

Research article

Open Access

The effect of low-level laser irradiation (In-Ga-Al-AsP - 660 nm) on melanoma *in vitro* and *in vivo*

Lúcio Frigo¹, Juliana SS Luppi¹, Giovani M Favero², Durnavei A Maria³, Sócrates C Penna⁴, Jan M Bjordal^{5,6}, Rene J Bensadoun⁷ and Rodrigo AB Lopes-Martins*⁴

Address: ¹Biological Sciences and Health Center, Cruzeiro do Sul University, Av. Dr. Ussiel Cirilo, 225 São Miguel Paulista, 08060-070 São Paulo, SP - Brasil, ²State University of Ponta Grossa, General Biology Department, Av. Gal. Carlos Cavalcanti, 4748 Ponta Grossa 84030-900, PR - Brasil, ³Laboratory of Biochemistry and Biophysics, Butantan Institute, Av. Dr. Vital Brasil, 1500 São Paulo 05599-000, SP - Brasil, ⁴Laboratory of Pharmacology and Phototherapy of Inflammation, Department of Pharmacology, Institute of Biomedical Sciences, University of São Paulo - São Paulo 05508-900 SP - Brasil, ⁵Institute for Physiotherapy, Bergen University College, Moellendalsvn. 6, 5009 Bergen, Norway, ⁶Section of Physiotherapy Science, Institute of Public Health and Primary Health Care, University of Bergen, Kalfarveien 31, 5018 Bergen, Norway and ⁷Service d'Oncologie Radiothérapique, CHU de Poitiers, 2, rue de la Milétrie, BP 577, 86021 - Poitiers Cedex, France

Email: Lúcio Frigo - luciofrigo@uol.com.br; Juliana SS Luppi - luciofrigo@uol.com.br; Giovani M Favero - gmfavero@yahoo.com.br; Durnavei A Maria - durvaneia@usp.br; Sócrates C Penna - scp@usp.br; Jan M Bjordal - jmb@hib.no; Rene J Bensadoun - rene-jean.bensadoun@nice.fnclcc.fr; Rodrigo AB Lopes-Martins* - rmartins@icb.usp.br

* Corresponding author

Published: 20 November 2009

Received: 19 December 2008

BMC Cancer 2009, 9:404 doi:10.1186/1471-2407-9-404

Accepted: 20 November 2009

This article is available from: <http://www.biomedcentral.com/1471-2407/9/404>

© 2009 Frigo et al; licensee BioMed Central Ltd.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/2.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Abstract

Background: It has been speculated that the biostimulatory effect of Low Level Laser Therapy could cause undesirable enhancement of tumor growth in neoplastic diseases. The aim of the present study is to analyze the behavior of melanoma cells (B16F10) *in vitro* and the *in vivo* development of melanoma in mice after laser irradiation.

Methods: We performed a controlled *in vitro* study on B16F10 melanoma cells to investigate cell viability and cell cycle changes by the Tripian Blue, MTT and cell quest histogram tests at 24, 48 and 72 h post irradiation. The *in vivo* mouse model (male Balb C, n = 21) of melanoma was used to analyze tumor volume and histological characteristics. Laser irradiation was performed three times (once a day for three consecutive days) with a 660 nm 50 mW CW laser, beam spot size 2 mm², irradiance 2.5 W/cm² and irradiation times of 60s (dose 150 J/cm²) and 420s (dose 1050 J/cm²) respectively.

Results: There were no statistically significant differences between the *in vitro* groups, except for an increase in the hypodiploid melanoma cells (8.48 ± 1.40% and 4.26 ± 0.60%) at 72 h post-irradiation. This cancer-protective effect was not reproduced in the *in vivo* experiment where outcome measures for the 150 J/cm² dose group were not significantly different from controls. For the 1050 J/cm² dose group, there were significant increases in tumor volume, blood vessels and cell abnormalities compared to the other groups.

Conclusion: LLLT Irradiation should be avoided over melanomas as the combination of high irradiance (2.5 W/cm²) and high dose (1050 J/cm²) significantly increases melanoma tumor growth *in vivo*.

Background

Malignant melanoma represents a burden to modern society and requires considerable efforts in terms of health service utilization. The incidence is increasing worldwide and in the Netherlands the prevalence is currently 16.1/100,000 with a mortality rate of 3.0/100,000[1].

Low level laser therapy (LLLT) has gained increasing popularity as a treatment for soft tissue injuries and joint conditions. It is applied transcutaneously with typical irradiances being 10 mW/cm² - 5,000 mW/cm², treatments times being in the range of 10 seconds - 2 minutes, with total energy delivered of 1 - 4 Joules(J)/cm² per point when targeting joints, tendons and muscles. The cellular proliferative potential of LLLT irradiation has attracted some negative speculation that this could also increase tumor growth in neoplastic diseases. Previous studies of LLLT irradiation of tumor cells *in vitro* have generated conflicting research data across a range of cultivated tumor cell lines and irradiation parameters [2-11] but there have been relatively few *in vivo* studies published [12,13]. *In vivo* studies are essential for the study of disease development and should be the main tool for studying the behavior of tumor cells. The complexity of the multicellular environment in an ongoing disease makes it hard to predict tumor behavior and cell culture studies alone are inadequate to for assessment of tumor responses.

