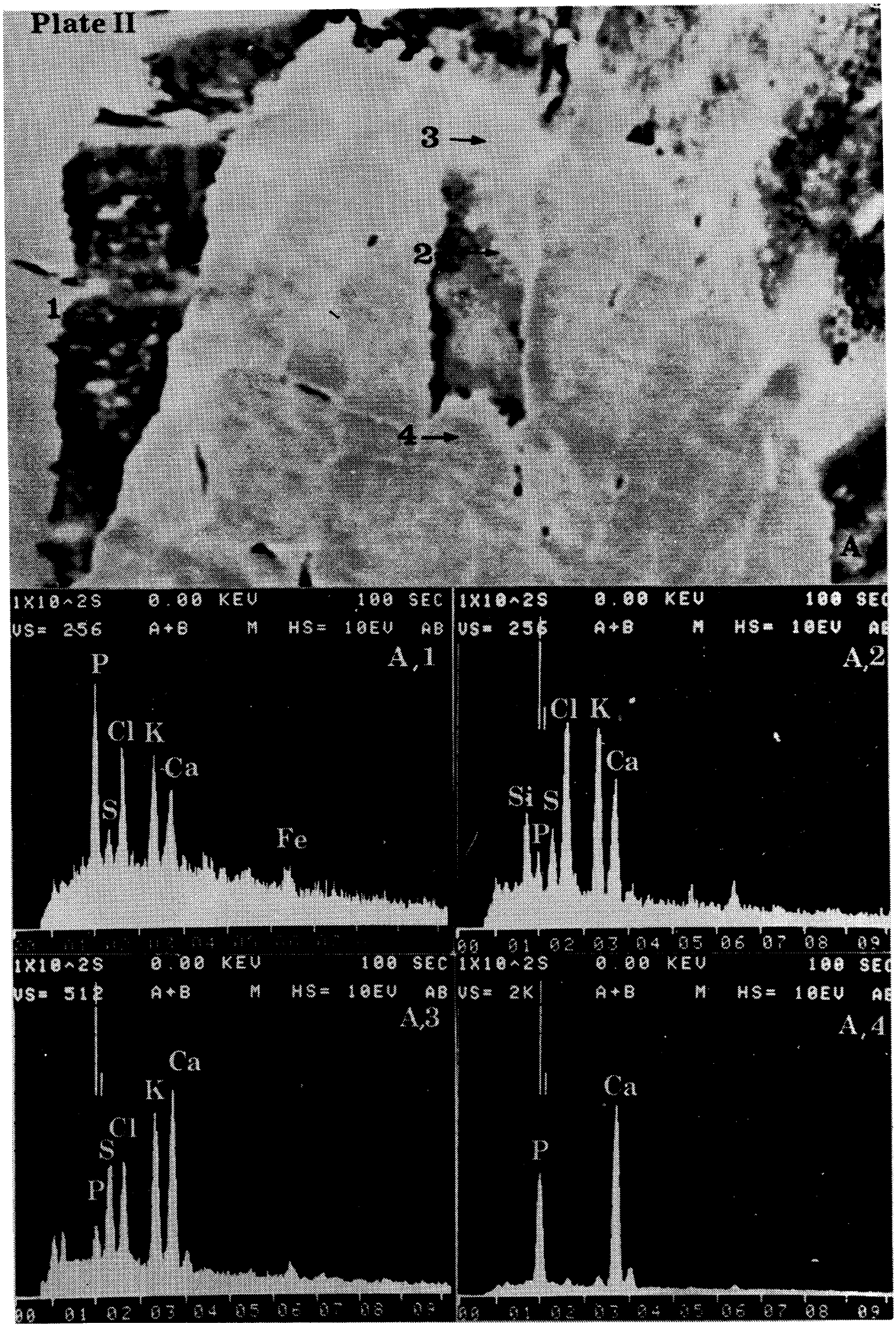
Plate II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A,2
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