Increases in cell proliferation and collagen biosynthesis after LLLT in wound healing improvement has already been observed in the pioneer work of Mester et al. [14]. The following decades were marked by a large quantity of research articles in LLLT. A better understanding of laser light modulatory mechanisms was obtained, but this effort also yielded conflicting results. There is a shortage of evidence about the effects of LLLT in malignant conditions such as melanoma. The complete biochemical mechanisms of cell proliferation after LLLT irradiation are still uncertain and we believe there is a need to study the effects of LLLT on tumor growth in suitable cell and animal models.

The aim of the present work is to study the effect of LLLT irradiation both *in vitro* and *in vivo*. For this purpose we decided to study cell viability and cell cycle changes in melanoma cells (B16F10) *in vitro*, and their behavior when injected subcutaneously into Balb C mice *in vivo*.

Methods

All the experimental procedures were submitted to and approved by the Ethical Committee at the Cruzeiro do Sul University.

Cell culture

B16F10 murine melanoma cells were obtained from ATCC (clone CRL 6457). Melanoma cells were grown in

RPMI 1640 medium supplemented with 10% fetal bovine serum (FBS), 100 U/mL penicillin/streptomycin and 24 mM NaHCO₃ at 37°C in a humidified atmosphere containing 5% CO₂. Cells were seeded at an initial density of 2 × 10⁴ cells/cm² (B16F10) for cell viability, which was determined by the MTT method and 1 × 10⁶ cells/cm² for the Trypan blue exclusion test.

In vitro laser irradiation

B16F10 cells were irradiated a total of three times (once a day for three consecutive days) in a 96 well culture plate for the MTT method and in a 12 well plate for Trypan blue and cytometric assays. Irradiation was performed with a 660 nm, 50 mW Continuous Wave (CW) laser, beam spot size 2 mm², irradiance 2.5 W/cm² (Quasar Medical - Dentoflex, São Paulo, Brasil). The seeded wells were spaced 5 cm apart in all directions and a thin aluminum sheet was placed halfway (2.5 cm) between them to prevent unintentional light scattering between the wells. The wells were randomly divided into a control group which received no irradiation, and a treatment group which received an LLLT dose of 150 J/cm² with an irradiance of 2.5 W/cm² for 60 seconds (3J), while a second group received sessions with an LLLT dose of 1050 J/cm² with an irradiance of 2.5 W/cm² for 420 seconds (21J). Total energy delivered after all three sessions was 9J and 63J respectively in the irradiated groups. A support device held the LLLT emission tip perpendicular to and 2 mm distant from the culture media. Irradiation was carefully timed and carried out in a dark laminar flux hood.

Animals

The animals were isogenic male Balb C mice (n = 21), which were randomized into one of three groups; a control group (n = 7), a "low" dose group (n = 7) and a "high" dose group (n = 7). The mice were injected subcutaneously with a suspension of 2 × 10⁶ B16F10 melanoma cells.

In vivo laser irradiation

After fifteen days of tumor growth the animals were irradiated three times (once a day for three consecutive days) at the site of the injected melanoma cells with the same laser and laser parameters as used in the *in vitro* study. Irradiation was performed with a 660 nm 660 nm, 50 mW Continuous Wave (CW) laser, beam spot size 2 mm², irradiance 2.5 W/cm² (Quasar Medical - Dentoflex, São Paulo, Brasil). Control Group: Received no irradiation Group 1: Received three LLLT sessions (once a day for three consecutive days) each of 60 seconds with a dose of 150 J/cm², (energy delivered per session was 3J, total energy delivered after three sessions was 9J) Group 2: Received three sessions (once a day for three consecutive days) each of 420 seconds with a dose of 1050 J/cm², (energy delivered per session was 21J, and total energy delivered after three sessions was 63J).

Outcome measures in vitro

Cell viability and cell changes were determined by MTT method and Trypan blue exclusion tests (B16F10). Cells were seeded at a density of 1×10^6 cells/Cm² (B16F10). At the end of the experiment, cells were treated with trypsin (0.05% trypsin in 0.02% EDTA) and washed 3 times with PBS, fixed in 70% ethanol, and stained with propidium iodide (PI) 50 mg/10 uL final concentration, these can distinguish hypodiploid (non-viable or dead cells) from diploid (viable) cells, for 30 min in the dark. All analyses were done using a FACScalibur flow cytometer (Becton Dickinson, San Jose, CA). The red fluorescence of PI was collected through a 585/42-nm band-pass filter, and the fluorescence signals were measured in a linear scale of 1024 channels. For each sample, at least 10,000 events were acquired and the data were analyzed using appropriate software (CELLQuest, Becton Dickinson, San Jose, CA). Cell viability was assessed by counting adherent and non-adherent cells and measured by the cellular permeability to propidium iodide. Cells in S/G₂/M (proliferating) and G₀/G₁ phases, and hypodiploid cells (cells under death process) were analyzed.

Outcome measures in vivo

Tumor cell growth area was estimated measuring length and width with a paquimeter device and using the formula: volume = length \times width² π div 6. Histological tumor analysis was performed after tumor volume measurements. Animals were anaesthetized with inhaled halothane and sacrificed by cervical dislocation. Tumor mass was immediately removed and immersed in a 4% phosphate buffered paraformaldehyde solution for 48 h. Specimens were dehydrated and embedded in paraffin prior to the 5 μ m microtome sections. Histological sections were collected on glass slides and hematoxylin-eosin stained. Analysis and photographs were carried out in a Nikon-YS100 photomicroscope.

Statistical analysis

The obtained data were first plotted for analysis of normal distribution, and statistical analysis was then performed with parametric tests if the data were normally distributed. The statistical level of significance was set at $P < 0.05$, and significance was tested statistically by an ANOVA-test. The mean values and its standard error (SE) were calculated, and differences between control group data and the irradiated group data were tested statistically with Bonferoni's test.

Results

In vitro experiments

The Trypan Blue dye exclusion test showed no statistical differences in proliferation or cell death numbers among irradiated groups and control group in the different times analyzed (Figure 1).

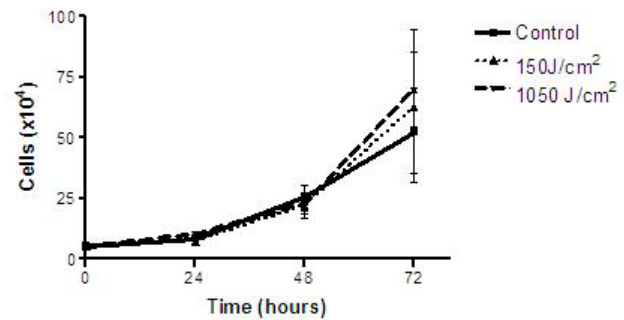


Figure 1
Total B16F10 cell count number by Trypan blue exclusion method. No significant differences were observed.

The MTT colorimetric test showed no statistical differences in proliferation or cell death numbers among irradiated groups and control group in the different times analyzed (Figure 2).

Cell cycle analysis in B16F10 cells showed no statistically significant differences in the cell numbers in G₀/G₁, S, G₂/M phases at 24 h, 48 h and 72 h among irradiated groups and control group (Figure 3, 4, 5).

There was statistically a significant difference ($p < 0.05$) in hypodiploid cells (possible cell death) at 72 h between the irradiated and control groups ($8.48 \pm 1.40\%$ and $4.26 \pm 0.60\%$). The increase in apoptosis was most prominent in the low dose 150 J/cm² group (Figure 6).

In vivo experiments

15 days after the B16F10 cell injections all the animals presented average tumor mass volume of 0.12 ± 0.04 cm³. The increase of the tumor mass volume of control and irradiated groups are shown in Figure 7.

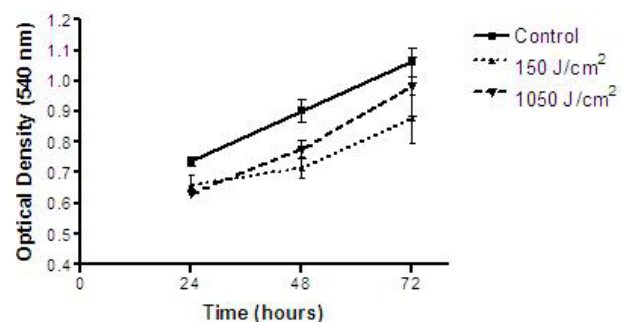


Figure 2
Proliferative-inhibitory effects of low-level laser irradiation in B16F10 cells evaluated by MTT colorimetric method. * = $p < 0.05$.

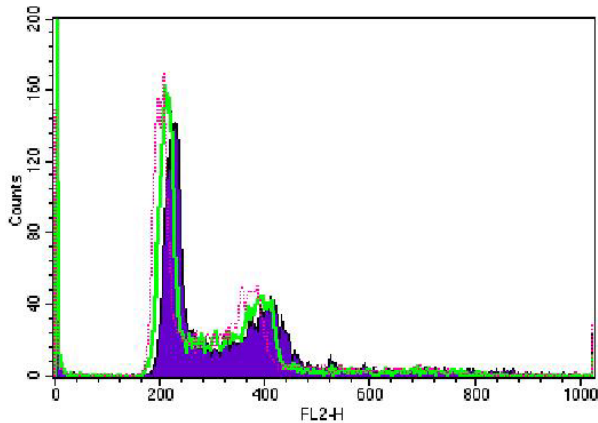


Figure 3
Cell-Quest acquisition histogram of B16F10 cell cycle phases after 24 h irradiation. Red dot refers to control group; green line refers to 150 J/cm² group and purple curve refers to 1050 J/cm² group.