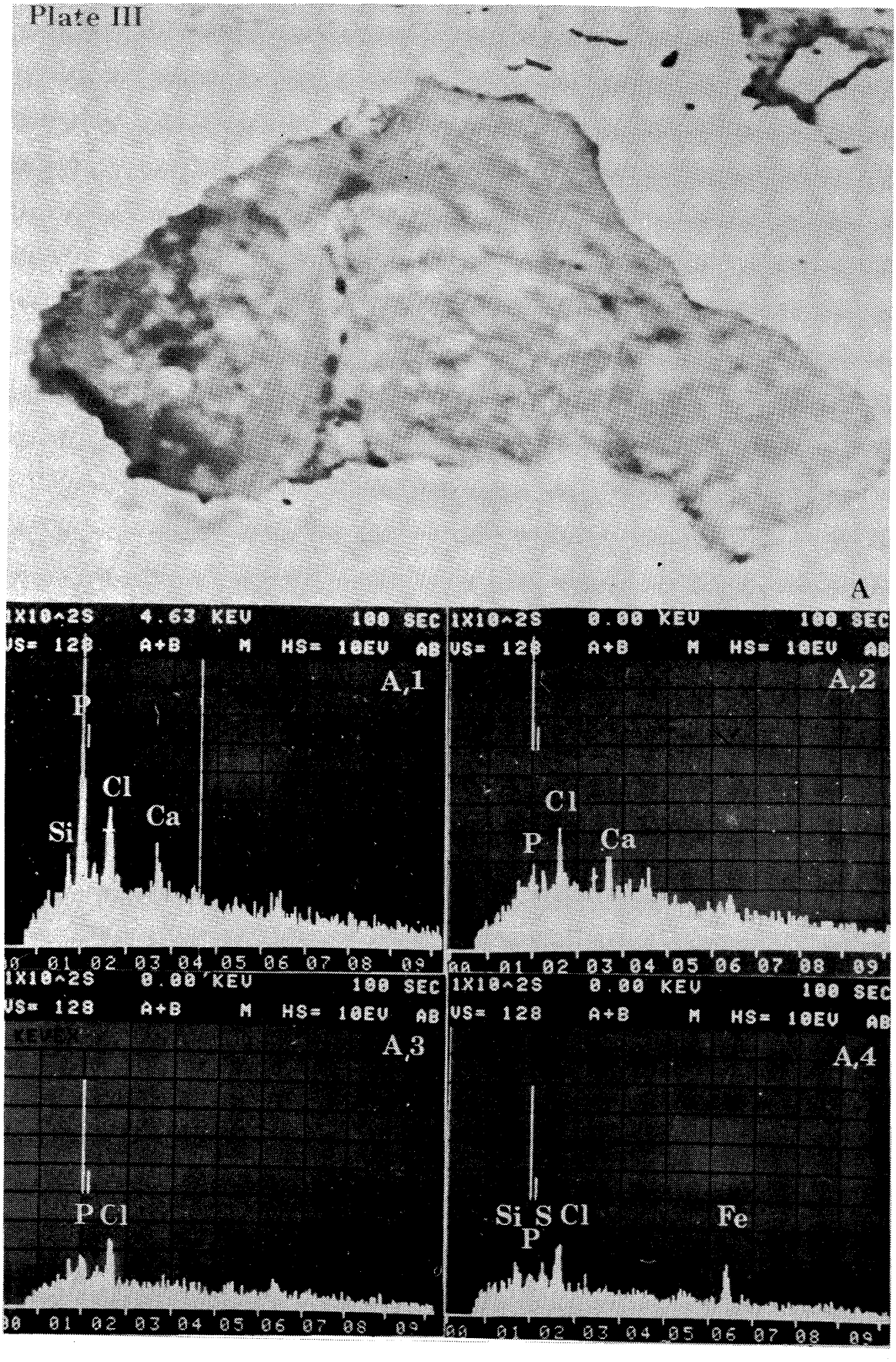
Plate III



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