At the 10th day, the tumor mass volume was significantly higher in the 1050 J/cm² group when compared to the 150 J/cm² and the control group. No significant difference in tumor volume was observed between the 150 J/cm² and the control group (Figure 8).

The macroscopic appearance of dissected tumor differed between the 1050 J/cm² group and the two other groups. In addition to a marked increase of the volume of this group, the connective tissue of the capsule appeared sticky to the tumor mass and to the adjacent muscle tissue. A

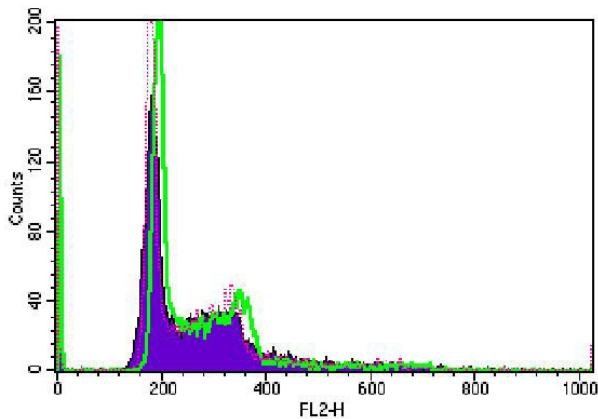


Figure 4
Cell-Quest acquisition histogram of B16F10 cell cycle phases after 48 h irradiation. Red dot refers to control group; green line refers to 150 J/cm² group and purple curve refers to 1050 J/cm² group.

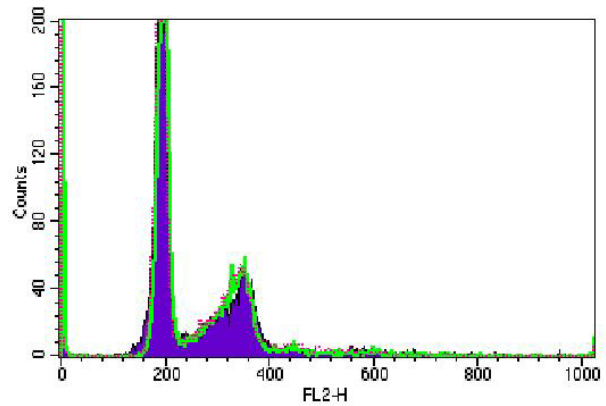


Figure 5
Cell-Quest acquisition histogram of B16F10 cell cycle phases after 72 h irradiation. Red dot refers to control group; green line refers to 150 J/cm² group and purple curve refers to 1050 J/cm² group.

greater number of blood vessels were also observed (Figure 9)

Histological sections of the control group revealed a dense mass of melanin producing melanoma cells invaded by lymphocytes, plasma cells and macrophages. A rich vascular bed filled with leukocytes and red blood cells can be observed. Some restricted areas of necrotic tissue were also present. In the connective tissue of the capsule, immunological cells spread through thin collagen fibers and edema areas (Figure 10).

In the histological sections of the 150 J/cm² group, immune cells were less frequent in the tumor mass, and large blood vessels were filled with leukocytes and red blood cells. Necrotic areas were slightly larger compared

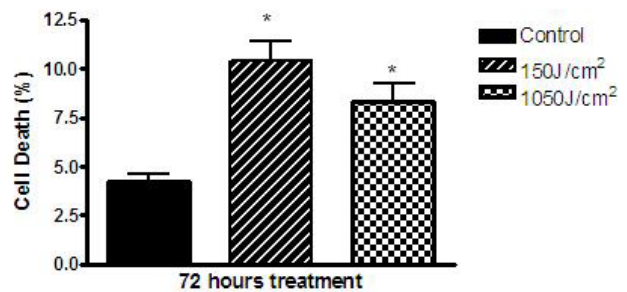


Figure 6
Graphic showing B16F10 cell death percentage after 72 h of laser irradiation. Statistically significant difference is indicated by asterisk (*P < 0,05).

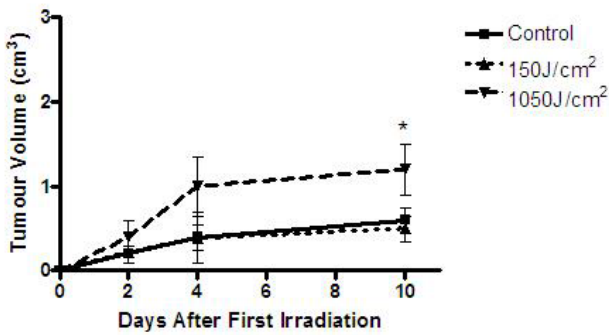


Figure 7
Graphic showing melanoma tumor growth in the three groups until the 10th day after irradiation.

to the control group. The connective tissue of the capsule had fewer immune cells in a greater area of thin fibers of collagen (Figure 11).

Histological sections of the 1050 J/cm² group showed remarkably atypical melanoma cells. Nuclei were of various sizes and shapes, and apoptotic figures and the frequency of mitotic cells were high. Necrotic areas were more common and extensive compared to the other groups. Immune cells were observed in greater numbers in the tumor mass and in the highly vascular capsule (Figure 12).

Discussion

In the present paper we have investigated the effects of LLLT on malignant melanoma, *in vitro* and *in vivo*. The question of a potential unwanted proliferative effect of low-level laser irradiation, has been raised by some authors [15,16]. We observed that laser irradiation with a

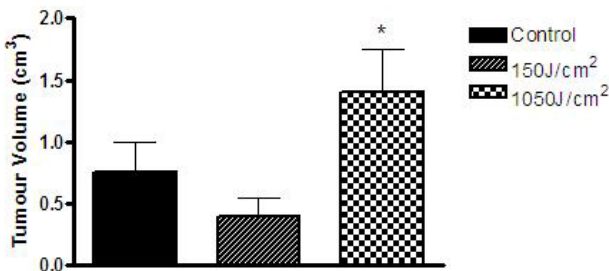


Figure 8
Graphic showing the percentage of volume growth on the 10th day after irradiation of the three groups. * = p < 0.05 when compared to the control group indicated by One Way ANOVA and Bonferroni's Multiple Comparison Test (*P < 0,05).

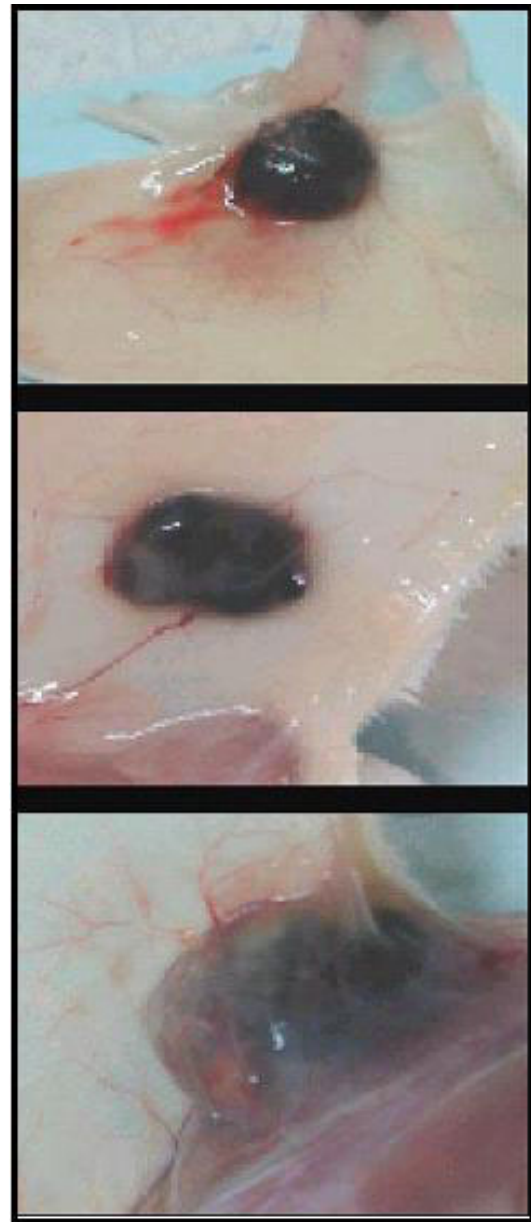


Figure 9
photography showing the macroscopic appearance of the tumor. A - control group, B - 150 J/Cm² group and C - 1050 J/Cm² group.

low LLLT dose of 150 J/cm² presented opposite effects when applied to each distinct situation. In the cultured melanoma cells, we found that the two LLLT doses presented a non-significant effect on tumor cells or even an inhibitory effect of cancer cell proliferation through increased apoptosis. In the *in vivo* experiment the low dose (150 J/cm²) was not inducing any changes in the cancer cell behavior. However, the high dose (1050 J/cm²) showed a significant increase in tumor mass volume and

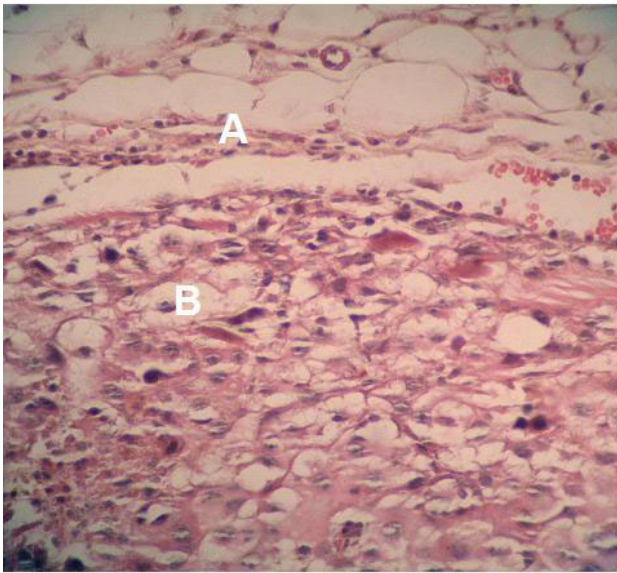


Figure 10
Hematoxylin-eosin stained photomicrograph showing tumor cells (A), connective tissue tumor capsule (B) Tumor mass in the control group. $\times 400$.

considerable histological alterations which indicate a worsening of the cancer. The results have several implications for research and clinical practice.

Cell culture is an important method for studying basic biological processes and to understand the possible cell reactions to treatments. Many kinds of tumor cell lines have been studied, ranging from carcinomas to sarcomas and myelomas [3,4,6,8,10,17]. We chose B16F10 melanoma cell line because it's a pigmented, highly aggressive and invasive tumor [18]. Our results of non-significant LLLT effects in the *in vitro* tests of cell viability are in accordance with the largely non-significant findings of other authors [9].

Our cell cycle analysis with flow cytometry method indicated a significant increase in cell death in 72 h of the 1050 J/cm² group. Some authors have previously found increased cell death *in vitro* after LLLT irradiation. LLLT fluences higher than 6 J/cm² seemed to increase cell death in melanoma cell lines (G361, LD50 and SKmel-23), and especially in melanin producing cells [19]. There seems to be an inverse relationship between laser fluence and melanoma cell growth in culture [11]. Other authors have reported an increase in G₀/G₁ phase of the cell cycle using HTB66 melanoma cell line [2], but our results did not support this finding. One important aspect of our findings is the discrepancy between the *in vitro* and *in vivo* experiments. It seems necessary to be careful in generalizing *in vitro* results, as cell-matrix interactions and cell behavior

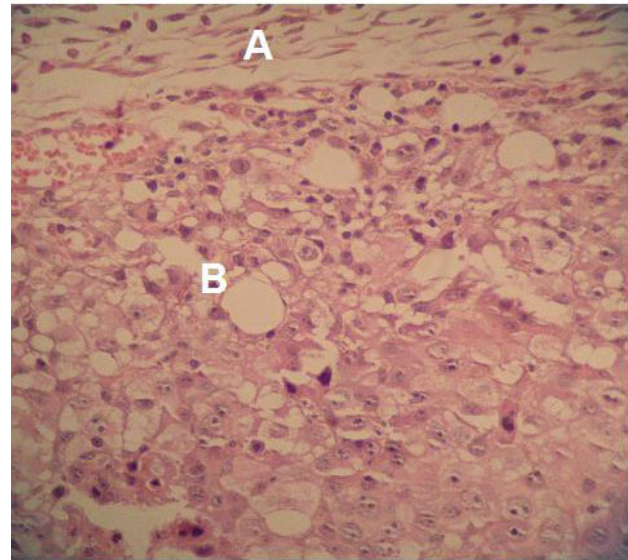


Figure 11
Hematoxylin-eosin stained photomicrograph showing (A), connective tissue tumor capsule (B) tumor mass in the 150 J/Cm². $\times 400$.

in the complex environment of tissues may produce unexpected reactions.

Our results demonstrated a significant tumor growth when the animals were irradiated with the high dose of

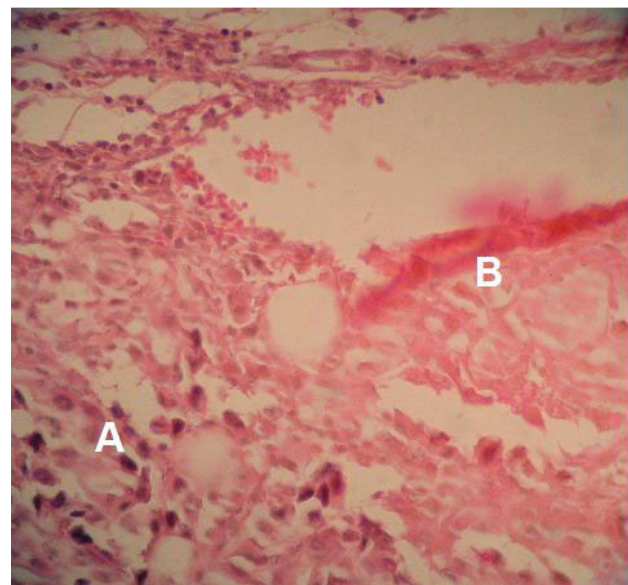


Figure 12
Hematoxylin-eosin stained photomicrograph showing tumor mass (A), necrotic area (B) in the 1050 J/Cm². $\times 400$.

1050 J/cm². This finding is in line with observations of enhanced Ehrlich ascites tumor growth after laser irradiation which have been reported in an early paper on LLLT [12]. However it seems that typical LLLT doses ranging from 1 - 4 Joules have no influence on tumor growth, or rather they can inhibit it in implanted glioma in mice [13].

Histological data also revealed that important differences in cell morphology were induced by high doses of laser irradiation. The immune cells (lymphocytes, plasma cells and macrophages) increased in the group irradiated with the high dose of 1050 J/cm². This group also presented significant areas of necrosis, a high number of atypical cells and an increase in the number of blood vessels.

Zhu et al. [8] reported differences in Focal Adhesion Kinases (FAK) and van Leeuwen et al. [7], showed differences in α -1 and β -4 subunits of integrin molecule. Both factors are important in tumor genesis and metastasis.

Many factors may contribute to tumor growth and most of them can be modulated by laser irradiation, for instance: low-level laser can enhance angiogenesis [20-22], growth factor synthesis [23-25], inflammatory metabolites [26] as well as modulate immunological cells and inflammation [27-29].

Conclusion

LLLT administered by a dose of 150 J/cm² appears safe with only minor effects on B16F10 melanoma cells proliferation *in vitro* and no significant effect on tumor growth *in vivo*. However, a high irradiance (2.5 W/cm²) combined with high dose of 1050 J/cm², can stimulate melanoma tumor growth with distinct histological features *in vivo*. Further studies are necessary to elucidate the main factors that are responsible for the different behaviors on tumor cells in response to laser light, and to determine laser irradiance and energy thresholds for stimulation of abnormal melanoma cell behavior.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

LF carried out melanoma injections in mice and histological analysis, JSSL and DAM carried out melanoma cell culture Trypan Blue dye exclusion test and MTT colorimetric test, GMF carried out of cell cycle analysis by flow cytometry and statistics, SCP and JMB were involved in drafting the manuscript and analysis, RABLM and RJB were involved in revising it critically and gave the final approval of the version to be published.

All authors have read and approved the final manuscript.

Acknowledgements

This study was supported by Fundação de Amparo à Pesquisa do Estado de São Paulo (FAPESP) 07/59124-0. and Cruzeiro do Sul University.

References

1. Hoekstra HJ: **The European approach to in-transit melanoma lesions.** *Int J: Hyperthermia* 2008, **24(3)**:227-237.
2. Chan HHL, Xiang L, Leung JCK, Tsang KWT, Lai K: **In vitro study examining the effect of sub-lethal QS 755 nm lasers on the expression of p16INK4a on melanoma cell lines.** *Lasers in Surgery and Medicine* 2003, **32**:88-93.
3. Kujawa J, Zavadnik IB, Lapshina A, Labieniec M, Bryszewska M: **Cell survival, DNA, and protein damage in B14 cells under low-intensity near-infrared (810 nm) laser irradiation.** *Photomedicine and Laser Surgery* 2004, **22(6)**:504-508.
4. Mognato M, Squizzato F, Facchin F, Zaghetto L, Corti L: **Cell growth modulation of human cells irradiated in vitro low-level laser therapy.** *Photomedicine and Laser Surgery* 2004, **22(6)**:523-526.
5. de Castro JL, Pinheiro AL, Werneck CE, Soares CP: **The effect of laser therapy on the proliferation of oral KB carcinoma cells: an in vitro study.** *Photomed Laser Surg* 2005, **23(6)**:586-9.
6. Sroka R, Schaffer M, Fuchs C, Pongratz T, Schrader-Reichard U, Busch M, Schaffer PM, Duhmke E, Baumgartner R: **Effects on the mitosis of normal and tumor cells induced by light treatment of different wavelengths.** *Lasers Surg Med* 1999, **25(3)**:263-71.
7. van Leeuwen RL, Dekker SK, Byers HR, Vermeer BJ, Grevelink JM: **Modulation of alpha 4 beta 1 and alpha 5 beta 1 integrin expression: heterogeneous effects of Q-switched ruby, Nd:YAG, and alexandrite lasers on melanoma cells in vitro.** *Lasers Surg Med* 1996, **18(1)**:63-71.
8. Zhu NW, Perks CM, Burd AR, Holly JM: **Changes in the levels of integrin and focal adhesion kinase (FAK) in human melanoma cells following 532 nm laser treatment.** *Int J Cancer* 1999, **82(3)**:353-8.
9. Marchesini R, Dasdia T, Melloni E, Rocca E: **Effect of low-energy laser irradiation on colony formation capability in different human tumor cells in vitro.** *Lasers Surg Med* 1989, **9(1)**:59-62.
10. Ocanã-Quero JM, Perez de la Lastra J, Gomez-Villamandos R, Moreno-Millan M: **Biological effect of helium-Neon (He-Ne) laser irradiation on mouse myeloma (Sp2-Ag14) cell line in vitro.** *Lasers Med Sci* 1998, **13**:214-18.
11. Jamieson CW, Litwin MS, Longo SE, Krementz ET: **Enhancement of melanoma cell culture growth rate by ruby laser radiation.** *Life Sci* 1969, **8(2)**:101-6.
12. Mester E, Lapis K, Tota JG: **Ultrastructural changes in Ehrlich ascites tumor cells following laser irradiation.** *Arch Geschwulstforsch* 1971, **38(3)**:210-20.
13. Abe M, Fujisawa K, Suzuki H, Sugimoto T, Kanno T: **Role of 830 nm low reactive level laser on the growth of an implanted glioma in mice.** *Keio J Med* 1993, **42(4)**:177-9.
14. Mester E: **The use of the laser beam in therapy.** *Orv Hetil* 1966, **107(22)**:1012-6.
15. Karu TI: **Effects of visible radiation on cultured cells.** *Photochem Photobiol* 1998, **52**:1089-1990.
16. Moore P, Ridgway TD, Higbee RG, Howard EW, Lucroy MD: **Effect of wavelength on low-intensity laser irradiation-stimulated cell proliferation in vitro.** *Lasers in Surgery and Medicine* 2005, **36**:8-12.
17. Morales JA, Ruiz-Gómez MJ, Gil-Carmona L, Souvirón A, Martínez-Morillo M: **He-Ne laser has no effect on cell cycle phases of human colon adenocarcinoma cells.** *Rev Esp Fisiol* 1995, **51(1)**:43-7.
18. Gray-Schopfer V, Wellbrock C, Marais R: **Melanoma biology and new targeted therapy.** *Nature* 2007, **445(7130)**:851-7.
19. Zhu NW, Kenealy J, Burd A, Graddidge T, Warr R, Rigby HS, Kemshead JT: **Sub-lethal effects of exposing the human melanoma cell line SKmel-23 to 532 nm laser light.** *Int J Cancer* 1997, **72(6)**:1104-12.
20. Schindl A, Merwald H, Schindl L, Kaun C, Wojta J: **Direct stimulatory effect of low-intensity 670 nm laser irradiation on human endothelial cell proliferation.** *British Journal of Dermatology* 2003, **148**:334-336.
21. Salate AC, Barbosa G, Gaspar P, Koeke PU, Parizotto NA, Benze BG, Foschiani D: **Effect of In-Ga-Al-P diode laser irradiation on**

- angiogenesis in partial ruptures of Achilles tendon in rats.** *Photomed Laser Surg* 2005, **23(5)**:470-5.
22. Ihsan FR: **Low-level laser therapy accelerates collateral circulation and enhances microcirculation.** *Photomed Laser Surg* 2005, **23(3)**:289-94.
 23. Yu W, Naim JO, Lanzafame RJ: **The effect of laser irradiation on the release of bFGF from 3T3 fibroblasts.** *Photochem Photobiol* 1994, **59(2)**:167-70.
 24. Zhang W, Wu C, Pan W, Tian L, Xia J: **Low-power Helium-Neon laser irradiation enhances the expression of VEGF in murine myocardium.** *Chin Med J* 2004, **117(10)**:1476-1480.
 25. Agaiby AD, Ghali LR, Wilson R, Dyson M: **Laser modulation of angiogenic factor production by T-lymphocytes.** *Lasers Surg Med* 2000, **26(4)**:357-63.
 26. Yu HS, Chang KL, Yu CL, Chen JW, Chen GS: **Low-energy helium-neon laser irradiation stimulates interleukin-1 alpha and interleukin-8 release from cultured human keratinocytes.** *J Invest Dermatol* 1996, **107(4)**:593-6.
 27. Dube A, Bansal H, Gupta PK: **Modulation of macrophage structure and function by low level He-Ne laser irradiation.** *Photochem Photobiol Sci* 2003, **2(8)**:851-5.
 28. Fujimaki Y, Shimoyama T, Liu Q, Umeda T, Nakaji S, Sugawara K: **Low-level laser irradiation attenuates production of reactive oxygen species by human neutrophils.** *J Clin Laser Med Surg* 2003, **21(3)**:165-70.
 29. Honmura A, Yanase M, Obata J, Haruki E: **Therapeutic effect of Ga-Al-As diode laser irradiation on experimentally induced inflammation in rats.** *Lasers Surg Med* 1992, **12(4)**:441-9.

Pre-publication history

The pre-publication history for this paper can be accessed here:

<http://www.biomedcentral.com/1471-2407/9/404/prepub>

Publish with **BioMed Central** and every scientist can read your work free of charge

"BioMed Central will be the most significant development for disseminating the results of biomedical research in our lifetime."

Sir Paul Nurse, Cancer Research UK

Your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours — you keep the copyright

Submit your manuscript here:
http://www.biomedcentral.com/info/publishing_adv.asp

